PeliKine compact[™] human IL-10

M1910



REF

Introduction

Assay procedure	was the demons cytokines by TH synthesis inhibito lineages, like TH: regulator, IL-10 i human and mous include IL-1, GM macrophages, ar compared with o substantial amou macrophage secr Nevertheless, IL enhancement of induction of diff lines [11], suppo expression on B- Bioassays for the produced by TH' These bioassays other substances The Pelikine com reproducible and as well as in cell-	the demonstration that supernatants from activated T-cells could inhibit the secretion of kines by TH1-cell clones [1,2]. Later it was found that IL-10, also known as cytokine besis inhibitory factor (CSIF) is an immunoregulatory protein produced by a number of cell ges, like TH2-cells, B-cells and activated monocytes [1,3-8]. As an immune system down- ator, IL-10 inhibits the synthesis of several cytokines that are normally secreted by both an and mouse monocytes/macrophages in response to activation by LPS. These cytokines de IL-1, GM-CSF, TNF, IL-6, IL-8, IL-10 and IL-12 [1,6,8]. IL-10 is secreted by activated ophages, and this is also inhibited by IL-10 [6]. The secretion of IL-10 is relatively late bared with other cytokines, which may explain why macrophages are able to secrete tantial amounts of various cytokines before IL-10 inhibition occurs. IFNγ also inhibits ophage secretion of IL-10 [9], resulting in direct cross-inhibition of IL-10 and IFNγ. wrtheless, IL-10 has also been shown to have up-regulating capacities, including the ncement of <i>in vitro</i> proliferation of IL-2 and/or IL-4 induced mouse thymocytes, the ction of differentiation and proliferation of activated B-cell lines into antibody secreting [11], supporting the growth of mast cell lines [12] and induction of class II MHC antigen ession on B-cells [11,12]. says for the quantification of IL-10, based on the inhibition of IFNγ and other cytokines uced by TH1-cells in response to stimulation by antigen have been used for several years. e bioassays, although sensitive, are time consuming and susceptible to interference by r substances. Pelikine compact™ human IL-10 ELISA kit [10] has been developed for faster, more ducible and specific quantification of human IL-10 in serum, plasma and other body fluids, ell as in cell-culture supernatant. Assay procedure for PeliKine TM compact ELISA kit.				
Kitcomponent list	Quantity	Kit component		Volume	Cap colour	
	1 vial	coating antibody	100-fold concentrated	375 <i>µ</i> l	red	
	1 vial	blocking reagent	50-fold concentrated	2 ml	transparent	
	2 vials	IL-10 standard (lyophilized)	see label	1000 <i>µ</i> i	-	
	1 vial	biotinylated antibody	100-fold concentrated	375 <i>µ</i> I	yellow	
	1 vial	streptavidin-poly-HRP conjugate	10,000-fold concentrated	20 <i>µ</i> I	brown	
	1 bottle	HPE-dilution buffer	5- fold concentrated	55 ml		
	3 pcs	microtiter plate + lid	-	-		
	10 pcs	plate seals	-	-		
Sensitivity Expected values Specificity	 MEAN calculated zero signal + 3 SD : 1 – 3 pg/ml (shake – static incubation) 2x (MEAN calculated zero signal) : 3 – 5 pg/ml (shake – static incubation) IL-10 values in fresh serum and plasma of healthy individuals are below 5 pg/ml. No crossreactivity was observed with the following recombinant human proteins: IL-1α, IL-1ß, IL-2 4, IL-5, IL-6, sIL-6r (GP80), IL-7, IL-8, IL-9, IL-11, IL-13, Macrophage Colony Stimulating Factor (M Granulocyte Colony Stimulating Factor (G-CSF), Granulocyte/Macrophage Colony Stimulating Factor CSF), Leukemia Inhibitory Factor (LIF), RANTES, Stem Cell Factor/ Mast Cell Factor (SCF/MCF), Transforming Growth Factor β-1 (TGFβ-1), Tumour Necrosis Factor α (TNFα), Tumour Necrosis Factor (TNFβ/Lymphotoxin), and Interferon γ (IFNγ). 					
References	 Fiorentino, D.F. <i>et al</i> (1989) J.Exp.Med. <u>170</u>: 2081 Moore, K.W. <i>et al</i> (1990) Science <u>248</u>: 1230 Zlotnik, A. and Moore, K.W. (1991) Cytokine <u>3</u>: 366 Yssel, H. <i>et al</i> (1992) J.Immunol. <u>1</u>: 2378 Benjamin, D. <i>et al</i> (1992) Blood <u>80</u>: 128 De Waal Malefyt, R. <i>et al</i> (1991) J.Exp.Med. <u>174</u>: 1209 Mosmann, T.R. (1994) Adv.Immunol. <u>56</u>: 1 Thomson, A.W. (1994) The cytokine handbook. Academic Press ISBN 0-12-689661-5 					

The initial discovery that led to the characterization and cDNA cloning of Interleukin 10 (IL-10)

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- 10. Llorente, L. *et al* (1993) Eur.Cyt.Netw. 4: 421
- 11. Go, N.F. et al (1990) J.Exp.Med. <u>172</u>: 1625
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Standard	A recombinant human IL-10 standard has been calibrated against the WHO International Reference Preparation (IL-10 92/516; National Institute for Biological Standards and Control, Potters Bar, Hertfordshire, U.K. 1 WHO Unit = 200 pg IL-10).			
	The kit contains two lyophilized vials with natural human IL-10.			
	Reconstitute the lyophilized standard by adding 1000 μ l of distilled water to the vial. Incubate for 10 minutes at room temperature and mix gently. After reconstitution the standard must be kept cold (2-8°C) and stored frozen after use (<-18°C, preferably<-70°C).			
Standard curve	Label 7 tubes, one tube for each dilution: 300, 120, 48, 19.2, 7.7, 3.1 and 1.2 pg/ml. Pipette 470 μ l of working-strength dilution buffer into the tube labelled 300 pg/ml and 300 μ l of working-strength dilution buffer into the other tubes. Transfer 30 μ l of the IL-10 standard (5000 pg/ml) into the first tube labelled 300 pg/ml, mix well and transfer 200 μ l of this dilution into the second tube labelled 120 pg/ml. Repeat the serial dilution's six more times by adding 200 μ l of the previous tube of diluted standard to the 300 μ l of dilution buffer.			
	The standard curve will contain 300, 120, 48, 19.2, 7.7, 3.1, 1.2 and 0 pg/ml (dilution buffer).			
	It is recommended to prepare two separate series for each assay.			
Samples	It is recommended to dilute the test samples at least 1:2 in working-strength dilution buffer. If high levels of IL-10 (outside the standard curve) are expected in the test samples, additional dilutions of sample i.e. 1:10 and 1:50 should also be prepared.			
Typical standard curve	A ₄₅₀			
	3.00			
	shaken			
	1.00 = incubation			
	static			

0.10

1

	STATIC INCUBATION	SHAKEN INCUBATION	
	Calculated mean absorbance at 450 nm		
substrate blank	0	0	
0 pg/ml	0.021	0.067	
1.2 pg/ml	0.033	0.077	
3.1 pg/ml	0.049	0.178	
7.7 pg/ml	0.089	0.215	
19.2 pg/ml	0.172	0.408	
48 pg/ml	0.380	0.950	
120 pg/ml	0.917	2.259	
300 pg/ml	1.886	> 3.000	

10

IL-10 (pg/ml)

incubation

100

DO NOT USE THESE DATA TO CONSTRUCT A STANDARD CURVE FOR SAMPLE VALUE CALCULATIONS