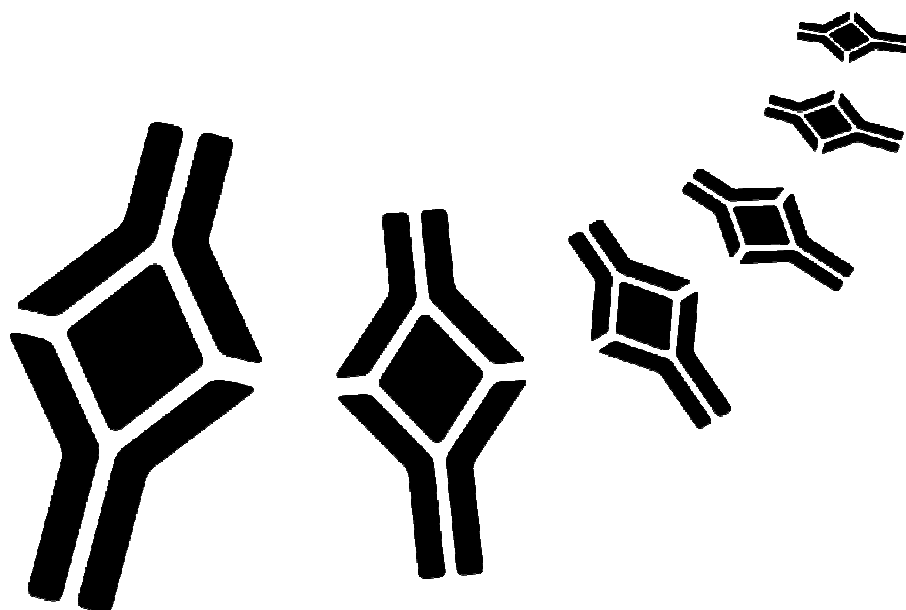


**BioVendor**

Research  
and Diagnostic Products



## HUMAN MxA PROTEIN ELISA

Product Data Sheet

Cat. No.: RK001R

For Research Use Only

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**»» This kit is manufactured by:  
BioVendor – Laboratorní medicína, a.s.**

**»» Use only the current version of Product Data Sheet enclosed with the kit!**

## 1. INTENDED USE

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The

Human MxA Protein ELISA is a sandwich enzyme immunoassay for the quantitative measurement of human MxA Protein.

### »» Features

- **It is intended for research use only.**
- The total assay time is less than 2 hours.
- The kit measures MxA protein in whole blood (cell lysate).
- Assay format is 96 wells.
- Standard is recombinant protein based.
- Components of the kit are provided ready to use, concentrated or lyophilized.

## 2. STORAGE, EXPIRATION

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Store the complete kit at 2-8°C. Under these conditions, the kit is stable until the expiration date (see label on the box).

For stability of opened reagents see Chapter 9.

### 3. INTRODUCTION

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Human MxA protein (Myxovirus resistance protein 1), the product of the MX1 gene, is a 76-kDa protein consisting of 662 amino acid residues and belonging to the dynamic superfamily of large GTPase.

MxA protein plays an important role in antiviral activity in cells against a wide variety of viruses, including influenza, parainfluenza, measles, coxsackie, hepatitis B virus, and Thogoto virus. The viruses are inhibited by MxA protein at an early stage in their life cycle, soon after host cell entry and before genome amplification. The mouse MxA (MX1 GTPase) accumulates in the cell nucleus where it associates with nuclear bodies and inhibits influenza and Thogoto viruses known to replicate in the nucleus. The human MxA protein accumulates in the cytoplasm and endoplasmic reticulum as well. The membrane compartment of endoplasmic reticulum seems to provide an interaction platform that facilitates viral target recognition. MxA appears to detect viral infection by sensing and trapping nucleocapsid-like structures. As a consequence, the viral components become unavailable for the generation of new virus particles.

The expression of viral MxA protein is induced exclusively and in a dose-dependent manner by IFN-alpha and IFN-beta, but not by IFN-gamma, IL-1, TNF-alpha or other cytokines.

In clinical diagnostics, MxA protein may offer advantages as a marker for viral infection over the other induced proteins such as 2', 5'-oligoadenylate synthetase, because of its very low basal concentration and long half-life. Several clinical studies have reported on the possible use of MxA protein expression in peripheral blood mononuclear cells as a marker distinguishing viral from bacterial disease, and reliable marker for type I IFN bioavailability during IFN treatment of chronic viral hepatitis and multiple sclerosis.

#### **Areas of investigation:**

Infection

## 4. TEST PRINCIPLE

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In the BioVendor Human MxA Protein ELISA, Standards and samples are incubated in microplate wells pre-coated with monoclonal anti-human MxA protein antibody. After 60 minutes incubation and washing, monoclonal anti-human MxA protein antibody, conjugated with horseradish peroxidase (HRP) is added to the wells and incubated for 30 minutes with captured MxA protein. Following another washing step, the remaining HRP conjugate is allowed to react with the substrate solution (TMB). The reaction is stopped by addition of acidic solution and absorbance of the resulting yellow product is measured spectrophotometrically at 450 nm. The absorbance is proportional to the concentration of MxA protein. A standard curve is constructed by plotting absorbance values against concentrations of Standards, and concentrations of unknown samples are determined using this standard curve.

