Rat Kidney Toxicity Panel 2

96 Well Plate Assay

Cat. # RKTX2-37K

MILLIPLEX® MAP

RAT KIDNEY TOXICITY PANEL 2 KIT
96 Well Plate Assay

RKTX2-37K

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FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.

INTRODUCTION

Absorption, distribution, metabolism and excretion (ADME) parameters are critical to all stages of a fully integrated drug development program and are used to augment the interpretation of toxicological findings. As toxicity has been found to be the leading cause of drug failure, this area of research is expanding in search of more sensitive, rapid methods for determining organ-specific damage as quickly as possible. Drug induced damage to kidney cells, also known as renal toxicity, results from drug excretion. The traditional methodology for determining renal toxicity has been to measure the blood urea nitrogen (BUN) and serum creatinine. These two tests only detect kidney damage a week after it begins to occur and only show that damage has occurred somewhere in the kidney. With Millipore's MILLIPLEX™ Rat Kidney Toxicity Panels coupled with the Luminex® xMAP® platform, you receive the advantage of speed and consistency, allowing quantitative multiplex detection of multiple analytes simultaneously, dramatically improving productivity.

Millipore's MILLIPLEX Rat Kidney Toxicity Panel 2 is the most versatile system available for nephrotoxicity research.

- MILLIPLEX MAP offers you the ability to:
    - Choose any combination of analytes from our panel of 3 analytes to design a custom kit that better meets your needs.
- A convenient "all-in-one" box format gives you the assurance that you will have all the necessary reagents you need to run your assay.

Millipore's MILLIPLEX Rat Kidney Toxicity Panel 2 kit is to be used for the simultaneous quantification of the following 3 rat kidney toxicity biomarkers in any combination: Albumin, β-2-Microglobulin and Cystatin C in 1:500 diluted rat urine samples. While β-2-Microglobulin and Cystatin C are quantified with typical sandwich assays, the Albumin assay uses a competitive format.

This kit may be used for the analysis of all or any combination of the above rat kidney toxicity biomarkers in urine samples.

This kit is for research purposes only.

Please read entire protocol before use.

It is important to use same assay incubation conditions throughout your study.

PRINCIPLE

MILLIPLEX® MAP is based on the Luminex® xMAP® technology — one of the fastest growing and most respected multiplex technologies offering applications throughout the life sciences and capable of performing a variety of bioassays including immunoassays on the surface of fluorescent-coded beads known as microspheres.

- Luminex uses proprietary techniques to internally color-code microspheres with two fluorescent dyes. Through precise concentrations of these dyes, 100 distinctly colored bead sets can be created, each of which is coated with a specific capture antibody.
- After an analyte from a test sample is captured by the bead, a biotinylated detection antibody is introduced.
- The reaction mixture is then incubated with Streptavidin-Phycoerythrin (SA-PE) conjugate, the reporter molecule, to complete the reaction on the surface of each microsphere.
- The microspheres are allowed to pass rapidly through a laser that excites the internal dyes marking the microsphere set. A second laser excites PE, the fluorescent dye on the reporter molecule.
- Finally, high-speed digital-signal processors identify each individual microsphere and quantify the result of its bioassay based on fluorescent reporter signals.

The capability of adding multiple conjugated beads to each sample results in the ability to obtain multiple results from each sample. Open-architecture xMAP® technology enables multiplexing of many types of bioassays reducing time, labor and costs over traditional methods.

STORAGE CONDITIONS UPON RECEIPT

- Recommended storage for kit components is 2 - 8°C.
- Once the standards and controls have been reconstituted, immediately transfer contents into polypropylene vials. **DO NOT STORE RECONSTITUTED STANDARDS OR CONTROLS IN GLASS VIALS**. For long-term storage, freeze reconstituted standards and controls at ≤ -20°C. Avoid multiple (>2) freeze/thaw cycles.
- **DO NOT FREEZE Antibody-Immobilized Beads, Detection Antibodies, and Streptavidin-Phycoerythrin**.

