



Revised 3 April 2008

For Veterinary Use Only

1 INTRODUCTION

For diagnosis of Canine Parvo Virus (CPV) infection or vaccination control, demonstration of antibody titer is the most commonly used method. The virus that is attached to the solid phase by use of monoclonal antibodies catches antibodies induced through infection or vaccination.

IgG antibody titers above dilutions of 1:810 are considered protected.

2 INTENDED USE

The principle of the CPV testkit is based on the detection of antibodies against Parvo virus. The Parvo virus is attached to the solid phase by use of a monoclonal antibody. After the attachment of the antigen (Parvo virus) sera containing antibodies are able to react with the bound antigen. After the antigen/antibody reaction, the attached antibodies can be detected by use of a polyclonal conjugate.

3 PRINCIPLE

The test is based on the reaction of CPV proteins with dog antibodies.

To this end CPV proteins have been coated to a 96-well microtiter plate.

The diluted dog serum/plasma sample is added to the wells of the coated plate.

After washing the bound dog antibodies are detected by a HRPO conjugated anti-species conjugate.

The colour reaction in the wells is directly related to the concentration of CPV antibodies in the serum/plasma sample.

4 CONTENTS

- 12 x 8-well microtiter strips
- 1 x stripholder
- 1 x 11 ml inactivated Canine Parvo Virus **antigen**.
- 1 x 18 ml ELISA buffer.
- 1 x 12 ml HRPO conjugate (conjugated anti-species antibodies)
- 1 x 0.5 ml **Positive control** (Freeze dried).
- 1 x 0.5 ml **Negative control** (Freeze dried).
- 1 x 20 ml Wash-solution (200 x concentrated), dilute in deionised water before use!
- 1 x 8 ml Substrate buffer A
- 1 x 8 ml **Substrate buffer B**
- 1 x 8 ml Stop solution
- 1 x plastic cover seal

5 HANDLING AND STORAGE OF SPECIMENS

The ELISA should be stored at 4-8°C. An unopened package can be used until the expiry date.

Avoid repeated freezing and thawing as this increases non-specific reactivity.





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Samples may be used fresh or may be kept frozen below -20°C before use.

6 WASHING PROTOCOL

In Elisa's, un-complexed components must be removed efficiently between each incubation step. This is accomplished by appropriate washing. It should be stressed that each washing step must be carried out with care to guarantee reproducible inter- and intra-assay results. It is essential to follow the washing procedures outlined below.

Washing may be done manually or with automatic equipment. Automatic washing equipment usually gives better results.

Manual washing

- 1. Empty each well by turning the microtiter plate upside down, followed by a firm vertical movement.
- 2. Fill all the wells with 250 µl washing solution.
- 3. This washing cycle (1 and 2) should be carried out at least 4 times.
- 4. Turn the plate upside down and empty the wells by a firm vertical movement.
- 5. Place the inverted plate on absorbent paper towels and tap the plate firmly to remove residual washing solution in the wells.
- 6. Take care that none of the wells dries out before the next reagent is dispensed.

Washing with automatic equipment

When using automatic plate wash equipment, check that all wells can be aspirated completely and that the washing solution is correctly dispensed, reaching the rim of each well during each rinsing cycle. The washer should be programmed to execute at least 4 washing cycles.

7 TEST PROTOCOL

- 1. Open the packet of strips, take out the strips to be used (see 4). Cover the remaining strips with a part of the provided seal; store them at 4°C and use within 10 days.
 - Wash the microtiter strips with washing solution according to washing protocol.

The washing solution provided must be diluted 200x in de-ionized water!

- 2. Reconstitute the controls in 0.5 ml aquabidest water, store them in aliquots at -20°C
- 3. Dispense 100 µl of inactivated Canine Parvo Virus antigen to all wells to be used.
- 4. Incubate 75 min. at 37°C.
- 5. Make 3-step dilutions of each sample in ELISA buffer, starting 1:30 (90; 270; 810) in a round bottomed microtiter plate.
 - Make also a 3-step dilution of the positive and negative control.
- 6. Wash as in 1.
- 7. Transfer 100 μl of this dilution to the CPV coated microtiter strips. Seal and incubate for 60 min. at 37°C
- 8 Wash as in 1
- 9. Dispense 100 µl conjugated anti-species antibody to all wells.
- 10. Seal and incubate 60 min. at 37°C.
- 11. Wash as in 1.





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12. Mix equal parts of buffer A and B with gentle shaking. Prepare immediately before use!

Dispense 100 µl substrate solution to each well.

Incubate 10-15 min. at room temperature (21°C.) If temperature is above 28 °C than incubate shorter for 5-10 min. instead of 10-15 min.

- 13. Add 50 µl stop solution to each well.
- 14. Read the absorbency values <u>immediately (within 10 min!)</u> at 450 nm.

8 VALIDATION OF THE TEST

The negative control (at the 1:30 dilution) should give an $OD \le 0.400$.

The end point titer of the positive control should be between 1:150 and 1:450 according to the instructions for interpretation of test results.

9 INTERPRETATION OF TEST RESULTS

The titer of the sample is the dilution which gives an extinction above 2 times the OD value of the negative control (at the 1:30 dilution).

The test is valid if the first two dilutions of the positive control are above 0.500 OD (450 nm).

In summary: 30 = no antibodies found.

90-270 = antibodies found.

 \geq 810 = high titer of antibodies found.

10 PRECAUTIONS

- Handle all biological material as though capable of transmitting CPV.
- Do not pipette by mouth.
- Do not eat, drink, smoke or prepare foods, or apply cosmetics within the designated work area.
- TMB is toxic by inhalation, through contact with skin or when swallowed; observe care when handling the substrate.
- Do not use components past the expiry date and do not mix components from different serial lots together.
- Optimal results will be obtained by strict adherence to this protocol. Careful pipetting and washing throughout this
 procedure are necessary to maintain precision and accuracy.
- Each well is ultimately used as an optical cuvette. Therefore, do not touch the under-surface of the microtiter plate and protect is from damage and dirt.

The entire risk as to the performance of these products is assumed by the purchaser. DRG shall not be liable for indirect, special or consequential damages of any kind resulting from use of the products.





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SYMBOLS USED WITH DRG ASSAY'S

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