

DRG® Anti-SS-A 60 (EIA-3575)



Revised 31 May 2007

RUO in the USA

PRINCIPLE OF THE PROCEDURE

Anti-SS-A 60 is an indirect solid phase enzyme immunometric assay (ELISA). It is designed for the quantitative measurement of IgG class autoantibodies directed against the extractable nuclear antigens SS-A 60 (Ro 60). The microplate is coated with SS-A 60 (Ro 60). The microplate can be divided into 12 modules of 8 wells each or can be used completely for 96 determinations. Each well can be separated from the module ("break-away"). The binding of present autoantibodies, as well as the formation of the sandwich complexes and enzymatic color reaction take place during three different reaction phases:

Phase 1:

Calibrators, controls and prediluted patient samples are pipetted into the wells of the microplate. Any present antibodies bind to the inner surface of the wells. After a 30 minutes incubation the microplate is washed with wash buffer for removing non-reactive serum components.

Phase 2:

An anti-human-IgG horseradish peroxidase conjugate solution is pipetted into the wells of the microplate to recognize the autoantibodies bound to the immobilized antigens. After a 15 minutes incubation any excessive enzyme conjugate, which is not specifically bound is washed away with wash buffer.

Phase 3:

A chromogenic substrate solution containing TMB (3,3',5,5'-Tetramethyl-benzidine) is dispensed into the wells. During 15 minutes of incubation the color of the solutions change into blue. Color development is stopped by adding 1 M hydrochloric acid as stop solution. The solutions color change into yellow. The amount of colour is directly proportional to the concentration of IgG present in the original sample.

To read the optical density a microplate reader with a 450 nm filter is required. Bi-chromatic measurement with a 600-690 nm reference is recommended. The optical density for each calibrator may be graphically plotted against the concentration of IgG and unknowns extrapolated from the curve.

CLINICAL RELEVANCE

Rheumatoid autoimmune diseases are often associated with the occurrence of autoanti-bodies against several nuclear or cytoplasmatic antigens. These so-called anti nuclear antigens (ANA) can be divided into three groups:

1. true anti nuclear antigens (ANA): dsDNA, ssDNA, histones, nucleolic RNA and DNP
 2. extractable nuclears antigens: Sm (Smith), n-RNP, Scl 70 and PM-1
 3. cytoplasmatic antigens: SS-A (Ro)*, SS-B (La)* and Jo-1
- SS-A (Ro) and SS-B (La) are co-localized in cytoplasm and nucleus

In patients with Sjögren-Syndrome antibodies against SS-A and SS-B often occur in combination. Due to the strong association of SS-A and SS-B antibodies to the HLA-DR3 and DR2 phenotypes a genetic predisposition is suspected. The anti SS-A protein passes the placenta and may cause the development of SLE in neonates. Immunoreactive proteins may occur in various combinations and bind also to 'host proteins' of viral origin. They induce synthesis of polyclonal autoantibodies, of the IgG, IgM and IgA class. Especially for mixed connective tissue diseases a relation to viral infections by EBV (Eppstein-Barr-Virus) is indicated. Each class of immunoglobulins causes a specific immunofluorescent pattern. Basically immunoflourescence titers correlate with the quantitation of IgG antibodies but the concentrations may vary considerably within each titer. Quantitation of IgG class antibodies extensively correlates with the diseases' activity. This makes it superior to immuno-fluorescence using Hep2 cells. The IF with Crithidia lucilliae sometimes results in deviating values. Today the best investigated immunoreactive antigens are double-stranded DNA

DRG® Anti-SS-A 60 (EIA-3575)



Revised 31 May 2007

RUO in the USA

(dsDNA), single stranded DNA (ssDNA), Sm (Smith), sn-RNP (small nuclear ribonucleoprotein particles), the complex RNP/Sm which is stabilized by ribonucleic acid as well as SS-A (Ro) and SS-B (La). The antigen Scl 70, a 70 kD molecular weight protein is associated with scleroderma.

