Human Amyloid β42 Brain
96-Well Plate
Cat. # EZBRAIN42

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96-Well Plate (Cat. # EZBRAIN42)

Millipore • 6 Research Park Drive • St. Charles, Missouri 63304 USA
Phone: (800) MILLIPORE • Fax: 636-441-8050
techserv_dd@millipore.com • www.millipore.com/bmia

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I. INTENDED USE

Amyloid beta peptides have been implicated in the etiology of Alzheimer's disease. Amyloid beta 40 is the most prominent peptide and Amyloid beta 42 is the neurotoxic form. The Amyloid beta 42/40-ratio (AB ratio) has been reported as a better indicator of the Alzheimer pathology. Millipore's Human Amyloid β42 Brain ELISA kit is used for the measurement of Amyloid beta 42 in brain samples and for other tissue samples in a 96-well format. This assay is for research use only and appropriate for the *in vitro* detection of human Amyloid β42 peptides in brain samples from e.g. Guinea pig, transgenic hAmyloid mice and cell extracts.

This kit is for research purpose only.

II. PRINCIPLES OF PROCEDURE

Prior to the ELISA procedure the brain samples are homogenized in the presence of the Lysis buffer provided in this kit. The homogenate is cleared by centrifugation and the supernatant is processed in the enzymatic assay. The ELISA applies a polystyrene, 96-well microtiter plate (12 flexible strips with 8 wells) coated with a monoclonal antibody (capture antibody) highly specific for the human Amyloid β42 peptide (antigen). During a first incubation, the antigen in the sample is selectively bound through its C-terminal end. In the same step a biotin conjugated antibody (detection antibody) binds to the N-terminal epitope of the immobilized antigen. After washing steps a streptavidin-peroxidase-conjugate is added to the wells. This leads to the formation of a complex of the antibody-Amyloid-antibody sandwich structure and the peroxidase linked over a streptavidin-biotin bridge. The addition of a substrate solution results in the catalysis of an enzymatic reaction with a colored product and the color intensity is measured by a spectrophotometer. The Amyloid β42 concentration in unknown samples are calculated from the standard curve.

Characteristics of the Aβ42 Brain ELISA:

- Highly sensitive, selective quantitative analysis of human Amyloid β42
- Test range from 16 to 500 pg/mL
- High reproducibility and accurate linearity of the standard curve
- Precoated strips (12x8) for flexible usage of samples according to individual customer requirements
- Low sample volumes (50 µL or less)

III. REAGENTS SUPPLIED

Each kit is sufficient to run one 96-well plate and contains the following reagents:

A. Human Amyloid β 1-42 ELISA Plate

Antibody-coated microtiter plate
Quantity: 1 strip plate
Preparation: Ready to Use
Note: Unused strips should be resealed in the foil pouch with the desiccant provided and stored at 2-8°C.

B. Adhesive Plate Sealer

Quantity: 2 sheets
Preparation: Ready to Use

C. Synthetic Amyloid β1-42 Standard

Concentration: 1000 ng/mL, lyophilized
Quantity: 1 bottle containing 250 µL / vial upon hydration.
Preparation: Reconstitute vial with 0.25 mL distilled or deionized water.

D. Amyloid β1-42 Quality Controls 1 and 2

One vial each, lyophilized, containing amyloid β1-42 at two different levels.
Quantity: 0.25 ml/vial upon hydration.
Preparation: Reconstitute each vial with 0.25 ml de-ionized water immediately before use.

E. Standard & Sample Diluent

Quantity: 1 bottle containing 25mL
Preparation: Ready to use, for dilution of standards or samples

F. Antibody Conjugate (100X)

Quantity: 1 bottle containing 100 µL at 100 fold concentrate
Preparation: Dilute with Antibody Conjugate Diluent 1:100 before use

G. Antibody Conjugate Diluent

Quantity: 1 bottle containing 8 mL
Preparation: Ready to Use, for use in diluting the Antibody Conjugate.

