DRG® Herpes Simplex Virus Type 1+2 IgG (HSV - 1+2) (EIA-3489)

Revised 3 Mar. 2011 rm (Vers. 7.1) **RUO in the USA**

This kit is intended for Research Use Only.

Not for use in diagnostic procedures.

Introduction

The **DRG Herpes Simplex Virus Type 1+2 IgG Enzyme Immunoassay Kit** provides materials for determination of IgG-class antibodies to Herpes Simplex Virus Type 1+ 2 in human serum.

PRINCIPLE of the test

The **DRG Herpes Simplex Virus Type 1+2 IgG ELISA** Kit is a solid phase enzyme-linked immunosorbent assay (ELISA)

Microtiter wells as a solid phase are coated with Herpes Simplex Virus Type 1 antigen.
Diluted specimens and **ready-for-use controls** are pipetted into these wells. During incubation Herpes Simplex Virus Type 1+2-specific antibodies of positive specimens and controls are bound to the immobilized antigens. After a washing step to remove unbound sample and control material horseradish peroxidase conjugated anti-human IgG antibodies are dispensed into the wells. During a second incubation this anti-IgG conjugate binds specifically to IgG antibodies resulting in the formation of enzyme-linked immune complexes.
After a second washing step to remove unbound conjugate the immune complexes formed (in case of positive results) are detected by incubation with TMB substrate and development of a blue color. The blue color turns into yellow by stopping the enzymatic indicator reaction with sulfuric acid.
The intensity of this color is directly proportional to the amount of Herpes Simplex Virus Type 1+2-specific IgG antibody in the specimen. Absorbance at 450 nm is read using an ELISA microtiter plate reader.

Precautions

- All reagents of this test kit which contain human serum or plasma have been tested and confirmed negative for HIV I/II, HBsAg and HCV by FDA approved procedures. All reagents, however, should be treated as potential biohazards in use and for disposal.
- The Controls and Standards have been found to be non-infectious in cell cultures.
- Avoid contact with *Stop Solution* containing 0.2 mol/L H_2SO_4. It may cause skin irritation and burns.
- Never pipette by mouth and avoid contact of reagents and specimens with skin and mucous membranes.
- Do not smoke, eat, drink or apply cosmetics in areas where specimens or kit reagents are handled.
- Wear disposable latex gloves when handling specimens and reagents. Microbial contamination of reagents or specimens may give false results.
- Handling should be in accordance with the procedures defined by an appropriate national biohazard safety guideline or regulation.
- Do not use reagents beyond expiry date as shown on the kit labels.
- All indicated volumes have to be performed according to the protocol. Optimal test results are only obtained when using calibrated pipettes and microtiter plate readers.

Revised 3 Mar. 2011 rm (Vers. 7.1) **RUO in the USA**

- Do not mix or use components from kits with different lot numbers. It is advised not to exchange wells of different plates even of the same lot. The kits may have been shipped or stored under different conditions and the binding characteristics of the plates may result slightly different.
- Chemicals and prepared or used reagents have to be treated as hazardous waste according the national biohazard safety guideline or regulation.
- For information on hazardous substances included in the kit please refer to Material Safety Data Sheets. Material Safety Data Sheets for this product are available upon request directly from DRG.

Kit Components

Contents of the Kit

1. ***Microtiterwells***, 12 x 8 (break apart) strips, 96 wells;
   Wells coated with Herpes Simplex Virus Type 1+2 antigen.
   (incl. 1 strip holder and 1 cover foil)
2. ***Sample Diluent*** ***, 1 vial, 100 mL, ready to use,
   colored yellow; pH 7.2 ± 0.2.
3. ***Pos. Control*** ***, 1 vial, 2.0 mL, ready to use;
   colored yellow, red cap.
4. ***Neg. Control*** ***, 1 vial, 2.0 mL, ready to use;
   colored yellow, yellow cap.
5. ***Cut-off Control*** ***, 1 vial, 2.0 mL, ready to use;
   colored yellow, black cap.
6. ***Enzyme Conjugate*** **, 1 vial, 20 mL, ready to use,
   colored red,
   antibody to human IgG conjugated to horseradish peroxidase.
7. ***Substrate Solution***, 1 vial, 14 mL, ready to use,
   Tetramethylbenzidine (TMB).
8. ***Stop Solution***, 1 vial, 14 mL, ready to use,
   contains 0.2 mol/l H_2SO_4,
   Avoid contact with the stop solution. It may cause skin irritations and burns.
9. ***Wash Solution*** *, 1 vial, 30 mL (20X concentrated for 600 mL), pH 7.2 ± 0.2
   see „Preparation of Reagents“.

