DRG® Swine Interleukin-10 (swIL-10) (EIA-3184 // EIA-3185)

Revised 24 May 2011 rm (Vers. 3.1) **For Veterinary Use Only**

Please use only the valid version of the package insert provided with the kit.

Contents and Storage

Storage

Store at 2 to 8°C.

Contents

Reagent Provided	96 Test Kit	192 Test Kit
Sw IL-10 Standard, lyophilized, recombinant Sw IL-10. Contains 0.1% sodium azide. Refer to vial label for quantity and reconstitution volume.	2 vials	4 vials
Standard Diluent Buffer. Contains 0.1% sodium azide; 25 mL per bottle.	1 bottle	2 bottles
Antibody Coated Wells. 12 x 8 Well Strips.	1 plate	2 plates
Sw IL-10 Biotin Conjugate (Biotin-labeled anti-IL-10). Contains 0.1% sodium azide; 11 mL per bottle.	1 bottle	2 bottles
Streptavidin-HRP (100X). Contains 3.3 mM thymol; 0.125 mL per vial.	1 vial	2 vials
Streptavidin-HRP Diluent. Contains 3.3 mM thymol; 25 mL per bottle.	1 bottle	1 bottle
Wash Buffer Concentrate (25X). 100 mL per bottle.	1 bottle	1 bottle
Stabilized Chromogen, Tetramethylbenzidine (TMB). 25 mL per bottle.	1 bottle	1 bottle
Stop Solution. 25 mL per bottle.	1 bottle	1 bottle
Plate Covers, adhesive strips.	3	6

Disposal Note

This kit contains materials with small quantities of sodium azide. Sodium azide reacts with lead and copper plumbing to form explosive metal azides. Upon disposal, flush drains with a large volume of water to prevent azide accumulation. Avoid ingestion and contact with eyes, skin and mucous membranes. In case of contact, rinse affected area with plenty of water. Observe all federal, state and local regulations for disposal.

Safety
All blood components and biological materials should be handled as potentially hazardous. Follow universal precautions as established by the Centers for Disease Control and Prevention and by the Occupational Safety and Health Administration when handling and disposing of infectious agents.

INTRODUCTION

Purpose

The Swine Interleukin-10 (Sw IL-10) ELISA is to be used for the determination of Sw IL-10 in swine serum, swine plasma, buffered solution, or cell culture medium. The assay will recognize both natural and recombinant Sw IL-10.
For Research Use Only. CAUTION: Not for human or animal therapeutic or diagnostic use.

Principle of the Method

The Sw IL-10 kit is a solid phase sandwich EnzymeLinked-Immuno-Sorbent Assay (ELISA). An antibody specific for Sw IL-10 has been coated onto the wells of the microtiter strips provided. Samples, including standards of known Sw IL-10 content, control specimens, and unknowns, are pipetted into these wells.
During the first incubation, the Sw IL-10 antigen binds to the immobilized (capture) antibody on one site. After washing, a biotinylated antibody specific for Sw IL-10 is added. During the second incubation, this antibody binds to the immobilized Sw IL-10 captured during the first incubation.
After removal of excess second antibody, Streptavidin-Peroxidase (enzyme) is added. This binds to the biotinylated antibody to complete the four-member sandwich. After a third incubation and washing to remove all the unbound enzyme, a substrate solution is added, which is acted upon by the bound enzyme to produce color. The intensity of this colored product is directly proportional to the concentration of Sw IL-10 present in the original specimen.

Background Information

Interleukin-10 (Hu IL-10) is a lymphokine produced by T helper lymphocytes, monocytes, macrophages and B-lymphocytes. IL-10 was first characterised as a cytokine synthesis inhibitory factor (CSIF) able to inhibit cytokine synthesis by Th1 clones activated in the presence of antigen presenting cells. However, in the absence of monocytes, IL-10 directly inhibits the growth of T-cells triggered by immobilised anti-CD3 monoclonal antibody. This inhibition of proliferation is a result of IL-2 production by the responding T-cells. In vitro, IL-10 is a very powerful inhibitor of monokines (including TNF-α, IL-1, IL-6 and IL-8) produced by LPS-activated monocytes and macrophages. The addition of IL-10 to B lymphocytes results in limited cell proliferation but most importantly in very high immunoglobulin production, a result of the transformation of B-cells into plasma cells. Finally, natural killer (NK) cells appear to be another target for the anti-inflammatory properties of IL-10. Indeed, recent data have shown that IL-10 can inhibit antigen induced IFN-γ production by NK-cells by inhibiting not only production but also the stimulatory effects of IL-12 and TNF on IFN-γ production.

