

DRG[®] MIP-2 (rat) ELISA (EIA-3789)

Revised 6 July 2005

RUO

INTENDED USE

The DRG Rat Macrophage Inflammatory Protein-2 (raMIP-2) ELISA is to be used for the *in vitro* quantitative determination of raMIP-2 in rat serum, buffered solution, or cell culture medium. The assay will recognize both natural and recombinant raMIP-2.

**This kit has been configured for research use only and is not to be used in diagnostic procedures.
Read entire protocol before use.**

PRINCIPLE OF THE METHOD

The DRG raMIP-2 kit is a solid phase sandwich Enzyme Linked-Immuno-Sorbent Assay (ELISA). A polyclonal antibody specific for raMIP-2 has been coated onto the wells of the microtiter strips provided. Samples, including standards of known raMIP-2 content, control specimens, and unknowns, are pipetted into these wells, followed by the addition of a biotinylated polyclonal second antibody.

During the first incubation, the raMIP-2 antigen binds simultaneously to the immobilized (capture) antibody on one site, and to the solution phase biotinylated antibody on a second site.

After removal of excess second antibody, Streptavidin-Peroxidase (enzyme) is added. This binds to the biotinylated antibody to complete the four-member sandwich. After a second incubation and washing to remove all the unbound enzyme, a substrate solution is added, which is acted upon by the bound enzyme to produce color. The intensity of this colored product is directly proportional to the concentration of raMIP-2 present in the original specimen.

REAGENTS PROVIDED

Note: Store all reagents at 2 - 8°C.

Reagent	96 Test Kit	192 Test Kit
<i>raMIP-2 Standard</i> , recombinant raMIP-2. Refer to vial label for quantity and reconstitution volume.	2 vials	4 vials
<i>Standard Diluent Buffer</i> . Contains 15 Mm sodium azide; 25 mL per bottle.	1 bottle	2 bottles
<i>raMIP-2 Antibody-Coated Wells</i> , 96 wells per plate.	1 plate	2 plates
<i>raMIP-2 Biotin Conjugate</i> , (Biotin-labeled anti-MIP-2). Contains 15 mM sodium azide; 6 mL per bottle.	1 bottle	2 bottles
<i>Streptavidin-Peroxidase (HRP)</i> , (100x) concentrate. Contains 3.3 mM thymol; 0.125 mL per vial.	1 vial	2 vials
<i>Streptavidin-Peroxidase (HRP) Diluent</i> . Contains 3.3 mM thymol; 25 mL per bottle.	1 bottle	1 bottle
<i>Wash Buffer Concentrate (25x)</i> ; 100 mL per bottle.	1 bottle	1 bottle
<i>Stabilized Chromogen</i> , Tetramethylbenzidine (TMB); 25 mL per bottle.	1 bottle	1 bottle
<i>Stop Solution</i> ; 25 mL per bottle.	1 bottle	1 bottle
<i>Plate Covers</i> , adhesive strips.	3	4

Disposal Note: This kit contains materials with small quantities of sodium azide. Sodium azide reacts with lead and copper plumbing to form explosive metal azides. Upon disposal, flush drains with a large volume of water to prevent azide accumulation. Avoid ingestion and contact with eyes, skin and mucous membranes. In case of contact, rinse affected area with plenty of water. Observe all federal, state and local regulations for disposal.

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SUPPLIES - NOT PROVIDED

1. Microtiter plate reader capable of measurement at or near 450 nm.
2. Calibrated adjustable precision pipettes, preferably with disposable plastic tips. (A manifold multi-channel pipette is desirable for large assays.)
3. Deionized or distilled H₂O.
4. Plate washer: automated or manual (squirt bottle, manifold dispenser, etc.).
5. Graph paper: linear (Cartesian), log-log, or semi-log, as desired.
6. Glass or plastic tubes for diluting and aliquoting standard.
7. Absorbent paper towels.
8. Calibrated beakers and graduated cylinders in various sizes.
9. 37°C incubator.