H. Enzyme Conjugate (100X) (Steptavidin-Peroxidase-Conjugate)

Quantity: 1 bottle containing 150 µL at 100 fold concentrate
Preparation: Dilute with Enzyme Conjugate Diluent 1:100 before use

I. Enzyme Conjugate Diluent

Pre-titered Streptavidin-Horseradish Peroxidase Conjugate in Buffer
Quantity: 1 bottle containing 13 mL
Preparation: Ready to Use, for dilution of Enzyme Conjugate

J. Washing Solution (20X)

Quantity: 2 bottles containing 25 mL/bottle at 20 fold concentrate
Preparation: Dilute 1:20 with deionized water before use

K. Substrate Solution

Quantity: 1 bottle containing 13 mL
Preparation: Ready to Use

L. Stop Solution

0.3 M HCl
Quantity: 1 bottle containing 12 mL
Preparation: Ready to Use **(Caution: Corrosive Solution)**

M. Lysis Buffer

Quantity: 1 bottle containing 40 mL
Preparation: Ready to Use

N. Protease Inhibitor Cocktail Tablets

1 tablet sufficient for 5 mL Lysis buffer
Quantity: 8 Tablets
Preparation: Ready to Use

IV. STORAGE & STABILITY

Prior to use, all components in the kit can be stored up to 2 weeks at 2-8°C. For longer storage (> 2 weeks), freeze Wash Buffer and reconstituted Standards and Quality Controls at ≤ –20°C. Minimize repeated freeze and thaw of the Standards and Quality Controls. Refer to expiration dates on all reagents prior to use. Do not mix reagents from different kits unless they have the same lot numbers

V. TECHNICAL GUIDELINES

- The hAmyloid β42-Brain ELISA is for *in vitro* use only.
- Carefully read and follow the test-instructions in this user guide included in every test-kit. Test performance and data calculation should always be done by qualified staff.
- Do not mix reagents from different test-kits.
- Some of the test components are concentrated solutions. After dilution the working solution should be used within 14 days (2 to 8 °C). Standard dilutions have to be diluted always just before the test starts.
- To calibrate the test-system (standard), the dilutions should be made according to the description in the test procedure. The resulting internal standard curve is a fixed component of each measurement. A transfer of the absorbance data from one test plate to another is not suitable.
- To avoid a cross contamination and carryover of reagents, the use of clean pipet tips for each sample pipetting is necessary.
- The pipetting of reagents and samples starts / stops kinetic reactions. To obtain a high precision for the test, be sure to treat each well of the microtiter plate in an identical manner.
- The washing solution has to be tapped out of the wells after the last washing step to assure the removal of buffer residues from the wells completely.

VI. REAGENT PRECAUTIONS

A. Hydrochloric Acid

Hydrochloric Acid is corrosive and can cause eye and skin burns. It is harmful if swallowed and can cause respiratory and digestive tract burns. Avoid contact with skin and eyes. Do not swallow or ingest.

VII. MATERIALS REQUIRED BUT NOT PROVIDED

In addition to the reagents provided in the test-kit, the following materials are essential for the performance of the peptide extraction and the ELISA-Test:

- Pre-cooled 70% formic acid (for analysis of insoluble Aβ)
- Tris pH 7.4 (for analysis of insoluble Aβ)
- Dry ice or liquid nitrogen
- Protease Inhibitor Tablets
- Pre-weighed 2 mL sample tubes (one tube per brain tissue sample)
- Homogenizer (e.g. hand-held motor homogenizer or 1 mL Dounce homogenizer)
- Phosphate-Buffered-Saline (PBS)
- Deionized water for dilution of Washing Solution
- Variable precision pipets (suitable for volumes from 10 to 1000 µL) *
- Vortex-mixer
- Timer
- Microtiter plate shaker
- Microtiter plate washer *
- Microtiter Plate Reader capable of reading absorbancy @ 450 nm
- Ice and ice container for sample preparation

* We recommend the use of multi channel pipets and automated plate-washers to achieve parallel working steps and simultaneous incubation times for best reproducibility.

VIII. SAMPLE COLLECTION AND STORAGE

- We recommend diluting the working solutions only for the intended use. The test plate is subdivided in strips of 8 wells for flexible sample handling

- The stability of Amyloid β is critical, because the peptides tend to aggregate in samples. For this reason, the preanalytic sample preparation is a major influencing parameter within the analysis of Amyloid peptides. Samples should be collected according to clinical approved standard procedures and immediately stored at -20 °C. During the handling of thawed samples, it is important to keep these chilled (for example working on ice).

- For the preparation of the samples polypropylene vials are recommended to avoid interaction with sample materials during storage.

- For research use only the test can be applied directly or in dilution to different sample materials (brain tissue extracts, cell extracts). More complex sample materials sometimes require a change of the test procedure.

- Samples should be collected in pyrogen/endotoxin-free tubes.

- Samples should be frozen if not analyzed shortly after collection.

- Dilution requirements for brain tissue vary. End user should determine optimum dilution for their samples

- Avoid multiple freeze-thaw cycles of the frozen samples.

