

**THIS KIT IS INTENDED FOR RESEARCH USE ONLY.**

**THIS KIT IS NOT INTENDED FOR DIAGNOSTIC PURPOSES.**

## 1 INTENDED USE

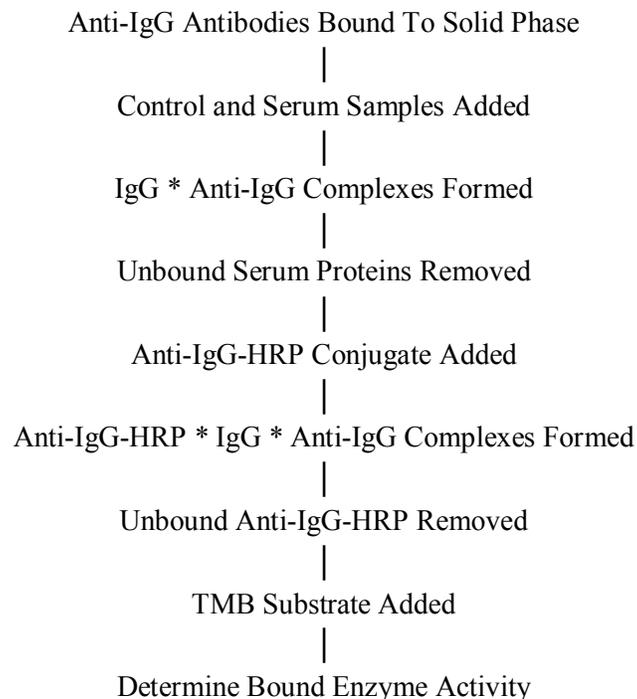
The Mouse IgG ELISA is a highly sensitive two-site enzyme-linked immunoassay (ELISA) for determination of IgG in mouse biological samples.

**For research use only.**

## 2 PRINCIPLE

The principle of the double antibody sandwich ELISA is represented in Figure 1. In this assay the IgG present in the serum sample reacts with the anti-IgG antibody which has been adsorbed to the surface of polystyrene microtiter wells. After the removal of unbound serum proteins by washing, anti-IgG antibody conjugated with horseradish peroxidase (HRP) is added. This HRP-conjugated antibody forms a complex with the previously bound serum IgG. Following another washing step, the enzyme bound to the immunosorbent is assayed by the addition of a chromogenic substrate, 3,3',5,5'-tetramethylbenzidine (TMB). The quantity of bound enzyme is proportional to the concentration of IgG in the sample tested; thus, the absorbance, at 450 nm, is a measure of the concentration of IgG in the test sample. The quantity of IgG in the test sample can be interpolated from the calibration curve constructed from the calibrators, and corrected for serum dilution.

*Figure 1.*



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**For Research Use Only****3 COMPONENTS****1. Diluent Concentrate**

One bottle containing 50 mL of a 5X concentrated diluent running buffer.

**2. Wash Solution Concentrate**

One bottle containing 50 mL of a 20X concentrated wash solution.

**3. Enzyme-Antibody Conjugate Concentrate**

One vial containing 150µL of a 100X concentrated affinity-purified anti-mouse IgG antibody conjugated with HRP in a stabilizing buffer.

**4. TMB Substrate Solution**

One vial containing 12 mL of TMB and hydrogen peroxide in citric acid buffer at pH 3.3.

**5. Stop Solution**

One vial containing 12 mL of 0.3 M sulfuric acid.

WARNING: Avoid contact with skin.

**6. Microtiter Plate**

Twelve removable eight-well strips in well holder frame.

Wells are coated with affinity-purified anti-mouse IgG.

**7. Mouse IgG Calibrator**

One vial containing 0.1 mL with 2.55 mg/mL of Mouse IgG Calibrator.

**8. Positive Control**

One vial containing 50 µL of serum with 0.1% sodium azide.

See the Control Certificate for the concentration.

**4 MATERIALS REQUIRED BUT NOT PROVIDED**

- Precision pipettes (2 µL to 200 µL) for making and dispensing dilutions.
- Test tubes
- Microplate washer/aspirator
- Distilled or de-ionized H<sub>2</sub>O
- Microplate reader
- Assorted glassware for the preparation of reagents and buffer solutions
- Timer
- Vortex mixer

**5 PRECAUTIONS**

1. Read the instructions carefully before beginning the assay.
2. This kit is for research use only.
3. Great care has been taken to ensure the quality and reliability of this product. However, it is possible that in certain cases, unusual results may be obtained due to high levels of interfering factors.
4. Preservatives:  
Positive Control contains 0.1% sodium azide.

