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RUO in the USA

REVISED 21 SEPT. 2010 RM (VERS. 3.1)

Please use only the valid version of the package insert provided with the kit.

1 NAME AND INTENDED USE

The ANA-8-Screen assay is intended to screen for the presence of antinuclear antibodies (ANAs) in human serum or plasma for certain systemic rheumatic diseases. This assay collectively detects, in one well, ANAs against SS-A/Ro, SS-B/La, RNP 70, Sm, RNP/Sm, Centromere B and Jo-1.

2 PRINCIPLE OF THE TEST

Purified antigens (SS-A/Ro, SS-B/La, RNP 70, Sm, RNP/Sm, Scl-70, centromere B and Jo-1) are bound to microwells. Antibodies against these antigens, if present in diluted serum or plasma, bind to the respective antigen. Washing of the microwells removes unspecific serum and plasma components. Horseradish peroxidase (HRP) conjugated anti-human IgG immunologically detects the bound donor antibodies forming a conjugate/antibody/antigen complex. Washing of the microwells removes unbound conjugate. An enzyme substrate in the presence of bound conjugate hydrolyzes to form a blue colour. The addition of an acid stops the reaction forming a yellow end-product.

The intensity of this yellow colour is measured photometrically at 450 nm.

3 WARNINGS AND PRECAUTIONS

- 1. All reagents of this kit are strictly intended for research use only.
- 2. Do not interchange kit components from different lots.
- 3. Components containing human serum were tested and found negative for HBsAg, HCV, HIV1 and HIV2 by FDA approved methods. No test can guarantee the absence of HBsAg, HCV, HIV1 or HIV2, and so all human serum based reagents in this kit must be handled as though capable of transmitting infection.
- 4. Avoid contact with the TMB (3,3′,5,5′-Tetramethyl-benzidine). If TMB comes into contact with skin, wash thoroughly with water and soap.
- 5. Avoid contact with the Stop Solution which is acid. If it comes into contact with skin, wash thoroughly with water and seek medical attention.
- 6. Some kit components (i.e. Controls, Sample buffer and Buffered Wash Solution) contain Sodium Azide as preservative. Sodium Azide (NaN₃) is highly toxic and reactive in pure form. At the product concentrations (0.09%), though not hazardous. Despite the classification as nonhazardous, we strongly recommend using prudent laboratory practices (see 8., 9., 10.).
- 7. Some kit components contain Proclin 300 as preservative. When disposing reagents containing Proclin 300, flush drains with copious amounts of water to dilute the components below active levels.
- 8. Wear disposable gloves while handling specimens or kit reagents and wash hands thoroughly afterwards.
- 9. Do not pipette by mouth.
- 10. Do not eat, drink, smoke or apply makeup in areas where specimens or kit reagents are handled.
- 11. Avoid contact between the buffered Peroxide Solution and easily oxidized materials; extreme temperature may initiate spontaneous combustion.





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Observe the guidelines for performing quality control in medical laboratories by assaying controls and/or pooled sera. During handling of all kit reagents, controls and serum samples observe the existing legal regulations.

4 CONTENTS OF THE KIT

Package size 96 determ.

Qty.1 Divisible **ANA microtiter strips**: 96 antigen-coated wells sealed in a foil pouch with desiccant.

Ready to use.

2 vials, 1.5 ml each Anti-ANA controls in a serum/buffer matrix (PBS, NaN₃ <0.1% (w/w)).

Negative Control (NC), Calibrator control (CC).

Ready to use

1 vial, 20 ml **Sample buffer** (Tris, NaN₃ <0.1% (w/w)), yellow,

concentrate (5x)

1 vial, 15 ml Enzyme conjugate solution (PBS, Proclin 300 <0.5% (v/v)), (light red)

containing polyclonal rabbit anti-human IgG; labelled with horseradish peroxidase.

Ready to use

1 vial, 15 ml **TMB substrate solution**.

Ready to use

1 vial, 15 ml Stop solution (contains acid).