REAGENTS SUPPLIED

Note: Store all reagents at 2 – 8°C

REAGENTS SUPPLIED	CATALOG NUMBER	VOLUME	QUANTITY
Rat Kidney Toxicity Panel 2 Standard	RKTX-8037-2	lyophilized	1 vial
Rat Kidney Toxicity Panel 2 Quality Controls 1 and 2	RKTX-6037-2	lyophilized	2 vials
Rat Kidney Toxicity Panel 2 Biotinylated Albumin (only supplied if Albumin is a requested analyte)	RKTX-BTAB	3.2 mL	1 bottle
Set of one 96-Well Microtiter Filter Plate with 2 Sealers	MX-PLATE	----------	1 plate 2 sealers
Assay Buffer Note: Contains 0.08% Sodium Azide	L-AB1	30 mL	2 bottles
10X Wash Buffer Note: Contains 0.05% Proclin	L-WB	30 mL	1 bottle
Rat Kidney Toxicity Panel 2 Detection Antibodies	RKTX-1037-2	5.5 mL	1 bottle
Streptavidin-Phycoerythrin	L-SAPE	5.5 mL	1 bottle
Bead Diluent	MXBD-2	3.5 mL	1 bottle
Mixing Bottle	----------	----------	1 bottle

Rat Kidney Toxicity Panel 2 Antibody-Immobilized Beads:

Bead/Analyte Name	Luminex Bead Region	Customizable Beads (20X Concentration, 200 µL)	
		Available	Cat. #
Anti – Albumin Bead	3	✓	RALBMN
Anti – β-2-Microglobulin Bead	6	✓	RB2MG
Anti – Cystatin C Bead	10	✓	RCYSTNC

MATERIALS REQUIRED BUT NOT PROVIDED

Reagents

1. Luminex Sheath Fluid (Luminex Catalogue #40-50000)

Instrumentation / Materials

1. Adjustable Pipettes with Tips capable of delivering 25 µL to 1000 µL
2. Multichannel Pipettes capable of delivering 5 µL to 50 µL or 25 µL to 200 µL
3. Reagent Reservoirs
4. Polypropylene Microfuge Tubes
5. Rubber Bands
6. Absorbent Pads
7. Laboratory Vortex Mixer
8. Sonicator (Branson Ultrasonic Cleaner Model #B200 or equivalent)
9. Titer Plate Shaker (Lab-Line Instruments Model #4625 or equivalent)
10. Vacuum Filtration Unit (Millipore Vacuum Manifold Catalog #MSVMHTS00 or equivalent with Millipore Vacuum Pump Catalog #WP6111560 or equivalent)
11. Luminex 100™ IS, 200™, HTS by Luminex Corporation
12. Plate Stand (Millipore Catalog # MX-STAND)

SAFETY PRECAUTIONS

- All biological materials should be handled as potentially hazardous. Follow universal precautions as established by the Centers for Disease Control and Prevention and by the Occupational Safety and Health Administration when handling and disposing of infectious agents.
- Sodium azide or Proclin has been added to some reagents as a preservative. Although the concentrations are low, sodium azide and Proclin may react with lead and copper plumbing to form highly explosive metal azides. On disposal, flush with a large volume of water to prevent azide buildup.

TECHNICAL GUIDELINES

To obtain reliable and reproducible results, the operator should carefully read this entire manual and fully understand all aspects of each assay step before running the

assay. The following notes should be reviewed and understood before the assay is set up.

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- Do not use beyond the expiration date on the label.
- Do not mix or substitute reagents with those from other lots or sources.
- The Antibody-Immobilized Beads are light sensitive and must be protected from light at all times. Cover the assay plate containing beads with opaque plate lid or aluminum foil during all incubation steps.
- It is important to allow all reagents to warm to room temperature (20-25°C) before use in the assay.
- The bottom of the Microtiter Filter Plate should not be in direct contact with any surface during assay setup or incubation times. The plate can be set on a plate holder or on the non-flat side of the plate cover or any other plate holder to raise the plate from the surface. A plate stand can be purchased separately from Millipore (Millipore Catalog #MX-STAND).
- Incomplete washing can adversely affect the assay outcome. All washing must be performed with the Wash Buffer provided.
- After the wash steps, keep the bottom of the Microtiter Filter Plate clean by blotting on paper towels or absorbent pads to prevent any leakage due to capillary action.
- Keep the vacuum suction on the plate as low as possible. It is recommended to have a vacuum setting that will remove 200 µL of buffer in ≥ 5 seconds (equivalent to < 100 mmHg).
- After hydration, all Standards and Controls must be transferred to polypropylene tubes.
- The Standards prepared by serial dilution must be used within 1 hour of preparation. Discard any unused standards except the standard stock, which may be stored at ≤ -20°C for 1 month and at ≤ -80°C for greater than one month.
- If samples fall outside the dynamic range of the assay, further dilute the samples with the appropriate diluent and repeat the assay.
- Any unused mixed Antibody-Immobilized Beads may be stored in the Mixing Bottle at 2-8°C for up to one month.
- During the preparation of the standard curve, make certain to mix the higher concentration well before making the next dilution. Use a new tip with each dilution.
- The plate should be read immediately after the assay is finished. If, however, the plate cannot be read immediately, seal the plate, cover with opaque lid and store the plate at 2-8°C for up to 24 hours. Prior to reading, agitate the plate on the plate shaker at room temperature for 10 minutes. Delay in reading a plate may result in decreased sensitivity for some analytes.
- The titer plate shaker should be set at a speed to provide maximum orbital mixing without splashing of liquid outside the wells. For the recommended