METHODS

Materials Needed But Not Provided

- Microtiter plate reader (at or near 450 nm) with software
- Calibrated adjustable precision pipettes
- Distilled or deionized water
- Plate washer: automated or manual (squirt bottle, manifold dispenser, etc.)
- Glass or plastic tubes for diluting solutions
- Absorbent paper towels
- Calibrated beakers and graduated cylinders

Procedural Notes

1. When not in use, kit components should be refrigerated. All reagents should be warmed to room temperature before use.
2. **Microtiter plates should be allowed to come to room temperature before opening the foil bags.** Once the desired number of strips has been removed, immediately reseal the bag and store at 2 to 8°C to maintain plate integrity.
3. Samples should be collected in pyrogen/endotoxin-free tubes.
4. Samples should be frozen if not analyzed shortly after collection. Avoid multiple freeze-thaw cycles of frozen samples. Thaw completely and mix well prior to analysis.
5. When possible, avoid use of badly hemolyzed or lipemic sera. If large amounts of particulate matter are present, centrifuge or filter prior to analysis.
6. It is recommended that all standards, controls and samples be run in duplicate.
7. When pipetting reagents, maintain a consistent order of addition from well-to-well. This ensures equal incubation times for all wells.
8. **Do not mix or interchange different reagent lots from various kit lots.**
9. Do not use reagents after the kit expiration date.
10. 10.Absorbances should be read immediately, but can be read up to 2 hours after assay completion. For best results, keep plate covered in the dark.
11. 11.In-house controls should be run with every assay. If control values fall outside pre-established ranges, the accuracy of the assay is suspect.
12. 12.All residual wash liquid must be drained from the wells by efficient aspiration or by decantation followed by tapping the plate forcefully on absorbent paper. **Never** insert absorbent paper directly into the wells.
13. Because Stabilized Chromogen is light sensitive, avoid prolonged exposure to light. Avoid contact between chromogen and metal, or color may develop.

Directions for Washing

- **Incomplete washing will adversely affect the test outcome.** All washing must be performed with the Wash Buffer Concentrate (25X) provided.

- Washing can be performed manually as follows: completely aspirate the liquid from all wells by gently lowering an aspiration tip into the bottom of each well. Take care not to scratch the inside of the well. After aspiration, fill the wells with at least 0.4 ml of diluted Wash Buffer. Let soak for 15 to 30 seconds, then aspirate the liquid. Repeat as directed under Assay Procedure. After the washing procedure, the plate is inverted and tapped dry on absorbent tissue.
- Alternatively, the diluted Wash Buffer may be put into a squirt bottle. If a squirt bottle is used, flood the plate with the diluted Wash Buffer, completely filling all wells. After the washing procedure, the plate is inverted and tapped dry on absorbent tissue.
- If using an automated washer, follow the washing instructions carefully.

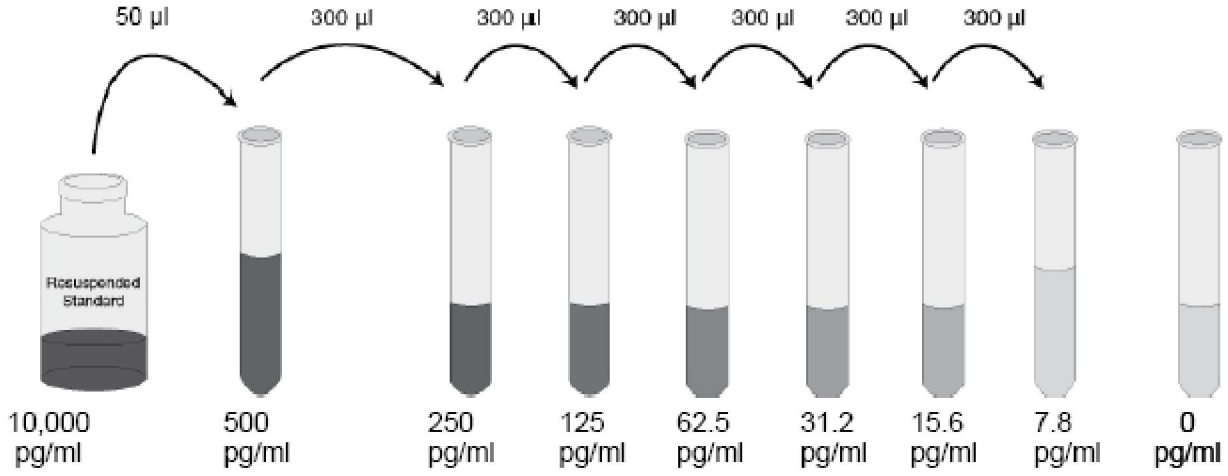
Preparation of Reagents

Dilution of Standard

Note: Either glass or plastic tubes may be used for standard dilutions.