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5. No additives or preservatives are necessary to maintain the integrity of the specimen. Avoid azide contamination.
6. Azide and thimerosal at concentrations higher than 0.1% inhibit the enzyme reaction.
7. Other precautions:
  - Do not interchange kit components from different lots.
  - Do not use kit components beyond the expiration date.
  - Protect reagents from direct sunlight.
  - Do not pipette by mouth.
  - Do not eat, drink, smoke or apply cosmetics where reagents are used.
  - Avoid all contact with the reagents by using gloves.
  - Stop solution contains diluted sulfuric acid. Irritation to eyes and skin is possible. Flush with water after contact.

## **6 REAGENT PREPARATION**

### **1. Diluent Concentrate**

The Diluent solution supplied is a 5X concentrate and must be diluted 1:5 with distilled or de-ionized water.

### **2. Wash Solution Concentrate**

The Wash Solution supplied is a 20X concentrate and must be diluted 1:20 with distilled or de-ionized water.

Crystal formation in the concentrate is not uncommon when storage temperatures are low. Warming of the concentrate to 30-35°C before dilution can dissolve crystals.

### **3. Enzyme-Antibody Conjugate Concentrate**

Calculate the required amount of working conjugate solution for each microtiter plate is prepared by adding 10 µL Enzyme-Antibody Conjugate to 990 µL of 1X Diluent for each test strip to be used for testing. Mix uniformly, but gently. Avoid foaming.

### **4. TMB Substrate Solution**

Ready to use as supplied.

### **5. Stop Solution**

Ready to use as supplied.

### **6. Microtiter Plate**

Ready to use as supplied. Unseal Microtiter Pouch and remove plate from pouch. Remove all strips and wells that will not be used in the assay and place back in pouch and re-seal along with desiccant.

### **7. Mouse IgG Calibrator**

The calibrator is now at a concentration of 2.55 mg/mL. Prepare the Mouse IgG Standards immediately prior to use according to the table below. Mix well between each step. Avoid foaming.

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Standard	Concentration (ng/mL)	Standard Volume added to 1X Diluent	Volume of 1X Diluent
A	25500	10 $\mu$ L Mouse IgG Standard	990 $\mu$ L
1	600	14 $\mu$ L Standard 0	581 $\mu$ L
2	300	300 $\mu$ L Standard 1	300 $\mu$ L
3	150	300 $\mu$ L Standard 2	300 $\mu$ L
4	75	300 $\mu$ L Standard 3	300 $\mu$ L
5	37.5	300 $\mu$ L Standard 4	300 $\mu$ L
6	18.75	300 $\mu$ L Standard 5	300 $\mu$ L
7	9.375	300 $\mu$ L Standard 6	300 $\mu$ L
0			500 $\mu$ L

### 8. Positive Control

The concentration and recommended dilution are provided on the Control Certificate.

Before use, briefly centrifuge the Positive Control to allow all of the liquid to collect in the bottom of the vial.

## 7 STORAGE AND STABILITY

### 1. Complete Kit

The expiration date for the kit is stated on the outer label. The recommended storage temperature is 4°C.

**Note: See long term storage recommendations below for the Mouse IgG Calibrator and Positive Control.**

### 2. Diluent

The 5X Diluent Concentrate is stable until the expiration date. The 1X working solution is stable for at least one week from the date of preparation. Both solutions should be stored at 4°C.

### 3. Wash Solution

The 20X Wash Solution Concentrate is stable until the expiration date. The 1X working solution is stable for at least one week from the date of preparation.

Both solutions can be stored at room temperature (RT, 16-25°C) or at 4°C.

### 4. Enzyme-Antibody Conjugate

Undiluted anti-IgG-HRP conjugate should be stored at 4°C and diluted immediately prior to use. The working conjugate solution is stable for one day at 4°C. The working conjugate solution is stable for up to 8 hours.

### 5. TMB Substrate Solution

The TMB Substrate Solution should be stored at 4°C and is stable until the expiration date.

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The Stop Solution should be stored at 4°C and is stable until the expiration date.

**7. Microtiter Plate**

Anti-mouse IgG coated wells are stable until the expiration date and should be stored at 4°C in the sealed foil pouch with a desiccant pack.

**8. Mouse IgG Calibrator**

Long Term Storage: Upon receipt aliquot the calibrator and store them frozen. They will be stable until the expiration date. Short Term Storage: the calibrator is stable for up to 14 days at 4°C. The working standard solutions should be prepared immediately prior to use and are stable for up to 8 hours.