PROCEDURAL NOTES/LAB QUALITY CONTROL

1. When not in use, kit components should be refrigerated. All reagents should be warmed to room temperature before use.
2. **Microtiter plates should be allowed to come to room temperature before opening the foil bags.** Once the desired number of strips has been removed, immediately reseal the bag and store at 2 - 8°C to maintain plate integrity.
3. Samples should be collected in pyrogen/endotoxin-free tubes.
4. Samples should be frozen if not analyzed shortly after collection. Avoid multiple freeze-thaw cycles of frozen samples. Thaw completely and mix well prior to analysis.
5. When possible, avoid use of badly hemolyzed or lipemic sera. If large amounts of particulate matter are present, centrifuge or filter prior to analysis.
6. It is recommended that all standards, controls and samples be run in duplicate.
7. Samples that are >640 pg/mL should be diluted with *Standard Diluent Buffer*.
8. When pipetting reagents, maintain a consistent order of addition from well-to-well. This ensures equal incubation times for all wells.
9. Cover or cap all reagents when not in use.
10. **Do not mix or interchange different reagent lots from various kit lots.**
11. Do not use reagents after the kit expiration date.
12. Read absorbances within 2 hours of assay completion.
13. In-house controls should be run with every assay. If control values fall outside pre-established ranges, the accuracy of the assay is suspect.
14. All residual wash liquid must be drained from the wells by efficient aspiration or by decantation followed by tapping the plate forcefully on absorbent paper. **Never** insert absorbent paper directly into the wells.
15. Because *Stabilized Chromogen* is light sensitive, avoid prolonged exposure to light. Also avoid contact between *Stabilized Chromogen* and metal, or color may develop.

SAFETY

All blood components and biological materials should be handled as potentially hazardous. Follow universal precautions as established by the Centers for Disease Control and Prevention and by the Occupational Safety and Health Administration when handling and disposing infectious agents.

DIRECTIONS FOR WASHING

Incomplete washing will adversely affect the test outcome. All washing must be performed with *Wash Buffer* provided. Washing can be performed manually as follows: completely aspirate the liquid from all wells by gently lowering an aspiration tip (aspiration device) into the bottom of each well. Take care not to scratch the inside of the well.

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After aspiration, fill the wells with at least 0.4 mL of diluted wash solution. Let soak for 15 to 30 seconds, then aspirate the liquid.

Repeat as directed under **ASSAY METHOD**. After the washing procedure, the plate is inverted and tapped dry on absorbent tissue.

Alternatively, the wash solution may be put into a squirt bottle. If a squirt bottle is used, flood the plate with wash buffer, completely filling all wells. After the washing procedure, the plate is inverted and tapped dry on absorbent tissue.

If using an automated washer, the operating instructions for washing equipment should be carefully followed.

REAGENT PREPARATION AND STORAGE**A. Reconstitution and Dilution of raMIP-2 Standard**

Note: Either glass or plastic tubes may be used for standard dilutions.

1. Reconstitute standard to 2000 pg/mL with *Standard Diluent Buffer*.

Refer to standard vial label for instructions. Swirl or mix gently and allow to sit for 10 minutes to ensure complete reconstitution. Use standard within 1 hour of reconstitution.

2. Add 0.160 mL of the reconstituted standard to a tube containing 0.340 mL *Standard Diluent Buffer*. Label as 640 pg/mL raMIP-2. Mix.

3. Add 0.150 mL of *Standard Diluent Buffer* to each of 6 tubes labeled 320, 160, 80, 40, 20 and 10 pg/mL raMIP-2.

4. Make serial dilutions of the standard as described in the following dilution table. Mix thoroughly between steps.

B. Dilution of raMIP-2 Standard

Discard all remaining reconstituted and diluted standards after completing assay. Return the *Standard Diluent Buffer* to the refrigerator.

Standard:	Add:	Into:
640 pg/mL	Prepare as described in Step 2.	
320 pg/mL	0.150 mL of the 640 pg/mL std.	0.150 mL of the Diluent Buffer
160 pg/mL	0.150 mL of the 320 pg/mL std.	0.150 mL of the Diluent Buffer
80 pg/mL	0.150 mL of the 160 pg/mL std.	0.150 mL of the Diluent Buffer
40 pg/mL	0.150 mL of the 80 pg/mL std.	0.150 mL of the Diluent Buffer
20 pg/mL	0.150 mL of the 40 pg/mL std.	0.150 mL of the Diluent Buffer
10 pg/mL	0.150 mL of the 20 pg/mL std.	0.150 mL of the Diluent Buffer
0 pg/mL	0.150 mL of the Diluent Buffer	An empty tube

C. Storage and Final Dilution of Streptavidin-HRP

Please Note: The *Streptavidin-HRP* 100x concentrate is in 50% glycerol.

