

Human Apolipoprotein AV (APO AV)

96-Well Plate

Cat. # EZHAP0AV71K

# HUMAN APOLIPOPROTEIN AV (APO AV) ELISA KIT 96-Well Plate (Cat. # EZHAPOAV71K)

١.	Intended Use	2
.	Principles Of Procedure	2
III.	Reagents Supplied	2
IV.	Storage and Stability	4
٧.	Reagent Precautions	4
VI.	Materials Required But Not Provided	5
VII.	Sample Collection And Storage	5
VIII.	Sample Preparation	5
IX.	Reagent Preparation	6
Χ.	,	8
XI.	Microtiter Plate Arrangement	11
XII.	Calculations	12
	Interpretation	12
XIV.	Standard Curve	13
XV.	Assay Characteristics	14
XVI.	Troubleshooting Guide	17
VII.	Replacement Reagents	17
$\times$	Ordering Information	18

# HUMAN APO AV ELISA KIT 96-Well Plate (Cat. # EZHAPOAV71K)

### I. INTENDED USE

This Human Apolipoprotein AV (Apo AV) ELISA kit is used for the non-radioactive quantification of Human Apo AV in serum, plasma, and cell culture samples. This kit specifically measures Human Apo AV and has no cross reactivity to Mouse or Rat Apo AV. One kit is sufficient to measure 38 unknown samples in duplicate.

This kit is for research purpose only.

### II. PRINCIPLES OF PROCEDURE

This assay is a Sandwich EUSA based, sequentially, on: 1) capture of Human Apo AV molecules from samples to the wells of a microtiter plate coated with a monoclonal anti-human Apo AV antibody, 2) washing of unbound materials from samples, 3) binding of a second biotinylated polyclonal anti-human Apo AV antibody to the captured molecules, 4) washing of unbound materials from samples, 5) binding of streptavidin-horseradish peroxidase conjugate to the immobilized biotinylated antibodies, 6) washing of excess of free enzyme conjugates, and 7) quantification of immobilized antibody-enzyme conjugates by monitoring horseradish peroxidase activities in the presence of the substrate 3,3',5,5'-tetramethylbenzidine. The enzyme activity is measured spectrophotometrically by the increased absorbance at 450 nm – 590 nm after acidification of formed products. Since the increase in absorbance is directly proportional to the amount of captured Human Apo AV in the unknown sample, the latter can be derived by interpolation from a reference curve generated in the same assay with reference standards of known concentrations of Human Apo AV.

#### III. REAGENTS SUPPLIED

Each kit is sufficient to run one 96-well plate and contains the following reagents:

### A. Microtiter Plate

Coated with Monoclonal anti-Human Apo AV Antibody

Quantity: 1 strip plate Preparation: Ready to Use

Note: Unused strips should be resealed in the foil pouch with the dessicant provided and

stored at 2-8 °C.

### B. Adhesive Plate Sealer

Quantity: 2 sheets

Preparation: Ready to Use

### III. REAGENTS SUPPLIED (continued)

### C. 10X Concentrate HRP Wash Buffer

10X concentrate of 50 mM Tris Buffered Saline containing Tween-20

Quantity: 2 bottles containing 50 ml each

Preparation: Dilute 1:10 with distilled or deionized water

#### D. Matrix Solution

Charcoal Stripped Human Serum

Quantity: 1ml

Preparation: Dilute 1:5 with Assay Buffer (AB-PTR)

### E. Human Apo AV Standard

Purified Recombinant Human Apo AV, lyophilized

Quantity: 0.5ml upon hydration

Preparation: Reconstitute with 0.5ml distilled or deionized water.

# F. Quality Controls 1 and 2

One vial each, lyophilized, containing recombinant human Apo AV at two different levels.

Quantity: 0.5ml upon hydration

Preparation: Reconstitute each control with 0.5ml distilled or deionized water.

## G. Assay Buffer (Sample Diluent)

0.05M Phosphosaline containing 0.025M EDTA, 0.08% Sodium Azide, 1% BSA

Quantity: 40 ml

Preparation: Ready to Use

# H. Assay Running Buffer

0.05M Phosphosaline containing 0.025M EDTA, 0.08% Sodium Azide, 1% BSA

Quantity: 11 ml

Preparation: Ready to Use

# I. Human Apo AV Detection Antibody (1000X)

Pre-titered Biotinylated Rabbit anti-Human Apo AV Antibody

Quantity: 20 µl

Preparation: Dilute 1:1000 with Assay Buffer (AB-PTR)

# J. Enzyme Solution

Pre-titered Streptavidin-Horseradish Peroxidase Conjugate in Buffer

Quantity: 12 ml

Preparation: Ready to Use

### III. REAGENTS SUPPLIED (continued)

# K. Substrate (Light sensitive, avoid unnecessary exposure to light)

3, 3', 5, 5'-tetramethylbenzidine in buffer

Quantity: 12 ml

Preparation: Ready to Use.