* contains 0.03 % ProClin 300
** contains 0.03 % ProClin 300 + 0.01 % Gentamicin sulphate
*** contains 0.03 % ProClin 300 + 0.015 % 5-bromo-5-nitro-1,3-dioxane (BND) +
0.010 % 2-methyl-2H-isothiazol-3-one (MIT)

Revised 3 Mar. 2011 rm (Vers. 7.1) RUO **in the USA**

Equipment and material required but not provided

- A microtiter plate calibrated reader (450/620nm ±10 nm)
  (e.g. the DRG Instruments Microtiter Plate Reader)
- Calibrated variable precision micropipettes
- Incubator 37 °C
- Manual or automatic equipment for rinsing wells
- Vortex tube mixer
- Deionised or (freshly) distilled water
- Timer
- Absorbent paper

Storage and stability of the Kit

When stored at 2 °C to 8 °C unopened reagents will retain reactivity until expiration date. Do not use reagents beyond this date.
Opened reagents must be stored at 2 °C to 8 °C. Microtiter wells must be stored at 2 °C to 8 °C. Once the foil bag has been opened, care should be taken to close it tightly again.
Opened kits retain activity for four months if stored as described above.

Preparation of Reagents

Allow all reagents and required number of strips to reach room temperature prior to use.

Wash Solution

Dilute *Wash Solution* **1+19** (e.g. 10 mL + 190 mL) with fresh and germ free redistilled water.
Consumption: ~ 5 mL per determination.
Crystals in the solution disappear by warming up to 37 °C in a water bath.
The diluted Wash Solution is stable for 4 weeks at 2 °C to 8 °C.

Disposal of the Kit

The disposal of the kit must be made according to the national regulations. Special information for this product is given in the Material Safety Data Sheets (see chapter 13 of this data sheet).

Damaged Test Kits

In case of any severe damage to the test kit or components, DRG has to be informed in writing, at the latest, one week after receiving the kit. Severely damaged single components should not be used for a test run. They have to be stored until a final solution has been found. After this, they should be disposed according to the official regulations.

Revised 3 Mar. 2011 rm (Vers. 7.1) RUO **in the USA**

SPECIMEN

Serum can be used in this assay.
Do not use haemolytic, icteric or lipaemic specimens.

Specimen Collection

Serum:

Collect blood by venipuncture (e.g. Sarstedt Monovette # 02.1388.001), allow to clot, and separate serum by centrifugation at room temperature. Do not centrifuge before complete clotting has occurred. Samples containing anticoagulant may require increased clotting time.

Specimen Storage

Specimens should be capped and may be stored for up to 24 hours at 2 °C to 8 °C prior to assaying. Specimens held for a longer time should be frozen only once at –20 °C prior to assay. Thawed samples should be inverted several times prior to testing.

Specimen Dilution

Prior to assaying dilute each specimen **1+100** with *Sample Diluent*; e.g. 10 µL of specimen + 1 mL of *Sample Diluent*, **mix well, let stand for 15 minutes, mix again gently**.

Please note: Controls are ready for use and must not be diluted!

test procedure

General Remarks

- Please read the test protocol carefully before performing the assay. Result reliability depends on strict adherence to the test protocol as described.
- **It is very important to bring all reagents, samples and controls to room temperature before starting the test run!**
- Once the test has been started, all steps should be completed without interruption.
- Use new disposal plastic pipette tips for each standard, control or sample in order to avoid cross contamination
- Absorbance is a function of the incubation time and temperature. Before starting the assay, it is recommended that all reagents are ready, caps removed, all needed wells secured in holder, etc. This will ensure equal elapsed time for each pipetting step without interruption.
- As a general rule the enzymatic reaction is linearly proportional to time and temperature.
- Close reagent vials tightly immediately after use to avoid evaporation and microbial contamination.
- After first opening and subsequent storage check conjugate and control vials for microbial contamination prior to further use.
- To avoid cross-contamination and falsely elevated results pipette samples and dispense conjugate without splashing accurately to the bottom of wells.
- During incubation cover microtiter strips with foil to avoid evaporation.

Revised 3 Mar. 2011 rm (Vers. 7.1) **RUO in the USA**

Assay Procedure

Prior to commencing the assay, dilute *Wash Solution*, **prepare samples as described in point 5.3** and establish carefully the **distribution and identification plan** supplied in the kit for all specimens and controls.

1. Select the required number of microtiter strips or wells and insert them into the holder.

   Please allocate at least:
   1 well (e.g. A1) for the substrate blank,
   1 well (e.g. B1) for the *Neg. Control,*
   2 wells (e.g. C1+D1) for the *Cut-off Control* and
   1 well (e.g. E1) for the *Pos. Control*.

   It is left to the user to determine controls and samples in duplicate.