This solution is viscous. To ensure accurate dilution, allow *Streptavidin-HRP* concentrate to reach room temperature. Gently mix.

Pipette *Streptavidin-HRP* concentrate slowly. Remove excess concentrate solution from pipette tip by gently wiping with clean absorbent paper.

1. Dilute 10 µL of this 100x concentrated solution with 1 mL of *Streptavidin-HRP Diluent* for each 8-well strip used in the assay.

Label as Streptavidin-HRP Working Solution.

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For Example:

For Example:

# of 8-Well Strips	Volume of Streptavidin-HRP Concentrate	Volume of Diluent
2	20 μ L solution	2 mL
4	40 μ L solution	4 mL
6	60 μ L solution	6 mL
8	80 μ L solution	8 mL
10	100 μ L solution	10 mL
12	120 μ L solution	12 mL

2. Return the unused *Streptavidin-HRP* concentrate to the refrigerator.

D. Dilution of Wash Buffer

Allow the 25x concentrate to reach room temperature and mix to ensure that any precipitated salts have redissolved. Dilute 1 volume of the 25x wash buffer concentrate with 24 volumes of deionized water (e.g., 50 mL may be diluted up to 1.25 liters, 100 mL may be diluted up to 2.5 liters).

Label as Working Wash Buffer.

Store both the concentrate and the Working Wash Buffer in the refrigerator. The diluted buffer should be used within 14 days.

ASSAY METHOD: PROCEDURE AND CALCULATIONS

Be sure to read the *Procedural Notes/Lab Quality Control* section before carrying out the assay.

Allow all reagents to reach room temperature before use. Gently mix all liquid reagents prior to use.

Note: A standard curve must be run with each assay.

1. Determine the number of 8-well strips needed for the assay. Insert these in the frame(s) for current use. (Re-bag extra strips and frame. Store these in the refrigerator for future use.)
2. Add 50 μ L of the *Standard Diluent Buffer* to zero wells. Well(s) reserved for chromogen blank should be left empty.
3. Add 50 μ L of standards, samples or controls to the appropriate microtiter wells. (See **REAGENT PREPARATION AND STORAGE**, Section B.)
4. Pipette 50 μ L of biotinylated anti-MIP-2 (*Biotin Conjugate*) solution into each well except the chromogen blank(s). Tap gently on the side of the plate to mix.
5. Cover plate with *plate cover* and incubate for **2 hours at 37°C**.
6. Thoroughly aspirate or decant solution from wells and discard the liquid. Wash wells 4 times. See **DIRECTIONS FOR WASHING**.
7. Add 100 μ L Streptavidin-HRP Working Solution to each well except the chromogen blank(s). (Prepare the working dilution as described in **REAGENT PREPARATION AND STORAGE**, Section C.)
8. Cover plate with the *plate cover* and incubate for **1 hour at room temperature**.
9. Thoroughly aspirate or decant solution from wells and discard the liquid. Wash wells 4 times. See **DIRECTIONS FOR WASHING**.
10. Add 100 μ L of *Stabilized Chromogen* to each well. The liquid in the wells will begin to turn blue.
11. Incubate for **30 minutes at room temperature and in the dark**.

Please Note: Do not cover the plate with aluminum foil or metalized mylar. The incubation time for chromogen substrate is often determined by the microtiter plate reader used. Many plate readers have the capacity to record a

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maximum optical density (O.D.) of 2.0. The O.D. values should be monitored and the substrate reaction stopped before the O.D. of the positive wells exceed the limits of the instrument. The O.D. values at 450 nm can only be read after the *Stop Solution* has been added to each well. If using a reader that records only to 2.0 O.D., stopping the assay after 20 to 25 minutes is suggested.