Ready to use

1 vial, 20 ml Wash solution (PBS, $NaN_3 < 0.1\%$ (w/w)),

concentrate (50x)

5 STORAGE AND STABILITY

- 1. Store the kit at 2-8°C
- 2. Keep microplate wells sealed in a dry bag with desiccants
- 3. The reagents are stable until expiration of the kit
- 4. Do not expose test reagents to heat, sun or strong light during storage and usage
- 5. Diluted sample buffer and wash buffer are stable for at least 30 days when stored at 2-8°C

6 MATERIALS REQUIRED

Equipment

- Microplate reader capable of endpoint measurements at 450 nm
- Multi-Channel Dispenser or repeatable pipet for $100 \ \mu l$
- Vortex mixer
- Pipets for 10 μl, 100 μl and 1000 μl
- Laboratory timing device
- Data reduction software





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Preparation of reagents

- Distilled or deionized water
- Graduated cylinder for 100 and 1000 ml
- Plastic container for storage of the wash solution

7 SPECIMEN COLLECTION, STORAGE AND HANDLING

- 1. Collect whole blood specimens using acceptable medical techniques to avoid hemolysis.
- 2. Allow blood to clot and separate the serum by centrifugation.
- 3. Test serum should be clear and non-hemolysed. Contamination by hemolysis or lipemia is best avoided, but does not interfere with this assay.
- 4. Specimens may be refrigerated at 2-8 °C for up to five days or stored at -20 °C up to six months.
- 5. Avoid repetitive freezing and thawing of serum samples. This may result in variable loss of autoantibody activity.
- 6. Testing of heat-inactivated sera is not recommended.

8 PROCEDURAL NOTES

- 1. Do not use kit components beyond their expiration dates.
- 2. Do not interchange kit components from different lots.
- 3. All materials must be at room temperature (20-28 °C).
- 4. Have all reagents and samples ready before start of the assay. Once started, the test must be performed without interruption to get the most reliable and consistent results.
- 5. Perform the assay steps only in the order indicated.
- 6. Always use fresh sample dilutions.
- 7. Pipette all reagents and samples into the bottom of the wells.
- 8. To avoid carryover contamination, change the tip between samples and different kit controls.
- 9. It is important to wash microwells thoroughly and remove the last droplets of wash buffer to achieve best results.
- 10. All incubation steps must be accurately timed.
- 11. Control sera or pools should routinely be assayed as unknowns to check performance of the reagents and the assay.
- 12. Do not re-use microplate wells.

For all controls, the respective concentrations are provided on the labels of each vial. Using these concentrations a calibration curve may be calculated to read off the donor results semi-quantitatively.







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9 PREPARATION OF REAGENTS

9.1 Preparation of sample buffer

Dilute the contents of each vial of the sample buffer concentrate (5x) with distilled or deionized water to a final volume of 100 ml prior to use.

Store refrigerated: stable at 2-8°C for at least 30 days after preparation or until the expiration date printed on the label.

9.2 Preparation of wash solution

Dilute the contents of each vial of the buffered wash solution concentrate (50x) with distilled or deionized water to a final volume of 1000 ml prior to use.

Store refrigerated: stable at 2-8°C for at least 30 days after preparation or until the expiration date printed on the label.

9.3 Sample preparation

Dilute all donor samples 1:100 with sample buffer before assay.

Therefore combine 10 µl of sample with 990 µl of sample buffer in a polystyrene tube. Mix well.

Controls are ready to use and need not be diluted.

10 TEST PROCEDURE

- 1. Prepare a sufficient number of microplate modules to accommodate controls and prediluted donor samples.
- 2. Pipet 100 μl of calibrators, controls and prediluted donor samples in duplicate into the wells.

		1	2	3	4	5	6
ı				,	7		U
	A	CC	P				
	В	NC	P				
	C	P1					
	D	P2					
	E	P3					
	F	P4					
	G	P5					
	Н	P6					

P1, P2... donor samples 1, 2 ...

CC: calibrator control

NC: negative control

- 3. Incubate for 30 minutes at room temperature (20-28°C)
- 4. Discard the contents of the microwells and wash 3 times with 300 μl of wash solution.
- 5. Dispense 100 μl of enzyme conjugate into each well





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- 6. Incubate for 15 minutes at room temperature
- 7. Discard the contents of the microwells and wash 3 times with 300 μ l of wash solution
- 8. Dispense 100 μl of TMB substrate solution into each well
- 9. Incubate for 15 minutes at room temperature
- 10. Add 100 μl of stop solution to each well of the modules and incubate for 5 minutes at room temperature
- 11. Read the optical density at 450 nm and calculate the results. Bi-chromatic measurement with a reference at 600-690 nm is recommended.

The developed colour is stable for at least 30 minutes. Read optical densities during this time.

11 REFERENCES / LITERATURE

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