1. Reconstitute standard to 10,000 pg/ml with Standard Diluent Buffer. Refer to standard vial label for instructions. Swirl or mix gently and allow to sit for 10 minutes to ensure complete reconstitution. Use standard within 1 hour of reconstitution.
2. Add 0.050 ml of the reconstituted standard to a tube containing 0.950 ml Standard Diluent Buffer. Label as 500 pg/ml Sw IL-10. Mix.
3. Add 0.300 ml of Standard Diluent Buffer to each of 6 tubes labeled 250, 125, 62.5, 31.2, 15.6 and 7.8 pg/ml Sw IL-10.
4. Make serial dilutions of the standard as described in the following dilution diagram. Mix thoroughly between steps.

Note: Remaining reconstituted standard should be discarded. Return the Standard Diluent Buffer to the refrigerator.



Preparing SAV-HRP

Note: Prepare within 15 minutes of usage. The Streptavidin-HRP (100X) is in 50% glycerol, which is viscous. To ensure accurate dilution, allow Streptavidin-HRP (100X) to reach room temperature. Gently mix. Pipette Streptavidin-HRP (100X) slowly. Remove excess concentrate solution from pipette tip by gently wiping with clean absorbent paper.

1. Dilute 10 µl of this 100X concentrated solution with 1 ml of Streptavidin-HRP Diluent for each 8-well strip used in the assay. Label as Streptavidin-HRP Working Solution.
2. Return the unused Streptavidin-HRP (100X) to the refrigerator.

# of 8-Well Strips	Volume of Streptavidin-HRP (100X)	Volume of Diluent
2	20 µl solution	2 ml
4	40 µl solution	4 ml
6	60 µl solution	6 ml
8	80 µl solution	8 ml
10	100 µl solution	10 ml
12	120 µl solution	12 ml

Dilution of Wash Buffer

1. Allow the Wash Buffer Concentrate (25X) to reach room temperature and mix to ensure that any precipitated salts have redissolved. Dilute 1 volume of the Wash Buffer Concentrate (25X) with 24 volumes of deionized water (e.g., 50 ml may be diluted up to 1.25 liters, 100 ml may be diluted up to 2.5 liters).
   Label as Working Wash Buffer.
2. Store both the concentrate and the Working Wash Buffer in the refrigerator.
   The diluted buffer should be used within 14 days.

Assay Procedure

Be sure to read the Procedural Notes section before carrying out the assay.
Allow all reagents to reach room temperature before use. Gently mix all liquid reagents prior to use.
Note: A standard curve must be run with each assay.

1. Determine the number of 8-well strips needed for the assay. Insert these in the frame(s) for current use. (Re-bag extra strips and frame. Store these in the refrigerator for future use.)
2. Add 100 µl of the Standard Diluent Buffer to the zero standard wells. Well(s) reserved for chromogen blank should be left empty.
3. For the standard curve, add 100 µl of standards to the appropriate microtiter wells.
   For buffered solutions or cell culture samples, add 100 µl of sample to each well.
   For controls, plasma and serum samples, add 50 µl of Standard Diluent Buffer to each well followed by 50 µl of sample. Tap gently on the side of plate to mix. *See Preparation of Reagents.*
4. Cover plate with plate cover and incubate for **1 hour at room temperature**.
5. Thoroughly aspirate or decant solution from wells and discard the liquid. Wash wells 4 times.
   See Directions for Washing.
6. Pipette 100 µl of biotinylated Sw IL-10 Biotin Conjugate solution into each well except the chromogen blank(s). Tap gently on the side of the plate to mix.
7. Cover plate with plate cover and incubate for **1 hour at room temperature**.