plate shaker, this would be a setting of 5-7, which is approximately 500-800 rpm.

- Ensure that the needle probe is clean. This may be achieved by sonication and/or alcohol flushes. Adjust probe height according to the protocols recommended by Luminex to the kit filter plate using 3 alignment discs prior to reading an assay.
- Vortex all reagents well before adding to plate.

SAMPLE COLLECTION AND STORAGE

A. Preparation of Urine Samples:

- Centrifuge the samples briefly to pellet debris. Assay immediately or aliquot and store samples at ≤ -20°C.
- Avoid multiple (>2) freeze/thaw cycles.
- When using frozen samples, it is recommended to thaw the samples completely, mix well by vortexing and centrifuge prior to use in the assay to remove particulates.
- **Rat urine sample should be diluted 1:500 using Assay Buffer as the sample diluent.** Dilute samples 1:10 in Assay Buffer (e.g. 2 µL sample into 18 µL Assay Buffer), then dilute again 1:50 in Assay Buffer (e.g. 2 µL sample into 98 µL Assay Buffer) to achieve a 1:500 dilution. Depending on strain used and treatment conditions, the dilution factor may vary. Use Assay Buffer for these dilutions as well.

NOTE:

- A maximum of 25 µL per well of urine sample can be used.
- All samples must be stored in polypropylene tubes. **DO NOT STORE SAMPLES IN GLASS.**
- Avoid debris, lipids and cells when using samples with gross hemolysis or lipemia.

PREPARATION OF REAGENTS FOR IMMUNOASSAY

A. Preparation of Antibody-Immobilized Beads

Sonicate each individual antibody-bead vial for 30 seconds then vortex for 1 minute. Add 150 µL from each antibody-bead vial to the Mixing Bottle and bring final volume to 3.0 mL with Bead Diluent. Vortex the mixed beads well. Unused portion may be stored at 2-8°C for up to one month.

Example 1: When using 1 antibody-immobilized bead, add 150 µL from the bead vial to the Mixing Bottle. Then add 2.85 mL Bead Diluent.

Example 2: When using 3 antibody-immobilized beads, add 150 µL from each of the 3 bead vials to the Mixing Bottle. Then add 2.55 mL Bead Diluent.

B. Preparation of Quality Controls

Before use, reconstitute Quality Control 1 and Quality Control 2 with 250 µL deionized water. Invert the vial several times to mix then vortex briefly. Allow the vial to sit for 5-10 minutes and then transfer the controls to appropriately labeled polypropylene microfuge tubes. Unused portions may be stored at ≤ -20°C for up to one month.

C. Preparation of Wash Buffer

Bring the 10X Wash Buffer to room temperature and mix to bring all salts into solution. Dilute 30 mL of 10X Wash Buffer with 270 mL deionized water. Store unused portion at 2-8°C for up to one month.

E. Preparation of Rat Kidney Toxicity Panel 2 Standard

1.) Prior to use, reconstitute the Rat Kidney Toxicity Panel 2 Standard with 250 µL deionized water (refer to table below for exact analyte concentrations). Invert the vial several times to mix. Vortex the vial for 10 seconds. Allow the vial to sit for 5-10 minutes and then transfer the standard to an appropriately labeled polypropylene microfuge tube. This will be used as Standard 7; the unused portion may be stored at ≤ -20°C for up to one month.