In rheumatoid autoimmune diseases various profiles of autoantibodies to these antigens can be detected. In a high incidence they are related to active and inactive systemic Lupus erythematoses, mixed connective tissue diseases (Sharp Syndrome), rheumatoid arthritis, Sjögren-Syndrome, Scleroderma, photosensitive dermatitis and drug-induced lupus. In Lupus patients typically anti-dsDNA antibodies can be detected. Patients without these antibodies very often show anti-ssDNA antibodies and anti-SS-A and anti-SS-B are present. A strong correlation between antibody concentration and severity of the disease has been observed with higher antibody concentrations in active phases of the disease. Thus quantitation is more informative compared to simple titering by immunofluorescence. Measurement of anti-ssDNA provides additional information regarding antibody specificity and activity. Except in chronic inflammatory processes anti-ssDNA antibodies are not found in healthy subjects. Most of these parameters are not specific for just one disease but they occur in various combinations. The pattern of different antibody combinations and their concentration together with the whole clinical picture of the patient are helpful diagnostic tools in the assessment of rheumatoid autoimmune diseases. The following graph gives brief information on the complexity of autoimmune diseases, occurring antibodies. It is not designed as a diagnostic schedule or program for ongoing diagnostic profiles.

Diseases (values in %)	Anti-								
	DsDNA	ssDNA	Histon	SS-A (Ro)	SS-B (La)	Sm	RNP/Sm	Scl 70	Jo-1
Systemic Lupus erythematoses (SLE)	> 90	> 90	30-50	10-30	30-50	10-30	10-30		
Drug induced Lupus (LE)		30-50	50-90						
Sharp-Syndrome / Mixed connective tissue diseases	10-30	10-30					> 90		
Rheumatoid Arthritis		30-50	30-50	10-30					
Sjögren's syndrome	10-30	10-30		> 90	> 90				
Scleroderma	10-30	10-30		10-30				> 90	
Photosensitive Dermatitis Dermatomyositis	10-30	10-30							50-90

Taken from: Thomas, L. Autoantikörper gegen Organe, Gewebe und Thrombozyten.

Labor and Diagnose 1992; Die Medizinische Verlagsgesellschaft Marburg, 4. Auflage: 980 - 990.

NORMAL VALUES

In a normal range study with serum samples from healthy blood donors the following ranges have been established with the Anti-SS-A 60 test:

Anti-SS-A 60 [U/ml]	
normal:	< 10
elevated:	> 10

Positive results should be verified concerning the entire clinical status of the patient. Also every decision for therapy should be taken individually. It is recommended that each laboratory establishes its own normal and pathological ranges of serum Anti-SS-A 60 antibodies. The above reference ranges should be regarded as guidelines only.



Revised 31 May 2007

RUO in the USA

SPECIFICITY

The microplate is coated with SS-A 60 highly purified by affinity chromatography. The Anti-SS-A 60 test kit is specific only for autoantibodies directed to SS-A 60. No crossreactivities to the other ENA-antigens have been observed.

CALIBRATION

The assay system is calibrated in relative arbitrary units.

WARNINGS AND PRECAUTIONS

All reagents of this test kit are strictly intended for in vitro use only. In the United States, this kit is intended for Research Use Only. Please adhere strictly to the sequence of pipetting steps provided in this protocol. Observe the guidelines for performing quality control in medical laboratories by assaying controls and/or pooled sera. All reagents should be stored refrigerated at 2 - 8 °C in their original container. Do not interchange kit components from different lots. The expiration dates stated on the labels of the shipping container and all vials have to be observed. Do not use kit components beyond their expiration dates. Allow all kit components and specimen to reach room temperature prior to use and mix well. During handling of all kit reagents, controls and serum samples observe the existing legal regulations. The following precautions should be taken handling potentially infectious materials:

- do not eat, drink or smoke in areas where specimens or kit reagents are handled
- do not pipette by mouth
- wear disposable gloves while handling specimens or kit reagents and wash hands thoroughly afterwards.