8. Thoroughly aspirate or decant solution from wells and discard the liquid. Wash wells 4 times. *See Directions for Washing.*
9. Add 100 µl Streptavidin-HRP Working Solution to each well except the chromogen blank(s). *See Preparation of Reagents.*
10. Cover plate with the plate cover and incubate for **30 minutes at room temperature**.
11. Thoroughly aspirate or decant solution from wells and discard the liquid. Wash wells 4 times. *See Directions for Washing.*
12. Add 100 µl of Stabilized Chromogen to each well. The liquid in the wells willbegin to turn blue.
13. Incubate for **30 minutes at room temperature and in the dark.**
    Note: Do not cover the plate with aluminum foil or metalized mylar.
    The incubation time for chromogen substrate is often determined by the microtiter plate reader used. Many plate readers have the capacity to record a maximum optical density (O.D.) of 2.0. The O.D. values should be monitored and the substrate reaction stopped before the O.D. of the positive wells exceeds the limits of the instrument. The O.D. values at 450 nm can only be read after the Stop Solution has been added to each well. If using areader that records only to 2.0 O.D., stopping the assay after 20 to 25 minutes is suggested.
14. Add 100 µl of Stop Solution to each well. Tap side of plate gently to mix. The solution in the wells should change from blue to yellow.
15. Read the absorbance of each well at 450 nm having blanked the plate reader against a chromogen blank composed of 100 µl each of Stabilized Chromogen and Stop Solution. Read the plate within 2 hours after adding the Stop Solution.
16. Use a curve fitting software to generate the standard curve. A four parameter algorithm provides the best standard curve fit.
17. Read the concentrations for unknown samples and controls from the standard curve. **Multiply value(s) obtained for serum sample(s) by 2 to correct for the 1:2 dilution.** (Samples producing signals greater than that of the highest standard should be diluted in Standard Diluent Buffer and reanalyzed, multiplying the concentration found by the appropriate dilution factor.)

Typical Data (Example)

The following data were obtained for the various standards over the range of 0 to 500 pg/ml Sw IL-10.

Standard Sw IL-10 (pg/ml)	Optical Density (450 nm)
500	2.71
250	1.36
125	0.74
62.5	0.40
31.2	0.22
15.6	0.14
7.8	0.11
0	0.06

PERFORMANCE CHARACTERISTICS

Sensitivity

The minimum detectable dose of Sw IL-10 is < 3.0 pg/ml. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 30 times.

Precision

1. Intra-Assay Precision

Samples of known Sw IL-10 concentration were assayed in replicates of 16 to determine precision within an assay.

	Sample 1	Sample 2	Sample 3
Mean (pg/ml)	59.9	182.0	311.5
SD	3.8	7.3	15.7
%CV	6.3	4.0	5.0
SD = Standard Deviation CV = Coefficient of Variation			

2. Inter-Assay Precision
Samples were assayed 48 times in multiple assays to determine precision between assays.

	Sample 1	Sample 2	Sample 3
Mean (pg/ml)	58.1	180.0	324.4
SD	4.2	8.8	30.5
%CV	7.2	4.9	9.4
SD = Standard Deviation CV = Coefficient of Variation			

Linearity of Dilution

Swine serum containing 447 pg/ml of measured Sw IL-10 was serially diluted in Standard Diluent Buffer over the range of the assay. Linear regression analysis of samples versus the expected concentration yielded a correlation coefficient of 0.99.

Recovery

The recovery of Sw IL-10 added to swine serum averaged 103%.
The recovery of Sw IL-10 in plasma averaged 105%.
The recovery of Sw IL-10 added to tissue culture medium containing 1% fetal bovine serum averaged 87%, while the recovery of Sw IL-10 added to tissue culture medium containing 10% fetal bovine serum averaged 92%.
Sera and plasma from Yorkshire and Chester-White pigs have been validated for use in this assay.
Other strains of swine have not been tested and consequently their use has not been validated

Specificity

Buffered solutions of a panel of substances at 10,000 pg/ml were assayed with the Sw IL-10 kit. The following substances were tested and found to have no cross-reactivity:
human IL-1β, IL-2, IL-4, IL-7, IL-8, IL-13, IFN-γ, SCF, TNF-α;
mouse IL-1β, IL-2, IL-4, IL-6, IFN-γ, TNF-α; rat IL-1β, IL-2, IL-10, IFN-γ, MCP-1, TNF- α.
Significant cross-reactivity was observed to human IL-10.