- Ensure that samples are free of particulate matter. If needed centrifuge or filter prior to analysis.

Brain Lysate Sample Preparation

We recommend keeping the brain samples frozen until Lysis buffer is added. The brain sample sizes should range between 50 to 90 mg. Clean the homogenizer thoroughly with deionized water after each homogenization.

1. Keeping on ice, dissolve protease inhibitor cocktail tablets in cold lysis buffer (1 tablet per 5 mL lysis buffer).

2. Mark and weigh appropriate number of micro tubes for brain samples to be prepared.

3. Pre chill micro tubes on dry ice.

4. Determine the exact wet mass of brain samples in the pre-weighed 2 mL sample tubes and transfer sample tubes on dry ice immediately thereafter.

5. Add 10x volume of cold Lysis buffer to the frozen brain sample immediately before the homogenization.

6. Homogenize the sample in the tube by using a hand-held homogenizer. Optionally, add brain sample and 10x volume of Lysis buffer to a Dounce homogenizer and homogenize thoroughly.

7. Rotate the homogenate for 2 hours at 4 °C.

8. Clear the lysate by centrifugation at 13000 rpm for 10 minutes at 4 °C.

9. For soluble Amyloid beta:
    a. Transfer supernatant to a new tube. No pellet should occur following a control centrifugation of the supernatant.
    b. Keep the lysate on ice.

10. For insoluble Amyloid beta:
    a. Resuspend the pellet in 2x volume of 70% pre-cooled formic acid.

b. Sonicate for >10 min and neutralize with 15x volume 1M Tris pH 7.4.

c. Centrifuge at 13000 rpm for 10 minutes at 4°C.

d. Transfer supernatant to a new tube.

e. Keep lysate on ice.

IX. REAGENT PREPARATION

A. Preparation of Human Amyloid β 1-42 Standard (DAY 1):

1. Use care in opening the lyophilized Standard vial. Using a pipette, reconstitute the Human Amyloid β 1-42 Standard with 0.25 ml distilled or deionized water to give a concentration of 1000 ng/ml. Invert and mix gently, let sit for 5 minutes then vortex gently.

2. Label eight tubes as Stock (S), 500 pg/ml, 250 pg/ml, 125 pg/ml, 62.5 pg/ml, 31.25 pg/ml, 16 pg/ml, and Blank. Using the chart below, add appropriate volumes of Standard and Sample Diluent to each of the eight tubes. Prepare dilutions according to the chart below and mix well.

   Note: Change tip for every dilution. Wet tip with Standard before dispensing. Unused portions of standard should be stored at ≤ -20°C. Avoid multiple freeze/thaw cycles.

Volume of Deionized Water to Add	Volume of Standard to Add	Standard Concentration (ng/ml)
0.25 ml	0	1000 ng/ml

Standard	Concentration of Amyloid β 1-42	Volume of Standard and Sample Diluent to Add	Volume of Standard to Add
Stock (S)	25,000 pg/ml	780 µl	20 µl of reconstituted std
Std 1	500 pg/ml	1470 µl	30 µl of Stock (S)
Std 2	250 pg/ml	150 µl	150 µl Std 1
Std 3	125 pg/ml	150 µl	150 µl Std 2
Std 4	62.5 pg/ml	150 µl	150 µl Std 3
Std 5	31.25 pg/ml	150 µl	150 µl Std 4
Std 6	16 pg/ml	150 µl	150 µl Std 5
Blank	0 pg/ml	150 µl	0

B. Quality Control 1 and 2 Preparation

Use care in opening the lyophilized Quality Control vials. Reconstitute each Amyloid β 1-42 Quality Control 1 and Quality Control 2 with 0.25 ml distilled or de-ionized water and gently invert to ensure complete hydration. Unused portions of the reconstituted Quality Controls should be stored in small aliquots at -20°C. Avoid further freeze/thaw cycles.

C. Preparation of Antibody Conjugate Solution (Day 1):

Dilute Antibody Conjugate (100x) 1:100 with Antibody Conjugate Diluent.