The test kit contains components of human origin which, when tested by FDA-licensed methods, were found negative for hepatitis B surface antigen and for HIV antibody. No known test can guarantee, however, that products derived from human blood will not be infectious. Handle, therefore, all reagents and human blood derivatives, like plasma or serum samples, as if capable of transmitting infection. Avoid contact with the TMB (3,3',5,5'-Tetramethyl-benzidine). If TMB comes into contact with skin wash thoroughly with water and soap. The stop solution contains hydrochloric acid. If it comes into contact with skin, wash thoroughly with water and seek medical attention. Avoid contact between the buffered Peroxide Solution and easily oxidized materials; extreme temperatures may initiate spontaneous combustion.

MATERIALS SUPPLIED

Package size	96 determ.
divisible microplate consisting of 12 modules of 8 wells each,	1
coated with highly purified SS-A 60	
Anti-SS-A 60 calibrators in a PBS/BSA matrix	6 vials, 1.5 ml each
containing respectively: 0; 6.3; 12.5; 25; 50 and 100 U/ml	
Anti-SS-A 60 controls in a serum/buffer matrix	2 vials, 1.5 ml each
(positive and negative), for the respective concentrations	
see the enclosed package insert	
sample buffer, yellow, Concentrate	1 vial, 20 ml
enzyme conjugate solution, (light red) containing polyclonal	1 vial, 15 ml
rabbit anti-h-IgG-IgG; labelled with horseradish peroxidase	
TMB substrate solution	1 vial, 15 ml
stop solution (1 M hydrochloric acid).....	1 vial, 15 ml
buffered wash solution, Concentrate.....	1 vial, 20 ml

**Revised 31 May 2007****RUO in the USA****CONTROLS**

A set of two controls is provided with the kit.

TECHNICAL DATA

Sample material:	serum or plasma
Required sample volume:	10 µl of sample to be diluted 1:100 with sample buffer 100 µl prediluted sample per single determination
Total incubation time:	60 minutes at room temperature (20 - 28 °C)
Calibration range:	6.3 - 100 U/ml
Sensitivity:	0.5 U/ml
Storage:	refrigerated at 2 - 8 °C
Shelf life:	12 months after manufacturing or until the expiration date printed on the labels
Package size:	96 tests

MATERIALS REQUIRED**Equipment**

- Microplate reader capable for endpoint measurements at 450 nm
- Vortex mixer
- Pipets for 10 µl, 100 µl and 1000 µl

Preparation of reagents

- distilled water
- graduated cylinder for 100 and 1000 ml
- plastic container for storage of the wash solution

Optional

- Multi-Chanel Dispenser
- or repeatable pipet for 100 µl
- data reduction software

SPECIMEN COLLECTION AND PREPARATION

For determination of Anti-SS-A 60 antibodies serum or plasma are the preferred sample matrixes.

All serum and plasma samples are prediluted 1 : 100 with sample buffer. Therefore 10 µl of sample may be diluted with 1000 µl of sample buffer. The patients need not to be fasting, and no special preparations are necessary. Collect blood by venipuncture into vacutainers and separate serum or plasma from the cells by centrifugation after clot formation. Samples may be stored refrigerated at 2 - 8 °C for at least 5 days. For longer storage of up to six months samples should be stored frozen at -20 °C. To avoid repeated thawing and freezing the samples should be aliquoted. Neither Bilirubin nor Hemolysis have significant effect on the procedure.

PREPARATION AND STORAGE OF REAGENTS

All components of this test kit are supplied in a liquid format and ready to use, except the sample buffer and wash buffer. When stored refrigerated at 2 - 8 °C the components are stable for at least 30 days after opening or until the expiration



Revised 31 May 2007

RUO in the USA

date printed on the labels. Remaining modules of the microplate should be stored refrigerated at 2 - 8 °C protected from moisture; store together with desiccant and carefully sealed in the plastic bag.