## L. Stop Solution (Caution: Corrosive Solution)

0.3 M HCl

Quantity: 12 ml

Preparation: Ready to Use

### IV. STORAGE AND STABILITY

Prior to use, all components in the kit can be stored up to 2 weeks at  $2\text{-}8^{\circ}\text{C}$ . For longer storage (> 2 weeks), freeze diluted HRP Wash Buffer, Assay Buffer, and reconstituted Apo AV Standards and Controls at  $\leq -20^{\circ}\text{C}$ . Minimize repeated freeze and thaw of the Apo AV Standards and Quality Controls. Refer to expiration dates on all reagents prior to use. Do not mix reagents from different kits unless they have the same lot numbers.

### V. REAGENT PRECAUTIONS

#### A. Sodium Azide

Sodium Azide has been added to certain reagents as a preservative. Although the concentrations are low, Sodium Azide may react with lead and copper plumbing to form highly explosive metal azides. On disposal, flush with a large volume of water to prevent azide build up.

# B. Hydrochloric Acid

Hydrochloric Acid is corrosive and can cause eye and skin burns. It is harmful if swallowed and can cause respiratory and digestive tract burns. Avoid contact with skin and eyes. Do not swallow or ingest.

### VI. MATERIALS REQUIRED BUT NOT PROVIDED

- 1. Pipettes and Pipette Tips: 10 µl 20 µl or 20 µl 100 µl
- 2. Multi-Channel Pipettes and Pipette Tips:  $5 \sim 50 \, \mu l$  and  $50 \sim 300 \, \mu l$
- 3. Buffer and Reagent Reservoirs
- 4. Vortex Mixer
- Deionized Water
- 6. Microtiter Plate Reader capable of reading absorbency at 450 nm
- 7. Orbital Microtiter Plate Shaker
- 8. Absorbent Paper or Cloth

### VII. SAMPLE COLLECTION AND STORAGE

- 1. To prepare serum samples, whole blood is directly drawn into a centrifuge tube that contains no anti-coagulant. Let blood clot at room temperature for 30 min.
- 2. Promptly centrifuge the clotted blood at 2,000 to 3,000 x g for 15 minutes at  $4 \pm 2^{\circ}$ C.
- 3. Transfer and store serum samples in separate tubes. Date and identify each sample.
- Use freshly prepared serum or aliquot and store samples at ≤ -20°C for later use. For long-term storage, keep at -70°C. Avoid freeze/thaw cycles.
- 5. To prepare plasma samples, whole blood should be collected into centrifuge tubes containing enough K<sub>3</sub>EDTA to achieve a final concentration of 1.735 mg/ml and centrifuged immediately after collection. Observe the same precautions in the preparation of serum samples.
- 6. If heparin is to be used as an anticoagulant, the effect on the assay outcome at the dose of heparin used should be pre-determined.
- 7. Avoid using samples with gross hemolysis or lipemia.

### VIII. SAMPLE PREPARATION

- 1. Allow all the reagents to come to room temperature.
- 2. Dilute serum or plasma samples 1:5 in Assay Buffer (AB-PTR). Recommended dilution: dilute 20 µl of serum or plasma in 80 µl of Assay Buffer (AB-PTR). Cellular extract and culture media dilutions will vary.

### IX. REAGENT PREPARATION

# A. Matrix Solution Preparation

Dilute Matrix Solution 1:5 in Assay Buffer (AB-PTR). Recommended dilution: dilute 500 µl Matrix Solution in 2 ml of Assay Buffer (AB-PTR). Mix well.

## B. Detection Antibody Preparation

Dilute (1000X) Detection Antibody 1:1000 in Assay Buffer (AB-PTR).

Recommended dilution: 12µl Detection Antibody in 11.988 ml of Assay Buffer (AB-PTR). Mix well.