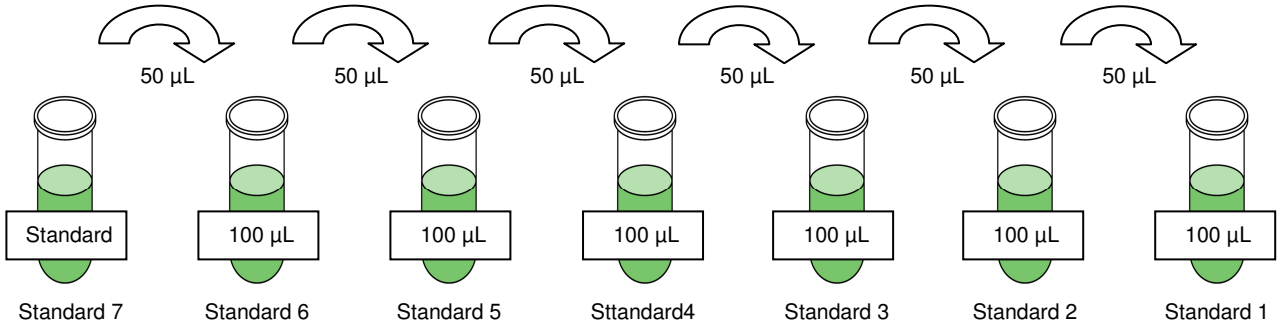
2.) Preparation of Working Standards

Label six polypropylene microfuge tubes Standard 6, Standard 5, Standard 4, Standard 3, Standard 2 and Standard 1. Add 100 µL of Assay Buffer to each of the six tubes. Prepare serial dilutions by adding 50 µL of the reconstituted standard (Standard 7) to the Standard 6 tube, mix well and transfer 50 µL of the Standard 6 tube to the Standard 5 tube mix well and transfer 50 µL of the Standard 5 tube to the Standard 4 tube, mix well and transfer 50 µL of the Standard 4 tube to the Standard 3 tube, mix well and transfer 50 µL of the Standard 3 tube to the Standard 2 tube, mix well and transfer 50 µL of the Standard 2 tube to the Standard 1 tube and mix well. The 0 pg/mL Standard (Background) will be Assay Buffer.

Standard (Tube #)	**Volume of Deionized Water to Add**	**Volume of Standard to Add**
Standard 7 (reconstituted standard)	250 µL	0

Standard (Tube #)	**Volume of Assay Buffer to Add**	**Volume of Standard to Add**
Standard 6	100 µL	50 µL of Standard 7
Standard 5	100 µL	50 µL of Standard 6
Standard 4	100 µL	50 µL of Standard 5
Standard 3	100 µL	50 µL of Standard 4
Standard 2	100 µL	50 µL of Standard 3
Standard 1	100 µL	50 µL of Standard 2

Preparation of Standards



After dilution, each tube has the following concentrations for each analyte:

Standard	Albumin (pg/mL)	β-2-Microglobulin (pg/mL)	Cystatin C (pg/mL)
Standard 1	5,487	137	34
Standard 2	16,461	412	103
Standard 3	49,383	1235	309
Standard 4	148,148	3704	926
Standard 5	444,444	11,111	2,778
Standard 6	1,333,333	33,333	8,333
Standard 7	4,000,000	100,000	25,000

IMMUNOASSAY PROCEDURE

- Prior to beginning this assay, it is imperative to read this protocol completely and to thoroughly understand the Technical Guidelines.
- Allow all reagents to warm to room temperature (20-25°C) before use in the assay.
- Diagram the placement of Background, Standards 1 through 7, Controls 1 and 2, and Samples on Well Map Worksheet in a vertical configuration. (Note: Most instruments will only read the 96-well plate vertically by default.) It is recommended to run the samples in duplicate.
- Set the filter plate on a plate holder at all times during reagent dispensing and incubation steps so that the bottom of the plate does not touch any surface.

1. Prewet the filter plate by pipetting 200 µL of Assay Buffer into each well of the Microtiter Filter Plate. Seal and mix on a plate shaker for 10 minutes at room temperature (20-25°C).
2. Remove Wash Buffer by vacuum. **(NOTE: DO NOT INVERT PLATE.)** Blot excess Wash Buffer from the bottom of the plate with an absorbent pad or paper towels.
3. Add 25 µL Assay Buffer to Background wells..
4. Add 25 µL of each Standard or Control into the appropriate wells.
5. Add 25 µL of Sample into the appropriate wells.
6. Add 25 µL of Biotinylated Albumin to each well. If Albumin is not one of the selected analytes for the panel, add 25 µL of Assay Buffer instead.
7. Vortex Mixing Bottle and add 25 µL of the Mixed Beads to each well. (Note: During addition of Mixed Beads, shake Mixing Bottle intermittently to avoid settling.)
8. Seal the plate with a plate sealer then cover it with the opaque lid. Wrap a rubber band around the plate holder, plate and lid and incubate with agitation on a plate shaker overnight at 4°C. Alternatively incubate with agitation on a plate shaker for 2 hours at room temperature.