Preparation of sample buffer

Dilute the contents of each vial of the sample buffer concentrate (5x) with distilled water to a final volume of 100 ml prior to use. Store refrigerated: stable at 2 - 8 °C for at least 30 days after preparation or until the expiration date printed on the label.

Preparation of buffered wash solution

Dilute the contents of each vial of the buffered wash solution concentrate (50x) with distilled water to a final volume of 1000 ml prior to use. Store refrigerated: stable at 2 - 8 °C for at least 30 days after preparation or until the expiration date printed on the label.

NOTES ON TECHNIQUE

Control sera or pools should routinely be assayed as unknowns to check performance of the reagents and the assay. For all controls, the respective concentrations are provided on the labels of each vial. Using these concentrations a calibration curve may be calculated to read off the patient results semi-quantitatively.

Pipetting and Sample Handling

Use a disposable-tip micropipette to dispense sera and plasma samples. Pipet directly to the bottom of the wells. To avoid carryover contamination change the tip between samples. Patient samples expected to contain high concentrations should be additionally diluted with sample buffer before. Additional dilutions must be considered during calculation.

IMMUNOASSAY PROCEDURE

Do not interchange components of different lots. All components should be at room temperature before use. Dilute all patient samples 1:100 with sample buffer before assay. Therefore combine 10 µl of sample with 1000 µl of sample buffer in a polystyrene tube. Mix well. Calibrators and controls are ready to use and need not to be diluted.

1. Prepare a sufficient number of microplate modules to accommodate calibrators, controls and prediluted patient samples in duplicates.

	1	2	3	4	5	6
A	SA	SE	P1	P5		
B	SA	SE	P1	P5		
C	SB	SF	P2	P..		
D	SB	SF	P2	P..		
E	SC	C1	P3			
F	SC	C1	P3			
G	SD	C2	P4			
H	SD	C2	P4			

SA - SF: standards A to F
P1, P2... patient sample 1, 2 ...
C1: positive control
C2: negative control

2. Pipet 100 µl of calibrators, controls and prediluted patient samples into the wells.
3. Incubate for 30 minutes at room temperature (20 - 28 °C).
4. Discard the contents of the microwells and wash 3 times with 300 µl of wash solution.



Revised 31 May 2007

RUO in the USA

5. Dispense 100 µl of enzyme conjugate solution into each well.
6. Incubate for 15 minutes at room temperature.
7. Discard the contents of the microwells and wash 3 times with 300 µl of wash solution.
8. Dispense 100 µl of TMB substrate solution into each well.
9. Incubate for 15 minutes at room temperature.
10. Add 100 µl of stop solution to each well of the modules and leave untouched for 5 minutes.
11. Read the optical density at 450 nm and calculate the results. Bi-chromatic measurement with reference at 600-650 nm is recommended. The developed color is stable for at least 30 minutes. Read optical densities during this time.

CALCULATION OF RESULTS

For the Anti-SS-A 60 test a 4-Parameter-Fit with lin-log coordinates for optical density and concentration is recommended. Spline Approximation and log-log coordinates are also suitable.

Recommended Lin-Log Plot

First calculate the averaged optical densities for each calibrator well. Use lin-log graph paper and plot the averaged optical density of each calibrator versus the concentration. Draw the best fitting curve approximating the path of all calibrator points. The calibrator points may also be connected with straight line segments. The concentration of unknowns may then be estimated from the calibration curve by interpolation.

CALCULATION EXAMPLE

The figures below show typical results for Anti-SS-A 60 test. These data are intended for illustration only and should not be used to calculate results from another run.