Example: 60 µL Antibody Conjugate (100x) + 5940 µL Antibody Conjugate Diluent = 6000 µL

D. Preparation of Washing Solution (Day 2) :

Dilute Washing Solution (20x) 1:20 with deionized water

Example: 50 mL Washing Solution (20x) + 950 mL deionized water = 1000 mL

E Preparation of Enzyme Conjugate Solution (Day 2):

Dilute Enzyme Conjugate (100x) 1:100 with Enzyme Conjugate Diluent

Example: 110 µL Enzyme Conjugate (100x) + 10890 µL Enzyme Conjugate = 11000 µL

X. ASSAY PROCEDURE

Day 1

- The following kit components are required for day 1:

  Lysis Buffer
  70% formic acid (not provided in the kit)
  8 well test strips
  Standard & Sample Diluent
  Synthetic Aß1-42 Standard
  Aß 1-42 Quality Controls 1,2
  Protease Inhibitor Tablets
  1M Tris pH7.4 (not provided in the kit)
  Antibody Conjugate Diluent
  Antibody Conjugate (100x)

- We recommend diluting the test reagents just before each application. The samples shall be chilled (at <4 °C, working on ice) during the complete test procedure to achieve high stability and optimal data results.
- All standards, quality controls or samples should be mixed gently just before pipetting. Accurate mixing and pipetting of the standard and quality control solutions are essential to the precision of the assay.

Note: If low values of Amyloid β are expected, a pre incubation step could be applied as follows: Add 100 µL of the brain homogenate sample to the well and incubate for 5 minutes on the micro plate shaker at room temperature. Incubate for another 60 minutes on 4 °C without shaking. Remove supernatant and proceed as below (starting with step 1).

1. Add 50 µL Antibody Conjugate Solution into all wells.
2. Add 50 µL of Standard and Sample Diluent to the background (0 pg/ml) wells.
3. Add in duplicate 50 µL standard in order of ascending concentration to the appropriate wells. Add sequentially 50 µL of samples in duplicate to the remaining wells. **For best results all additions should be completed within 30 minutes**.
4. Cover the plate with a plate sealer and thoroughly mix the contents of the wells for a period of 5 minutes on an orbital plate shaker (500-600 rpm/min). Incubate without shaking overnight (16-20 hours) at 2 to 8 °C.

Note: The test reagents needed on the following day can be taken out of the refrigerator to allow them to reach room temperature overnight.

Day 2

- The following kit components are required for day 2:

  Washing Solution (20x)
  Enzyme Conjugate (100x)
  Enzyme Conjugate Diluent
  Substrate Solution
  Stop Solution

- Caution- All reagents must be at room temperature before use.

1. Remove plate sealer and decant solutions from the plate. Tap as before to remove residual solutions in the wells.
2. Wash test plate 5 times with 300 µL Washing Solution per well, remove the remaining fluid by tapping the plate on an absorbing paper.
3. Add 100 µL Enzyme Conjugate Solution to each well, cover the plate with a plate sealer and incubate for 30 minutes at room temperature (20 to 28 ℃) on an orbital shaker (500-600 rpm/min).
4. Remove plate sealer and decant solutions from the plate. Tap as before to remove residual solutions in the wells.
5. Wash test plate 5 times with 300 µL Washing Solution per well, remove the remaining fluid by tapping the plate on an absorbing paper.
6. Add 100 µL Substrate Solution to each well. Cover plate with sealer and shake on the plate shaker for 5 - 30 minutes (A longer development time may be needed if using a plate washer). Blue color should be formed in wells of standards with intensity proportional to increasing concentrations of Amyloid β 1-42.

   NOTE: Please be aware that the color may develop more quickly or more slowly than the recommended incubation time depending on the localized room temperature. Please visually monitor the color development to optimize the incubation time.

7. Remove sealer and add 100 µL Stop Solution **(Caution: Corrosive solution)** and shake plate by hand to ensure complete mixing of solution in all wells. The blue color should turn to yellow after acidification. Read absorbance at 450 nm and 590 nm in a plate reader within 5 minutes and ensure that there are no air bubbles in any well. Record the difference in absorbance units. The absorbance of the highest Amyloid β 1-42 standard should be approximately 2.0 – 3.2, or not to exceed the capability of the plate reader used.