## C. Standard and Quality Control Preparation

## Human Apo AV Standard Preparation

- 1. Use care in opening the lyophilized Standard vial. Using a pipette, reconstitute the Human Apo AV Standard with 0.5 ml distilled or deionized water to give a concentration described in the analysis sheet. Invert and mix gently, let sit for 5 minutes then vortex gently.
- 2. Label six tubes 1, 2, 3, 4, 5, and 6. Add 0.25 ml Assay Buffer (AB-PTR) to each of the six tubes. Prepare serial dilutions by adding 0.25 ml of the reconstituted standard to Tube 1, mix well and transfer 0.25 ml of Tube 2 to Tube 3, mix well and transfer 0.25 ml of Tube 3 to Tube 4, mix well and transfer 0.25 ml of Tube 5, mix well and transfer 0.25 ml of Tube 5 to Tube 6 and mix well.

Note: Do not use a Repeater pipette. Change tip for every dilution. Wet tip with Standard before dispensing. Unused portions of standard should be stored at ≤ -20°C. Avoid multiple freeze/thaw cycles.

# C. Standard and Quality Control Preparation

# Human Apo AV Standard Preparation (continued)

Volume of Deionized Water to Add	Volume of Standard to Add	Standard Concentration (ng/ml)
0.5 ml	0	X (refer to analysis sheet For exact concentration)

Tube #	Volume of Assay Buffer (AB-PTR) to Add	Volume of Standard to Add	Standard Concentration (ng/ml)
Tube 1	0.25 ml	0.25 ml of reconstituted standard	X/2
Tube 2	0.25 ml	0.25 ml of Tube 1	X/4
Tube 3	0.25 ml	0.25 ml of Tube 2	X/8
Tube 4	0.25 ml	0.25 ml of Tube 3	X/16
Tube 5	0.25 ml	0.25 ml of Tube 4	X/32
Tube 6	0.25 ml	0.25 ml of Tube 5	X/64

# D. Human Apo AV Quality Control 1 and 2 Preparation

Use care in opening the lyophilized Quality Control vials. Using a pipette, reconstitute each of the Human Apo AV Quality Control 1 and Quality Control 2 with 0.5 ml distilled or deionized water into the glass vials. Invert and mix gently, let sit for 5 minutes then mix well.

### X. ASSAY PROCEDURE

Pre-warm all reagents to room temperature prior to setting up the assay.

- 1. Dilute the 10X Wash Buffer concentrate 10 fold by mixing the entire content of each bottle of Wash Buffer with 450 ml deionized or distilled water. (Dilute both bottles with 900 ml deionized water).
- 2. Remove the required number of strips from the Microtiter Assay Plate. Unused strips should be resealed in the foil pouch and stored at 2-8°C. Assemble the strips in an empty plate holder and wash each well 3 times with 300 µl of 1X HRP wash buffer per wash. Decant wash buffer and remove the residual amount from all wells by inverting the plate and tapping it smartly onto absorbent towels several times. Do not let wells dry before proceeding to the next step. If an automated machine is used for the assay, follow the manufacturer's instructions for all washing steps described in this protocol.
- 3. Add in duplicate 60 µl **Assay Running Buffer (EARB-3)** to all Human Apo AV Standard, QC 1, and QC 2 wells.
- 4. Add in duplicate 80 µl Assay Running Buffer (EARB-3) to blank and sample wells.
- 5. Add in duplicate 20 µl (1:5 diluted) Matrix Solution to the Background, Standard, QC1 and QC2 wells.
- 6. Add in duplicate 20 µl Human Apo AV Standards in the order of ascending concentration to the appropriate wells. Add in duplicate 20 µl QC1 and 20 µl QC2 to the appropriate wells. Add sequentially 20µl of the (1:5 diluted) unknown samples in duplicate to the remaining wells. For best result all additions should be completed within 30 minutes.
- 7. Cover the plate with plate sealer and incubate at room temperature for 1.5 hours on an orbital microtiter plate shaker set to rotate at moderate speed, approximately 400 to 500 rpm.
- 8. Remove plate sealer and decant solutions from the plate. Tap as before to remove residual solutions in the wells.
- 9. Wash wells 3 times with 1X Wash Buffer, 300 µl per well per wash. Decant and tap firmly after each wash to remove residual buffer.
- 10. Add 100 µl of 1:1000 diluted Detection Antibody to each well. Cover plate with sealer and incubate with moderate shaking at room temperature for 1 hour on the microtiter plate shaker.