Limitations of the Procedure

Do not extrapolate the standard curve beyond the top standard point; the dose-response is non-linear in this region and accuracy is difficult to obtain. Dilute all samples above the top standard point with Standard Diluent Buffer; reanalyze these and multiply results by the appropriate dilution factor.
The influence of various drugs, aberrant sera (hemolyzed, hyperlipidemic, jaundiced, etc.) and the use of biological fluids in place of serum samples have not been thoroughly investigated. The rate of degradation of native Sw IL-10 in various matrices has not been investigated. The immunoassay literature contains frequent references to aberrant signals seen with some sera, attributed to heterophilic antibodies. Though such samples have not been seen to date, the possibility of this occurrence cannot be excluded.

Appendix

Troubleshooting Guide

Elevated background

Cause: Insufficient washing and/or draining of wells after washing. Solution containing either biotin or Streptavidin-HRP can elevate the background if residual is left in the well.

Solution: Wash according to the protocol. Verify the function of automated plate washer. At the end of each washing step, invert plate on absorbent tissue on countertop and allow to completely drain and tap forcefully if necessary to remove residual fluid.

Cause: Contamination of substrate solution with metal ions or oxidizing reagents.

Solution: Use distilled/deionized water for dilution of wash buffer and use plastic equipment. DO NOT COVER plate with foil.

Cause: Contamination of pipette, dispensing reservoir or substrate solution with SAV-HRP conjugate.

Solution: Do not use chromogen that appears blue prior to dispensing onto the plate. Obtain new vial of chromogen.

Cause: Incubation time is too long or incubation temperature is too high.

Solution: Reduce incubation time and/or temperature.

Elevated sample/standard ODs

Cause: Incorrect dilution of standard stock solution; intermediary dilutions not followed correctly.

Solution: Follow the protocol instructions regarding the dilution of the standard.

Cause: Incorrect dilution of the Streptavidin-HRP Working Solution.

Solution: Warm solution of Streptavidin-HRP (100X) to room temperature, draw up slowly and wipe tip with kim-wipe to remove excess. Dilute ONLY in Streptavidin-HRP Diluent provided.

Cause: Incubation times extended.

Solution: Follow incubation times outlined in protocol.

Cause: Incubations carried out at 37°C when RT is dictated.

Solution: Perform incubations at RT (= 25 ± 2°C) when instructed in the protocol.

Poor standard curve

Cause: Improper preparation of standard stock solution.

Solution: Dilute lyophilized standard as directed by the vial label only with the standard diluent buffer or in a diluent that most closely matches the matrix of your sample.

Cause: Reagents (lyophilized standard, standard diluent buffer, etc.) from different kits, either different cytokine or different lot number, were substituted.

Solution: NEVER substitute any components from another kit.

Cause: Errors in pipetting the standard or subsequent steps.

Solution: Always dispense into wells quickly and in the same order. Do not touch the pipette tip on the individual microwells when dispensing. Use calibrated pipettes and the appropriate tips for that device.

Weak/no color develops

Cause: Reagents not at RT (25 ± 2°C) at start of assay.
Solution: Allow ALL reagents to warm to RT prior to commencing assay.

Cause: Incorrect storage of components, e.g., not stored at 2 to 8°C.
Solution: Store all components exactly as directed in protocol and on labels.

Cause: Working Streptavidin-HRP solution made up longer than 15 minutes before use in assay.
Solution: Use the diluted Streptavidin-HRP within 15 minutes of dilution.

Cause: TMB solution lost activity.
Solution 1: The TMB solution should be clear before it is dispensed into the wells of the microtiter plate. An intense aqua blue color indicates that the product is contaminated. Please contact Technical Support if this problem is noted. To avoid contamination, we recommend that the quantity required for an assay be dispensed into a disposable trough for pipetting. Any TMB solution left in the trough should be discarded.
Solution 2: Avoid contact of the TMB solution with items containing metal ions.

Cause: Attempt to measure analyte in a matrix for which the ELISA assay has not been optimized.
Solution: Please contact Technical Support for advice when using nonvalidated sample types.

Cause: Wells have been scratched with pipette tip or washing tips.
Solution: Use caution when dispensing and aspirating into and out of microwells.

Poor Precision

Cause: Errors in pipetting the standards, samples or subsequent steps.
Solution: Always dispense into wells quickly and in the same order. Do not touch the pipette tip on the individual microwells when dispensing. Use calibrated pipettes and the appropriate tips for that device. Check for any leaks in the pipette tip.

Cause: Repetitive use of tips for several samples or different reagents.
Solution: Use fresh tips for each sample or reagent transfer.

Cause: Wells have been scratched with pipette tip or washing tips.
Solution: Use caution when dispensing and aspirating into and out of microwells.

References / Literature

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