12. Add 100 μ L of *Stop Solution* to each well. Tap side of plate gently to mix. The solution in the wells should change from blue to yellow.
13. Read the absorbance of each well at 450 nm having blanked the plate reader against a chromogen blank composed of 100 μ L each of *Stabilized Chromogen* and *Stop Solution*. Read the plate within 2 hours after adding the *Stop Solution*.
14. Plot on graph paper the absorbance of the standards against the standard concentration. (Optimally, the background absorbance may be subtracted from *all* data points, including standards, unknowns and controls, prior to plotting.) Draw the best smooth curve through these points to construct the standard curve. If using curve fitting software, the four parameter algorithm provides the best curve fit.
15. Read the raMIP-2 concentrations for unknown samples and controls from the standard curve plotted in Step 14. (Samples producing signals greater than that of the highest standard (640 pg/mL) should be diluted in *Standard Diluent Buffer* and re-analyzed, multiplying the concentration found by the appropriate dilution factor.)

TYPICAL DATA

The following data were obtained for the various standards over the range of 0 to 640 pg/mL raMIP-2.

Standard raMIP-2 (pg/ml)	Optical Density (450 nm)
0	0.166
	0.164
10	0.248
	0.231
20	0.289
	0.308
40	0.388
	0.462
80	0.617
	0.711
160	1.097
	1.115
320	1.745
	1.852
640	3.169
	3.288

LIMITATIONS OF THE PROCEDURE

Do not extrapolate the standard curve beyond the 640 pg/mL standard point; the dose-response is non-linear in this region and accuracy is difficult to obtain. Dilute samples >640 pg/mL with *Standard Diluent Buffer*; re-analyze these and multiply results by the appropriate dilution factor.

The influence of various drugs, aberrant sera (hemolyzed, hyperlipidemic, jaundiced, etc.) and the use of biological fluids in place of serum samples have not been thoroughly investigated. The rate of degradation of native raMIP-2 in various matrices has not been investigated. The immunoassay literature contains frequent references to aberrant signals seen with some sera, attributed to heterophilic antibodies.

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Though such samples have not been seen to date, the possibility of this occurrence cannot be excluded.

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Not for human therapeutic or diagnostic use.

PERFORMANCE CHARACTERISTICS

SENSITIVITY

The minimum detectable dose of raMIP-2 is <1 pg/mL. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 32 times.

PRECISION

1. Intra-Assay Precision

Samples of known raMIP-2 concentration were assayed in replicates of 10 to determine precision within an assay.

	<u>Sample 1</u>	<u>Sample 2</u>	<u>Sample 3</u>
Mean (pg/mL)	55.3	85.2	171.2
SD	3.5	6.3	6.7
%CV	6.3	7.4	3.9

SD = Standard Deviation

CV = Coefficient of Variation

2. Inter-Assay Precision

Samples were assayed 10 times in multiple assays to determine precision between assays.

	<u>Sample 1</u>	<u>Sample 2</u>	<u>Sample 3</u>
Mean (pg/mL)	40.4	88.2	155.6
SD	4.6	6.6	10.8
%CV	11.4	7.5	6.9

SD = Standard Deviation

CV = Coefficient of Variation

LINEARITY OF DILUTION

A rat serum pool containing 501.3 pg/mL of measured raMIP-2 was serially diluted in *Standard Diluent Buffer* over the range of the assay.

Linear regression analysis of samples versus the expected concentration yielded a correlation coefficient of 1.00.

RECOVERY

The recovery of raMIP-2 added to rat serum averaged 88% (range 82% to 93%). The recovery of raMIP-2 added to tissue culture medium containing 10% fetal calf serum averaged 91%, while the recovery of raMIP-2 added to tissue culture medium containing 1% fetal calf serum averaged 109%.

SPECIFICITY

Buffered solutions of a panel of substances at 10,000 pg/mL were assayed with the DRG raMIP-2 kit. The following substances were tested and found to have no cross-reactivity: mouse IL-2, IL-4, IL-6, TNF-a, IFN-y, MCP-1; rat IFN-y, MCP-1, CINC; human IL-1b, IL-2, IL-4, IL-6, IL-7, IL-10, TNF-a, IFN-y.