Add 200 µL Assay Buffer per well

↓ Shake 10 min, RT

↓ Vacuum

- Add 25 µL Assay Buffer to Background wells
- Add 25 µL Standard or Control to appropriate wells
- Add 25 µL Sample to sample wells
- Add 25 µL Biotinylated Albumin (use if Albumin is one of the selected analytes) or 25 µL Assay Buffer (use if Albumin is excluded) to each well
- Add 25 µL Beads to each well

↓ Incubate overnight at 4°C with shaking

9. Gently remove fluid by vacuum. **(NOTE: DO NOT INVERT PLATE.)**

10. Wash plate 2 times with 200 µL/well of Wash Buffer, removing Wash Buffer by vacuum filtration between each wash. Blot excess Wash Buffer from the bottom the plate by with an absorbent pad or paper towels.

11. Add 50 µL of Detection Antibodies into each well. (Note: Allow the Detection Antibodies to warm to room temperature prior to addition.)

12. Seal, cover with lid, and incubate with agitation on a plate shaker for 1 hour at room temperature (20-25°C). **DO NOT VACUUM AFTER INCUBATION.**

13. Add 50 µL Streptavidin-Phycoerythrin to each well containing the 50 µL of Detection Antibodies.

14. Seal, cover with lid and incubate with agitation on a plate shaker for 30 minutes at room temperature (20-25°C).

15. Gently remove all contents by vacuum. **(NOTE: DO NOT INVERT PLATE.)**

16. Wash plate 2 times with 200 µL/well Wash Buffer, removing Wash Buffer by vacuum filtration between each wash. Wipe any excess buffer on the bottom of the plate with an absorbent pad or paper towel.

17. Add 100 µL of Sheath Fluid to all wells. Resuspend the beads by incubating with agitation on a plate shaker for 5 minutes at room temperature.

18. Run plate on Luminex 100™ IS, 200™, HTS.

19. Save and analyze the Median Fluorescent Intensity (MFI) data using a 5-parameter logistic or spline curve-fitting method to calculate analyte concentrations in samples. Multiply calculated concentrations by 500 to determine sample concentration.

Vacuum and wash 2X with 200 µL Wash Buffer

Add 50 µL Detection Antibodies per well

Incubate 1 hour at RT

Do not vacuum

Add 50 µL Streptavidin-Phycoerythrin per well

Incubate for 30 minutes at RT

Vacuum and wash 2X with 200µL Wash Buffer

Add 100 µL Sheath Fluid per well

Read on Luminex (50µL, 50 beads per bead set)

EQUIPMENT SETTINGS

These specifications are for the Luminex 100™ IS v.1.7, Luminex 100™ IS v2.1/2.2, Luminex 200™ v2.3, xPONENT®, and Luminex HTS. Luminex instruments with other software (e.g. MasterPlex®, StarStation®, LiquiChip®, Bio-Plex®, LABScan®100) would need to follow instrument instructions for gate settings and additional specifications from the vendors.

Events:	50 per bead	
Sample Size:	50 µL	
Gate Settings	8,000 to 15,000	
Time Out	60 seconds	
Bead Set:	3-plex Bead Regions	
	Albumin	3
	β-2-Microglobulin	6
	Cystatin C	10

QUALITY CONTROLS

The ranges for each analyte in Quality Control 1 and 2 are provided on the card insert or can be located at the MILLIPORE website www.millipore.com/techlibrary/index.do using the catalog number as the keyword.

ASSAY CHARACTERISTICS

Assay Sensitivities (minimum detectable concentrations, pg/mL)

MinDC: Minimum Detectable Concentration is calculated using the Milliplex Analyst Software from Millipore. It measures the true limits of detection for an assay by mathematically determining what the empirical MinDC would be if an infinite number of standard concentrations were run for the assay under the same conditions.