Assay Procedure

	Day 1				Day 2						
	Step 1	Step 2	Step 3	Step 4	Step 1-2	Step 3	Step 4-5	Step 6		Step 7	Step 7
Well #	Ab. Conj Solution	Standard & Sample Diluent	Standards/QCs & Samples	Seal, agitate for 5 minutes, then incubate overnight at 2 to 8 ℃.	Wash test plate 5 times with 300 µL Washing Solution per well, remove the remaining fluid by tapping the plate on an absorbing paper.	Enzyme Conj Solution	Seal., agitate, and incubate at room temperature for 30 minutes Remove residual buffer by tapping smartly on absorbent towels. Wash 5X with 300 µL Wash Buffer Remove the remaining fluid by tapping the plate on an absorbing paper.	Substrate	Seal, agitate and incubate 5 to 30 min at room temperature.	Stop Solution	Read Absorbance at 450 nm and 590 nm.
A1, B1	50 µL	50 µL	----			100 µL		100 µL		100 µL	
C1, D1	50 µL	----	50 µl of 16 pg/mL Standard								
E1, F1	50 µL	----	50 µl of 31.25 pg/mL Standard								
G1, H1	50 µL	----	50 µl of 62.5 pg/mL Standard								
A2, B2	50 µL	----	50 µl of 125 pg/mL Standard								
C2, D2	50 µL	----	50 µl of 250 pg/mL Standard								
E2, F2	50 µL	----	50 µl of 500 pg/mL Standard								
G2, H2	50 µL	----	50 µl of Quality Control 1								
A3, B3	50 µL	----	50 µl of Quality Control 2								
C3, D3	50 µL	----	50 µl of Sample								
E3, F3	50 µL	----	50 µl of Sample								
G3, H3 ↓	50 µL	----	50 µl of Sample								

XI. MICROTITER PLATE ARRANGEMENT

	1	2	3	4	5	6	7	8	9	10	11	12
A	(Blank) 0 pg/mL	125 pg/mL	QC 2	Etc.								
B	(Blank) 0 pg/mL	125 pg/mL	QC 2									
C	16 pg/mL	250 pg/mL	Sample 1									
D	16 pg/mL	250 pg/mL	Sample 1									
E	31.25 pg/mL	500 pg/mL	Sample 2									
F	31.25 pg/mL	500 pg/mL	Sample 2									
G	62.5 pg/mL	QC 1	Sample 3									
H	62.5 pg/mL	QC 1	Sample 3									

XII. INTERPRETATION

Analysis of the measured absorbance data (mean, standard deviation) for the standards and for the samples is performed with the help of a microtiter plate reader software.

The **blank** (zero standards) is **not** integrated into the calculation of the standard curve. The blank is taken only as a control for a non-specific binding of the antibody; the mean absorbance of the blank shall be below 0.2.

Construct a standard curve by plotting the mean absorbance of standard 1-6 on the vertical axis versus the corresponding Aβ1-42 concentration on the horizontal axis. The data can be calculated by linear fit (linear regression) or by a point to point fit (cubic spline). The test results are not valid if the standard 1 (500 pg/mL) shows an absorbance below 0.6 in magnitude. Please control your test handling (XIV. TROUBLESHOOTING GUIDE).

The assay will be considered accepted when all Quality Control values fall within the calculated QC range. If any QCs fall outside of thecontrol range, review results with a supervisor.

The resulting Aβ42-concentrations of the samples can be calculated with this standard curve. Only samples that are in the measured range of the standard curve can be calculated.
If the Aβ-concentration value of a sample exceeds 500 pg/mL, the test sample must be measured again by using a higher sample dilution (with appropriate amount of Standard & Sample Diluent)**.**

XIII. ASSAY CHARACTERISTICS

A. Sensitivity

The lowest level of Amyloid ß1-42 standard used in this assay is 8 pg/mL (50 µL sample size).

B. Specificity

The Amyloid β42 ELISA uses monoclonal anti-Aβ antibodies with high selectivity for human Aβ. The capture antibody recognizes the C-terminal end of Amyloid β1-42, which causes a high selectivity for Aβ42. The cross-reactivity of the used antibodies to other Amyloid peptides was tested by ELISA and BIACORE and shows no significant cross-reactivity to Aβ1-38, Aβ1-39, Aβ1-42, Aβ1-43 and Aβ1-44.

C. Precision

Analyte	Intra-Assay (%CV)	Inter-Assay (% CV)
Amyloid β 1-42	<10%	<10%

XIV. QUALITY CONTROLS

The ranges for Quality Control 1 and 2 are provided on the card insert or can be located at the Millipore website www.millipore.com\bmia.