No	Position	OD 1	OD 2	Mean	Conc. 1	Conc. 2	Mean	decl.Conc.	CV %
STA	A 1 / B 1	0.022	0.021	0.022	0.1	0.1	0.1	0	3.3
STB	C 1 / D 1	0.441	0.433	0.437	6.3	6.2	6.25	6.3	1.3
STC	E 1 / F 1	0.753	0.738	0.746	12.8	12.4	12.6	12.5	1.4
STD	G 1 / H 1	1.141	1.173	1.157	24.7	25.9	25.2	25.0	1.9
STE	A 2 / B 2	1.603	1.570	1.586	49.9	47.5	48.8	50	0
STF	C 2 / D 2	2.022	2.011	2.017	102.8	100.6	101.7	100	1.7

ASSAY CHARACTERISTICS

Sensitivity

The lower detection limit for the Anti-SS-A 60 test was determined at 0.5 U/ml.

Parallelism

In dilution experiments sera with high antibody concentrations were diluted with sample buffer and assayed in the Anti-SS-A 60 kit. The assay showed linearity over the full measuring range.

Precision

Statistics were calculated for each of three samples from the results of 24 determinations in a single run for Intra-Assay precision and the run-to-run precision was calculated from the results of 5 different runs with 6 determinations each:


Revised 31 May 2007
RUO in the USA

Intra-Assay		
Sample No	Mean [U/ml]	CV [%]
1	14.7	3.1
2	33.3	2.7
3	69.1	3.2

Inter-Assay		
Sample No	Mean [U/ml]	CV [%]
1	17.3	5.9
2	31.9	6.3
3	62.3	1.8

REFERENCES

1. Antinuclear Antibodies
The Lancet 1984; 15: 611 - 613.
2. Froelich, Ch. J., Wallmann, H., Skosey, J. L. and Teodorescu, M.
Clinical Value of an Integrated ELISA System for the Detection of 6 Autoantibodies (ssDNA, dsDNA, Sm, RNP/Sm, SSA and SSB).
The Journal of Rheumatology 1990; Vol 17, No 2: 192 - 200.
3. Harley, J. B. and Gaither, K. K.
Autoantibodies.
Rheumatic Disease Clinics of North America 1988; Vol 14, No 1: 43 - 56.
4. Slobbe, R.L. et al.
Detection and occurrence of the 60- and 52-kD Ro (SS-A) antigens and of autoantibodies against these proteins.
Clin. Exp. Immunol. 1991; 86: 99 - 105.
5. Hartung, K. and Deicher, H.
Systemischer Lupus erythematosus.
Allergologie 1985; Jahrgang 8, Nr. 7: 275 - 281.
6. Hartung, K. and Deicher, H.
Symptome und Symptommuster beim systemischen Lupus erythematosus.
Die Internistische Welt 1987; 9: 236 - 241.
7. Takeda, J. et al.
Antigenic Domains on the U1 Small Nuclear Ribonucleoprotein-Associated 70K Polypeptide: A Comparison of Regions Selectively Recognized by Human and Mouse Autoantibodies and by Monoclonal Antibodies.
Clinical Immunology and Immunopathology 1991; 61: 55 - 68.
8. Reichlin, M. and Van Venrooij, W. J.
Autoantibodies to the URNP particles: relationship to clinical diagnosis and nephritis.
Clin. exp. Immunol. 1991; 83: 286 - 290.
9. Hietarinta, M., Lassila, O. and Hietaharju, A.
Association of Anti-U1RNP- and Anti-Scl-70-antibodies with Neurological Manifestations in Systemic Sclerosis (Scleroderma).
Scand. J. Rheumatol. 1994; 23: 64 - 67.
10. Reveille, J.D. et al.
Racial Differences in the Frequencies of Scleroderma-related Autoantibodies.
Arthritis and Rheumatism 1992; Vol 35, No 2: 216 - 218.
11. Genth, E. and Mierau, R.
Jo-1-(Antisynthetase-)Syndrom - verbessern Autoantikörper die Klassifikation von Myositiden?
Zeitschrift für Rheumatologie 1993; 52: 259 - 263.