2. Dispense
   100 µL of *Neg. Control* into well B1
   100 µL of *Cut-off Control* into wells C1 and D1
   100 µL of *Pos. Control* into well E1 and
   100 µL of each diluted sample with new disposable tips into appropriate wells.
   Leave well A1 for substrate blank!

3. Cover wells with foil supplied in the kit. Incubate for **60 minutes at 37 °C.**

4. Briskly shake out the contents of the wells.
   Rinse the wells **5 times** with diluted *Wash Solution* (**300 µL per well**). Strike the wells sharply on absorbent paper to remove residual droplets.
   Important note:
   The sensitivity and precision of this assay is markedly influenced by the correct performance of the washing procedure!

5. Dispense **100 µL** *Enzyme Conjugate* into each well, **except A1**.

6. Cover wells with foil. Incubate for **30 minutes at room temperature (20 °C to 25 °C).**
   Do not expose to direct sun light!

7. Briskly shake out the contents of the wells.
   Rinse the wells **5 times** with diluted *Wash Solution* (300 µL per well). Strike the wells sharply on absorbent paper to remove residual droplets.

8. Add **100 µL** of *Substrate Solution* into all wells.

9. Cover wells with foil. Incubate for **exactly 15 minutes at room temperature (20 °C to 25 °C) in the dark**.

10. Stop the enzymatic reaction by adding **100 µL** of *Stop Solution* to each well.
    Any blue color developed during the incubation turns into yellow.
    Note: Highly positive samples can cause dark precipitates of the chromogen!

11. Read the optical density at **450/620 nm** with a microtiter plate reader **within 30 minutes** after adding the *Stop Solution*.

Measurement

Adjust the ELISA microplate or microstrip reader **to zero** using the **substrate blank in well A1**.

If - due to technical reasons - the ELISA reader cannot be adjusted to zero using the substrate blank in well A1, subtract the absorbance value of well A1 from all other absorbance values measured in order to obtain reliable results!

Measure the absorbance of all wells **at 450 nm** and record the absorbance values for each control and sample in the distribution and identification plan.

Dual wavelength reading using 620 nm as reference wavelength is recommended.

Where applicable **calculate the mean absorbance values** of all duplicates.

Results

Validation of the Test Run

The test run may be considered valid provided the following criteria are met:

Substrate blank in A1:	Absorbance value **lower than 0.100**
***Neg. Control* in B1:**	Absorbance value **lower than 0.200**
***Cut-off Control* in C1/D1 :**	Absorbance value **between 0.250 – 0.850**
***Pos. Control* in E1:**	Absorbance value **greater than 0.600**

The absorbance value of the Pos. Control should be greater than the absorbance value of the Cut-off Control!

Calculation

Mean absorbance value of Cut-off Control [CO]

Calculate the mean absorbance value of the two (2) Cut-off Control determinations (e.g. in C1/D1).

Example: *(0.44 + 0.46) : 2 = 0.45 = CO*

REFERENCES / LiterAture

1. Corey,L.,P.G.Spear:infections with herpes simplex viruses.N.Eng.J.Med.314 (1986) 686-691.
2. Dörries,R.,R.Kaiser,H.Imrich et al.: NeuereAspekte zur Diagnose zentralnervöser Virusinfektionen.Lab.Med.15 (1991) 99-102.

Revised 3 Mar. 2011 rm (Vers. 7.1) RUO **in the USA**

Version 2011-02-17~rm

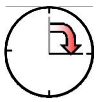
DRG® Herpes Simplex Virus Type 1+2 IgG (HSV - 1+2) (EIA-3489)

Revised 3 Mar. 2011 rm (Vers. 7.1) **RUO in the USA**

Short Instructions for Use

	All reagents and specimens must be allowed to come to room temperature (18-25°C) before use.
	Leave well A1 for substrate Blank. Dispense 100 µl of Controls into appropriate wells.
	Dispense 100 µl of pretreated sample into selected wells. **(Please note special sample treatment, point 5.3!)**
 60 min	Cover wells with foil. Incubate for **60 minutes** at 37°C.
	Briskly shake out the contents of the wells.
	Rinse the wells **5 times** with diluted *Wash Solution* (300 µl per well).
	Strike the wells sharply on absorbent paper to remove residual droplets.
	Dispense 100 µl of *Enzyme Conjugate* into each well.
30 min 	Incubate for **30 minutes** at room temperature.
	Briskly shake out the contents of the wells.
	Rinse the wells **5 times** with diluted *Wash Solution* (300 µl per well).
	Strike the wells sharply on absorbent paper to remove residual droplets.

Revised 3 Mar. 2011 rm (Vers. 7.1) RUO **in the USA**

	Add 100 µl of *Substrate Solution* to each well.
	Incubate for **15 minutes** at room temperature.
	Stop the reaction by adding 100 µl of *Stop Solution* to each well.
	Determine the absorbance of each well at 450 nm.