Analyte	*2 Hour Protocol**	*Overnight Protocol***
Albumin	3588.5	9373.5
β-2-Microglobulin	39.0	74.2
Cystatin C	5.7	7.1

*mean minDC + 2 standard deviations, N=3 assays
**mean minDC + 2 standard deviations, N=7 assays

Precision

Intra-assay precision is generated from the mean of the %CV's from two to four reportable results across different concentrations of analytes in five different assays. Inter-assay precision is generated from the mean of the %CV's from two to four reportable results across different concentrations of analytes across five different assays.

Analyte	*Intra-Assay (%CV)*	*Inter-Assay (%CV)*
Albumin	19.5	19.5
β-2-Microglobulin	9.0	9.0
Cystatin C	14.3	14.3

Accuracy

Spike Recovery: The data represent mean percent recovery of 3 levels of spiked standards in urine samples.

Analyte	*Spike Recovery in Urine*
Albumin	103.8
β-2-Microglobulin	118.9
Cystatin C	111.5

Sample Linearity of Dilution (% Recovery):

Dilution Linearity, defined as the analyte levels measured in diluted samples as a percentage of neat urine samples, was determined by measuring analyte concentrations in 5 independent samples. Samples were first diluted 1:250 in Assay Buffer then further diluted to 1:1000, 1:4000 and 1:16000 in the kit Assay Buffer. These samples were analyzed in duplicate. The average percent recovery is reported.

Analyte	***Average Percent Recovery***
Albumin	156.3
β-2-Microglobulin	105.4
Cystatin C	133.7

Cross-Reactivity

There was no or negligible cross-reactivity between the antibodies for an analyte and any of the other analytes in this panel.

TROUBLESHOOTING GUIDE

Problem	Probable Cause	Solution
Filter plate will not vacuum	Vacuum pressure is insufficient	Increase vacuum pressure such that 0.2mL buffer can be suctioned in 3-5 seconds.
	Samples have insoluble particles	Centrifuge samples just prior to assay setup and use supernatant. If high lipid concentration, after centrifugation, remove lipid layer and use supernatant.
	Sample too viscous	May need to dilute sample.
Insufficient bead count	Vacuum pressure too high	Adjust vacuum pressure such that 0.2mL buffer can be suctioned in 3-5 seconds.
	Bead mix prepared incorrectly	Sonicate bead vials and vortex just prior to adding to Mixing Bottle according to protocol. Agitate bead mix intermittently in reservoir while pipetting into the plate.
	Samples cause interference due to particulate matter or viscosity	See above. Also sample probe may need to be cleaned with alcohol flush, backflush and washes; or, if needed, probe should be removed and sonicated.
	Probe height not adjusted correctly	Adjust probe to 3 alignment discs in well H6.
Plate leaked	Vacuum pressure too high	Adjust vacuum pressure such that 0.2mL buffer can be suctioned in 3-5 seconds. May need to transfer contents to a new (prewetted) plate and continue.
	Plate set directly on table or absorbent towels during incubations or reagent additions	Set plate on plate holder or raised edge so bottom of filter is not touching any surface.
	Insufficient blotting of filter plate bottom causing wicking	Blot the bottom of the filter plate well with absorbent towels after each wash step.
	Pipette touching plate filter during additions	Pipette to the side of well.
	Probe height not adjusted correctly	Adjust probe to 3 alignment discs in well H6.
Background is too high	Background wells were contaminated	Avoid cross-well contamination by using sealer appropriately and by pipetting with multichannel pipets without touching reagent in plate.
	Matrix used has endogenous analyte or interference	Check matrix ingredients for crossreacting components (e.g. interleukin modified tissue culture medium).
	Insufficient washes	Increase number of washes.