### X. ASSAY PROCEDURE (continued)

- 11. Remove plate sealer and decant solutions from the plate. Tap as before to remove residual solutions in the wells.
- 12. Wash wells 3 times with 1X Wash Buffer, 300 µl per well per wash. Decant and tap firmly after each wash to remove residual buffer.
- 13. Add 100 µl Enzyme Solution to each well. Cover plate with sealer and incubate with moderate shaking at room temperature for 30 minutes on the microtiter plate shaker.
- 14. Remove plate sealer and decant solutions from the plate. Tap as before to remove residual solutions in the wells.
- 15. Wash wells 3 times with 1X Wash Buffer, 300 µl per well per wash. Decant and tap firmly after each wash to remove residual buffer.
- 16. Add 100 µl of Substrate Solution to each well, cover plate with sealer and shake on the plate shaker for 5 to 20 minutes. Blue color should be formed in wells of Apo AV standards with intensity proportional to increasing concentrations of Apo AV.

**Note:** Please be aware that the color may develop more quickly or more slowly than the recommended incubation time depending on the localized room temperature. Please visually monitor the color development to optimize the incubation time.

17. Remove sealer and add 100 µl Stop Solution [CAUTION: CORROSIVE SOLUTION] and shake plate by hand to ensure complete mixing of solution in all wells. The blue color should turn to yellow after acidification. Read absorbance at 450 nm and 590 nm in a plate reader within 5 minutes and ensure that there are no air bubbles in any well. Record the difference of absorbance units. The absorbance of highest Apo AV standard should be approximately 2.2-2.8, or not to exceed the capability of the plate reader used.

A4, B4 ↓	G3, H3	E3, F3	C3, D3	A3, B3	G2, H2	E2, F2	C2, D2	A2, B2	G1, H1	El, Fl	C1, D1	A1, B1	VVe∥#	
,	Dilute each bottle of 10X Wash Buffer with 450ml Deionized Water.										Step 1			
		Re	emove						Wash   / on ak		nt towe	els		Step 2
80 µl	80 µl	الر 80	60 µl	60 µl	60 µl	60 µl	60 µl	60 µl	60 µl	60 µl	60 µl	80 µl	Assay Running Buffer	Step 3-4
ı	I	I	20 µl	20 µl	20 µl	20 µl	20 µl	20 µl	20 µl	20 µl	20 µl	20 µl	1:5 Matrix Solution	Step 5
20 µl of Sample	20 µl of Sample	20 µl of Sample	20 µl of QC 2	20 µl of QC 1	20 µl of reconstituted standard	20 µl of Tube 1	20 µl of Tube 2	20 µl of Tube 3	20 µl of Tube 4	20 µl of Tube 5	20 µl of Tube 6		Standards/ Controls/ Samples	Step 6
			Seal,						oom To h Buffe	emperc er	ature.			Step 7-9
•												100 µl	1:1000 Diluted Detection Antibody	Step 10
			Sea						om Ter h Buffe	nperati er	ıre.			Step 11-12
•												100 µl	Enzyme Solution	Step 13
			Seal,						Room T h Buffe	emper	ature.			Step 14-15
-												100 μΙ	Substrate	Step 16
		S	eal, A	gitate,	Incuba	ite 5 -	20 mir	nutes a	t Room	Тетр	erature	-	1	Step 17
-												100 μΙ	Stop Solution	Step 18
			Read	Absor	bance	at 450	O nm a	nd 59	O nm.					Step 19

# XI. MICROTITER PLATE ARRANGEMENT

# Human Apo AV ELISA

	]	2	3	4	5	6	7	8	9	10	]]	12
А	Blank	Tube 3	QC 1	Etc.								
В	Blank	Tube 3	QC 1	Etc.								
С	Tube 6	Tube 2	QC2									
D	Tube 6	Tube 2	QC2									
Е	Tube 5	Tube 1	Sample									
F	Tube 5	Tube 1	Sample									
G	Tube 4	Reconstituted Standard	Sample									
Н	Tube4	Reconstituted Standard	Sample									

#### XII. CALCULATIONS

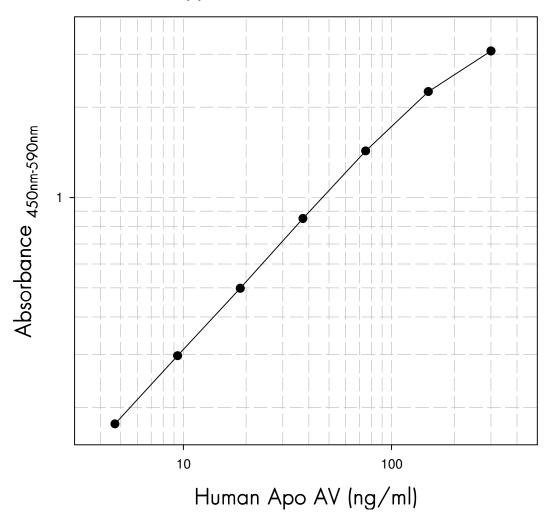
The dose-response curve of this assay fits best to a sigmoidal 4- or 5-parameter logistic equation. The results of unknown samples can be calculated with any computer program having a 4- or 5-parameter logistic function.