**9. Positive Control**

For storage longer than 7 days keep frozen until the expiration date. Storage less than 7 days can be at 4°C. Avoid multiple freeze/thaw cycles.

**8 INDICATIONS OF INSTABILITY**

If the test is performing correctly, the results observed with the standard solutions should be within 20% of the expected values.

**9 SPECIMEN COLLECTION AND HANDLING**

Blood should be collected by venipuncture and the serum separated from the cells, after clot formation, by centrifugation. For plasma samples, blood should be collected into a container with an anticoagulant and then centrifuged. Care should be taken to minimize hemolysis; excessive hemolysis can impact your results. Assay immediately or aliquot and store samples at -20°C. Avoid repeated freezing/thawing.

For any sample that might contain pathogens, care must be taken to prevent contact with open wounds. No additives or preservatives are necessary to maintain the integrity of the specimen. Avoid azide contamination.

**10 ASSAY PROTOCOL****10.1 Dilution of Samples**

The assay for quantification of IgG in samples requires that each test sample be diluted before use. For a single step determination a dilution of **1/50,000** is appropriate for most serum/plasma samples. For absolute quantification of samples that yield results outside the range of the calibration curve, a lesser or greater dilution might be required. If unsure of a sample level, a serial dilution with one or two representative samples before running the entire plate is highly recommended.

To prepare a 1:50,000 dilution of sample, transfer 5 µL of sample to 995 µL of 1X Diluent. This gives you a 1:200 dilution. Mix thoroughly. Then dilute this by transferring 4 µL to 996 µL of 1X Diluent. You now have a 1:50,000 dilution of your sample. Make sure to mix thoroughly at each stage.

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1. Bring all reagents to RT before use.
2. Pipette 100  $\mu$ L of
  - Calibrator 0 (0.0 ng/mL) in duplicate
  - Calibrator 1 (9.375 ng/mL) in duplicate
  - Calibrator 2 (18.75 ng/mL) in duplicate
  - Calibrator 3 (37.5 ng/mL) in duplicate
  - Calibrator 4 (75 ng/mL) in duplicate
  - Calibrator 5 (150 ng/mL) in duplicate
  - Calibrator 6 (300 ng/mL) in duplicate
  - Calibrator 7 (600 ng/mL) in duplicate
3. Pipette 100  $\mu$ L of diluted Positive Control (in duplicate) into pre-designated wells.
  4. Pipette 100  $\mu$ L of sample (in duplicate) into pre-designated wells.
5. Incubate the Microtiter Plate at 22°C (RT) for sixty ( $60 \pm 2$ ) minutes. Keep plate level during incubation.
6. Following incubation, aspirate the contents of the wells.
7. Completely fill each well with appropriately diluted Wash Solution and aspirate. Repeat three times, for a total of four washes. If washing manually: completely fill wells with diluted Wash Solution, invert the plate and pour/shake out the contents in a waste container. Follow this by sharply striking the wells on absorbent paper to remove residual Wash Solution. Repeat three times for a total of four washes.
8. Pipette 100  $\mu$ L of appropriately diluted Enzyme-Antibody Conjugate to each well. Incubate in the dark at 22°C (RT) for thirty ( $30 \pm 2$ ) minutes. Keep plate covered in the dark and level during incubation.
9. Wash and blot the wells as described in Steps 6 and 7.
10. Pipette 100  $\mu$ L of TMB Substrate Solution into each well.
11. Incubate in the dark at RT for precisely ten (10) minutes.
12. After ten minutes, add 100  $\mu$ L of Stop Solution to each well.
13. Determine the absorbance at 450 nm of the contents of each well. Zero the plate reader to air.

The absorbance of the final reaction mixture can be measured up to 2 hours after the addition of the Stop Solution. However, good laboratory practice dictates that the measurement be made as soon as possible.

**11 RESULTS**

1. Subtract the average background value from the test values for each sample.
2. Using the results observed for the standards construct a calibration curve. The appropriate curve fit is that of a four-parameter logistics curve. A second order polynomial (quadratic) or other curve fits may also be used.
3. Interpolate test sample values from the calibration curve.  
Correct for serum dilution factor to arrive at IgG concentration in original sample.

**12 QUALITY CONTROL**

In accord with good laboratory practice, the assays for total IgG require meticulous quality control. Each laboratory should use routine quality control procedures to establish inter- and intra-assay precision and performance characteristics.

**13 LIMITATION OF THE PROCEDURE**

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of the information contained in the package insert instructions and with adherence to good laboratory practice.
2. Factors that might affect the performance of the assay include proper instrument function, cleanliness of glassware, quality of distilled or de-ionized water, and accuracy of reagent and sample pipetting.