Beads not in region or gate	Luminex not calibrated correctly or recently	Calibrate Luminex based on instrument manufacturer's instructions, at least once a week or if temperature has changed by >3°C.
	Gate settings not adjusted correctly	Some Luminex instruments (e.g. Bioplex) require different gate settings than those described in the kit protocol. Use instrument default settings.
	Wrong bead regions in protocol template	Check kit protocol for correct bead regions or analyte selection.
	Incorrect sample type used	Samples containing organic solvents or if highly viscous should be diluted or dialyzed as required.
	Instrument not washed or primed	Prime the Luminex 4 times to eliminate air bubbles. Wash 4 times with sheath fluid or water if there is any remnant alcohol or sanitizing liquid.
	Beads were exposed to light	Keep plate and bead mix covered with dark lid or aluminum foil during all incubation steps.
Signal for whole plate is same as background	Incorrect or no Detection Antibody was added	Add appropriate Detection Antibody and continue.
	Streptavidin-Phycoerythrin was not added	Add Streptavidin-Phycoerythrin according to protocol. If Detection Antibody has already been vacuumed out, sensitivity may be low.
Low signal for standard curve	Detection Antibody may have been vacuumed out prior to adding Streptavidin Phycoerythrin	May need to repeat assay if desired sensitivity not achieved.
	Incubations done at incorrect temperatures, timings or agitation	Assay conditions need to be checked.
Signals too high, standard curves are saturated	Calibration target value set too high	With some Luminex instruments (e.g. Bio-plex) default target setting for RP1 calibrator is set at High PMT. Use low target value for calibration and reanalyze plate.
	Plate incubation was too long with standard curve and samples	Use shorter incubation time.
Sample readings are out of range	Samples contain no or below detectable levels of analyte	If below detectable levels, it may be possible to use higher sample volume. Check with tech support for appropriate protocol modifications.
	Samples contain analyte concentrations higher than highest standard point	Samples may require dilution and reanalysis for that particular analyte.
	Standard curve was saturated at higher end of curve	See above.

High variation in samples and/or standards	Multichannel pipet may not be calibrated	Calibrate pipets.
	Plate washing was not uniform	Confirm all reagents are vacuumed out completely in all wash steps.
	Samples may have high particulate matter or other interfering substances	See above.
	Plate agitation was insufficient	Plate should be agitated during all incubation steps using a vertical plate shaker at a speed where beads are in constant motion without causing splashing.
	Cross-well contamination	Check when reusing plate sealer that no reagent has touched sealer. Care should be taken when using same pipet tips that are used for reagent additions and that pipet tip does not touch reagent in plate.

REPLACEMENT REAGENTS

Components	Cat #
Rat Kidney Toxicity Panel 2 Standard	RKTX-8037-2
Rat Kidney Toxicity Panel 2 Quality Controls 1 and 2	RKTX-6037-2
Rat Kidney Toxicity Panel 2 Biotinylated Albumin	RKTX-BTAB
Rat Kidney Toxicity Panel 2 Detection Antibodies	RKTX-1037-2
Streptavidin-Phycoerythrin	L-SAPE
Assay Buffer	L-AB1
Bead Diluent	MXBD-2
Set of two 96-Well Filter Plates with Sealers	MX-PLATE
10X Wash Buffer	L-WB

Antibody-Immobilized Beads

Kidney Toxicity Biomarker	Bead #	Cat. #
Albumin	3	RALBMN
β-2-Microglobulin	6	RB2MG
Cystatin C	10	RCYSTNC

ORDERING INFORMATION

To place an order:

To assure the clarity of your custom Rat Kidney Toxicity Panel 2 kit order, please FAX the following information to our customer service department:

- Your name, telephone and/or fax number
- Customer account number
- Shipping and billing address
- Purchase order number
- Catalog number and description of product
- Quantity of kits
- Selection of MILLIPLEX® Rat Kidney Toxicity Panel 2 Analyte Requirements

FAX: (636) 441-8050
Toll-Free US: (800) MILLIPORE

Mail Orders: Millipore Corp.
6 Research Park Drive
St. Charles, Missouri 63304 U.S.A.

For International Customers:

To best serve our international customers in placing an order or obtaining additional information about MILLIPLEX® MAP products, please contact your multiplex specialist or sales representative or email our European Customer Service at customerserviceEU@Millipore.com.

Conditions of Sale

All products are for research use only. They are not intended for use in clinical diagnosis or for administration to humans or animals. All products are intended for *in vitro* use only.

Material Safety Data Sheets (MSDS)

Material Safety Data Sheets for Millipore products may be ordered by fax or phone or through our website at www.millipore.com/techlibrary/index.do

WELL MAP

	1	2	3	4	5	6	7	8	9	10	11	12
A	0 pg/mL Standard (Background)	Standard 4	QC-1 Control	Etc.								
B	0 pg/mL Standard (Background)	Standard 4	QC-1 Control									
C	Standard 1	Standard 5	QC-2 Control									
D	Standard 1	Standard 5	QC-2 Control									
E	Standard 2	Standard 6	Sample 1									
F	Standard 2	Standard 6	Sample 1									
G	Standard 3	Standard 7	Sample 2									
H	Standard 3	Standard 7	Sample 2									