Note: When sample volumes assayed differ from 20  $\mu$ l, an appropriate mathematical adjustment must be made to accommodate for the dilution factor (e.g., if 10  $\mu$ l of sample is used, then calculated data must be multiplied by 2). When sample volume assayed is less than 20  $\mu$ l, compensate the volume deficit with assay buffer (sample diluent).

#### XIII. INTERPRETATION

- 1. The assay will be considered accepted when all Quality Control values fall within the calculated Quality Control Range. If any QC's fall outside the control range, review results with a supervisor.
- 2. If the difference between duplicate results of a sample is >15% CV, repeat the sample.
- 3. The limit of sensitivity of this assay is 1.0 ng/ml Human Apo AV (20 µl sample size).
- 4. The appropriate range of this assay is 4.96 ng/ml to 300 ng/ml Human Apo AV (20 μl sample size). Any result greater than 300 ng/ml in a 20 μl sample should be diluted using assay diluent, and the assay repeated until the results fall within range.

# Human Apo AV ELISA Assay Typical Standard Curve



Typical Standard Curve, not to be used to calculate data.

### XV. ASSAY CHARACTERISTICS

### A. Sensitivity

The lowest level of Apo AV that can be detected by this assay is 1.0 ng/ml when using a 20  $\mu l$  sample size.

## B. Specificity

The antibody pair used in this assay is specific to Human Apo AV. No species cross-reactivity is observed in the assay to rat, mouse, canine, porcine, or sheep samples. However, there is a high cross-reactivity to non-human primate samples that has not been evaluated or calibrated.

No cross reactivity is observed to the following human analytes: Apo A1, Apo AII, Apo B, Apo CII, Apo CIII, Apo E, and other human cytokines and endocrine hormones tested.

### C. Precision

### Intra-Assay Variation

mar way to		
Sample No.	Mean Apo AV Levels	Intra-Assay% CV
	(ng/ml)	
1	5.8	6.44
2	6.53	6.18
3	7.2	3.57
4	7.36	2.88
5	8.06	5.33
6	12.18	3.29
7	12.96	2.83
8	13.34	3.78

### Inter-Assay Variation

Sample No.	Mean Apo AV Levels	Inter-Assay % CV
	(ng/ml)	
1	7.85	12.36
2	9.17	8.15
3	9.62	5.36
4	10.20	10.07
5	15.47	6.00
6	16.82	6.53
7	17.4	5.59
8	21.92	5.56

The assay variations of Human Apo AV ELISA kits were studied on eight human serum samples with varying concentrations of endogenous Apo AV. The mean Intra-Assay variation was calculated from results of 10 replicate determinations in each assay of the indicated samples. The mean Inter-Assay variations of each sample were calculated from results of three separate assays with duplicate samples in each assay.

# XV. ASSAY CHARACTERISTICS (continued)

D. RecoverySpike & Recovery of Human Apo AV in Serum

Sample No. 1	Apo AV Added ng/ml 0 4.69 37.5 150	Expected ng/ml 20.3 25.0 57.8 170.3	Observed ng/ml 20.3 25.1 47.2 140.0	% of Recovery 100 82 82
2	0 4.69 37.5 150	26.4 31.1 63.9 176.4	26.4 31.2 59.3 159.1	100 93 90
3	0 4.69 37.5 150	30.5 35.2 68.0 180.5	30.5 34.6 61.4 173.5	98 90 96
4	0 4.69 37.5 150	35.0 39.7 72.5 185.0	35.0 39.4 64.5 144.7	99 89 78
5	0 4.69 37.5 150	43.6 48.29 81.1 193.6	43.6 50.5 78 177.8	104.58 96.18 91.84
6	0 4.69 37.5 150	89.7 94.39 127.2 239.7	89.7 93.7 125.4 223.1	99.27 98.58 93.07

Varying amounts of Human Apo AV were added to eight human serum samples and the Apo AV content was determined in three separate assays. The % of recovery = observed Apo AV concentrations/expected Apo AV concentrations x 100%.