XV. TROUBLESHOOTING GUIDE

Problem	Cause	Recommend
No signal	wrong test reagents used	Ensure that only the reagents for the specific test-lot are used.
	test reagents damaged	Don't use the test-kit after expiration date.
Weak signal	test reagents used in a wrong dilution	Control used test dilutions carefully (usually a dilution factor of 100 is used).
	wrong filter (wavelength)	Check your wavelength in your microtiter plate photometer.
	incubation time too short temperature too low	Check the information of incubation times of the lot in the product data sheet. (The incubation time of the enzyme substrate is applied for temperatures from 20 to 28 °C!); extend the substrate incubation time, if absorption is below 1.0
	reagents not at right temperature	Make sure that the reagents used for day 2 have reached room temperature (20 to 28 °C) before using within the test-kit.
	sodium azide, mercaptoethanol or DTT can interfere with peroxidase activity at high concentrations	Only use samples which contain no or low contents (< 0.1 %) of sodium azide, mercaptoethanol or DTT.
High signal	test reagents used in a wrong dilution	Check used test dilutions carefully (usually a dilution factor of 100 is used).
	incubation time too long temperature too high	Check the information of incubation times of the lot in the product data sheet. (The incubation time of the enzyme substrate is applied for temperatures from 20 to 28°C!); shorten the substrate incubation time, if absorption is above 3.0

Problem	Cause	Recommend
High background (blank)	insufficient washing steps	Wash plate carefully and remove the liquid after each washing carefully.
	contamination of the washing solution	Confirm that the water is not contaminated. Use always double distilled water for the reconstitution and dilution of the washing solution.
	contamination of reagents or vials/tubes from previous experiments	Avoid pipetting directly out of the reagent vials, if test reagents should be used in further measurements. (Oxidative active contaminants can influence the enzyme substrate by non-specific color development).
	test reagents (antibody- and enzyme conjugate) used in wrong dilutions	Check used test dilutions for antibody- and enzyme conjugate carefully (usually a dilution factor of 100 is used).
Low precision (= random error)	non-homogeneous samples e.g. cloudy solution, particles in the sample	Check that the samples are taken, prepared and stored according to a recommended sample procedure (polypropylene tubes, storage of clear samples at -20 °C).
	insufficient mixing of samples and standards	Mix samples and standards before pipetting carefully.
	variation in pipetting	Check your pipettes and calibrate if necessary.
	carry over between samples and/or standards	Change pipet tips after each pipetting.
	insufficient mixing of reagents during incubation	Mix reagents on the test plate after pipetting by moving the test plate carefully; use an orbital microtiter plate shaker on the recommended test steps for optimal mixing of reagents.
	insufficient washing	Check that the automatic microtiter plate washer is working correctly; residues of liquids must be removed completely after each washing step.
	evaporation of liquids	Check the contact of the cover seal with the plate during the incubation steps.

Problem	Cause	Recommend
Calculated data are too high or too low (=systematic error, deviation of data from „typical data")	calculation of the dilution factor is not correct	Check the dilution factor used for the sample dilution within the data calculation.
	modification of the test procedure	Follow the instructions in the product data sheet carefully (incubation time, dilution etc.).
	incorrect sample treatment	Check that the samples are taken, prepared and stored according to a recommended sample procedure (polypropylene tubes, storage of clear samples at -20 °C).

XVI. REPLACEMENT REAGENTS

Reagents	Cat. #
Human Amyloid β1-42 ELISA Plate	4TS
Lysis Buffer	LB
Protease Inhibitor Cocktail Tablets	PICT
Synthetic Aβ 1-42 Standard	2STM
Amyloid β 1-42 Quality Control 1,2	2QC
Standard & Sample Diluent	SD
Antibody Conjugate	HSAC
Antibody Conjugate Diluent	HSAD
Enzyme Conjugate	2EC
Enzyme Conjugate Diluent	2ED
Washing Solution	WS
Substrate Solution	ES
Stop Solution	ET-TMB

XVII. ORDERING INFORMATION

A. To place an order:

For USA Customers:

Please provide the following information to our customer service department to expedite your telephone, fax or mail order:

1. Your name, telephone and/or fax number
2. Customer account number
3. Shipping and billing address
4. Purchase order number
5. Catalog number and description of product
6. Quantity and product size

TELEPHONE ORDERS:
Toll Free US (800) MILLIPORE
FAX ORDERS: (636) 441-8050
MAIL ORDERS: Millipore
6 Research Park Drive
St. Charles, Missouri 63304 U.S.A.

For International Customers:
To best serve our international customers, it is Millipore's policy to sell our products through a network of distributors. To place an order or to obtain additional information about Millipore products, please contact your local distributor.

B. Conditions of Sale

All products are for research or manufacturing use only. They are not intended for use in clinical diagnosis or for administration to human or animals. All products are intended for *in vitro* use only.

C. Material Safety Data Sheets (MSDS)

Material safety data sheets for Millipore products may be ordered by fax or phone. See Section A above for details on ordering.

XVIII. REFERENCES

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