# XV. ASSAY CHARACTERISTICS (continued)

E. Linearity

Effect of Serum Dilution

Sample	Volume	Expected	Observed	% Of
No.	Sampled	ng/ml	ng/ml	Expected
1	20μl 10μl 5μl 2.5μl	19.60 9.80 4.90 2.45	19.6 9.1 4.6 2.3	92.86 93.88 93.88
2	20µl 10µl 5µl 2.5µl	36.50 18.25 9.13 4.56	36.5 16.2 8.1 3.9	88.77 88.77 85.48
3	20µl 10µl 5µl 2.5µl	39.30 19.65 9.83 4.91	39.3 17.8 8.8 4.2	90.59 89.57 85.50
4	20μl 10μl 5μl 2.5μl	39.80 19.90 9.95 4.98	39.8 21 10 5.8	105.53 100.50 116.58
5	20µl 10µl 5µl 2.5µl	48.00 24.00 12.00 6.00	48 23.9 12.9 6.3	99.58 107.50 105.00
6	20μΙ 10μΙ 5μΙ 2.5μΙ	66.40 33.20 16.60 8.30	66.4 32.1 15.9 7.9	96.69 95.78 95.18

Six human serum samples with the indicated sample volumes were assayed in three separate experiments. Required amounts of assay buffer were added to compensate for lost volumes below 20  $\mu$ l. The resulting dilution factors of 1, 2, 4, and 8 representing 20  $\mu$ l, 10  $\mu$ l, 5  $\mu$ l, and 2.5  $\mu$ l sample volumes assayed, respectively, were applied in the calculation of % expected = observed/expected x 100%.

### XVI. TROUBLESHOOTING GUIDE

- 1. To obtain reliable and reproducible results the operator should carefully read this manual and fully understand all aspects of each assay step before attempting to run the assay.
- 2. Throughout the assay the operator should adhere strictly to the procedures with good laboratory practice.
- 3. Have all necessary reagents and equipment ready on hand before starting. Once the assay has been started all steps should be completed with precise timing and without interruption.
- 4. Avoid cross contamination of any reagents or samples to be used in the assay.
- 5. Make sure all reagents and samples are added to the bottom of each well.
- 6. Careful and complete mixing of solutions in the well is critical. Poor assay precision will result from incomplete mixing or cross well contamination due to inappropriate mixing.
- 7. Remove any air bubble formed in the well after acidification of substrate solution because bubbles interfere with spectrophotometric readings.
- 8. Do not let the absorbency reading of the highest standard reach 3.0 units or higher after acidification.
- 9. High absorbance in background or blank wells could be due to 1) cross well contamination by standard solution or sample or 2) inadequate washing of wells with HRP Wash Buffer or 3) overexposure to light after substrate has been added.

### XVII. REPLACEMENT REAGENTS

Reagents	Cat. #
Microtiter Plate	EP71
10X HRP Wash Buffer Concentrate (50 ml)	EWB-HRP
Human Apo AV Standard	E8071-K
Quality Controls 1 and 2	E6071-K
Matrix Solution	<b>EMTX</b>
Assay Buffer (Sample Diluent)	AB-PTR
Assay Running Buffer	EARB-3
Human Apo AV Detection Antibody (1000X)	E1071
Enzyme Solution	EHRP-3
Substrate	ESS-TMB
Stop Solution	ET-TMB

### XVIII. ORDERING INFORMATION

# A. To place an order:

### For USA Customers:

Please provide the following information to our customer service department to expedite your telephone, fax or mail order:

- 1. Your name, telephone and/or fax number
- 2. Customer account number
- 3. Shipping and billing address
- 4. Purchase order number
- 5. Catalog number and description of product
- 6. Quantity and product size

TELEPHONE ORDERS:

Toll Free US: (800) MILLIPORE

FAX ORDERS: (636) 441-8050

MAIL ORDERS: Millipore

6 Research Park Drive

St. Charles, Missouri 63304 U.S.A.

#### For International Customers:

To best serve our international customers, it is Millipore's policy to sell our products through a network of distributors. To place an order or to obtain additional information about Millipore products, please contact your local distributor.

### B. Conditions of Sale

All products are for research or manufacturing use only. They are not intended for use in clinical diagnosis or for administration to human or animals. All products are intended for *in vitro* use only.

# C. Material Safety Data Sheets (MSDS)

Material safety data sheets for Millipore products may be ordered by fax or phone. See Section A above for details on ordering.