**Since the human MxA protein is induced by only 1-type interferon (interferon  $\alpha/\beta$ ) and expressed mainly in mononuclear cells, lysis procedure of blood cells is required before the assay. In this MxA ELISA test kit, Dilution Buffer is used for the lysis of blood cells.**

## 5. PRECAUTIONS

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- **For professional use only.**
- Wear gloves and laboratory coats when handling immunodiagnostic materials.
- Do not drink, eat or smoke in the areas where immunodiagnostic materials are being handled.
- Avoid contact with the acidic Stop Solution and Substrate Solution, which contains hydrogen peroxide and tetramethylbenzidine (TMB). Wear gloves and eye and clothing protection when handling these reagents. Stop and/or Substrate Solutions may cause skin/eyes irritation. In case of contact with the Stop Solution and the Substrate Solution wash skin/eyes thoroughly with water and seek medical attention, when necessary.
- The materials must not be pipetted by mouth.

## 6. TECHNICAL HINTS

- Reagents with different lot numbers should not be mixed.
- Use thoroughly clean glassware.
- Use deionized (distilled) water, stored in clean containers.
- Avoid any contamination among samples and reagents. For this purpose, disposable tips should be used for each sample and reagent.
- Substrate Solution should remain colourless until added to the plate. Keep Substrate Solution protected from light.
- Stop Solution should remain colourless until added to the plate. The colour developed in the wells will turn from blue to yellow immediately after the addition of the Stop Solution. Wells that are green in colour indicate that the Stop Solution has not mixed thoroughly with the Substrate Solution.
- Dispose of consumable materials and unused contents in accordance with applicable national regulatory requirements.

## 7. REAGENT SUPPLIED

<i>Kit Components</i>	<i>State</i>	<i>Quantity</i>
Antibody Coated Microtiter Strips	ready to use	96 wells
Conjugate Solution	ready to use	11 ml
Master Standard	lyophilized	1 vial
Dilution Buffer	ready to use	30 ml
Wash Solution Concentrate (10x)	concentrated	100 ml
Substrate Solution	ready to use	11 ml
Stop Solution	ready to use	11 ml
Product Data Sheet + Quality Control Sheet		1 pc

## 8. MATERIAL REQUIRED BUT NOT SUPPLIED

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- Deionized (distilled) water
- Test tubes for diluting samples
- Glassware (graduated cylinder and bottle) for Wash Solution (Dilution Buffer)
- Precision pipettes to deliver 10-1000 µl with disposable tips
- Multichannel pipette to deliver 100 µl with disposable tips
- Absorbent material (e.g. paper towels) for blotting the microtiter plate after washing
- Vortex mixer
- Orbital microplate shaker capable of approximately 300 rpm
- Microplate washer (optional). [Manual washing is possible but not preferable.]
- Microplate reader with 450 ± 10 nm filter
- Software package facilitating data generation and analysis (optional)

## 9. PREPARATION OF REAGENTS

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- All reagents need to be brought to room temperature prior to use.
- Always prepare only the appropriate quantity of reagents for your test.
- Do not use components after the expiration date marked on their label.

- Assay reagents supplied ready to use:

### **Antibody Coated Microtiter Strips**

#### Stability and storage:

Return the unused strips to the provided aluminium zip-sealed bag with desiccant and seal carefully. Remaining Microtiter Strips are stable 3 months stored at 2-8°C and protected from the moisture.

### **Conjugate Solution**

Conjugate Solution is ready to use, do not dilute it.

#### Stability and storage:

Opened Standards are stable 3 months when stored at 2-8°C.

### **Dilution Buffer**

### **Substrate Solution**

### **Stop Solution**

#### Stability and storage:

Opened reagents are stable 3 months when stored at 2-8°C.

- **Assay reagents supplied concentrated or lyophilized:**

#### **Human MxA Protein Master Standard**

**Refer to the Quality Control Sheet for current volume of distilled water needed for reconstitution of standard!!!**

Reconstitute the lyophilized Master Standard with distilled water just prior to the assay. Let it dissolve at least 15 minutes with occasionally gently shaking (not to foam). The resulting concentration of the MxA protein in the stock solution is **24 ng/ml**.

Prepare set of standards using Dilution Buffer as follows:

Volume of Standard	Dilution Buffer	Concentration
Stock	-	24 ng/ml
250 µl of stock	250 µl	12 ng/ml
250 µl of 12 ng/ml	250 µl	6 ng/ml
250 µl of 6 ng/ml	250 µl	3 ng/ml
250 µl of 3 ng/ml	250 µl	1.5 ng/ml
250 µl of 1.5 ng/ml	250 µl	0.75 ng/ml
250 µl of 0.75 ng/ml	250 µl	0.38 ng/ml

**Prepared Standards are ready to use, do not dilute them.**

#### Stability and storage:

Standard stock solution (25 ng/ml) should be aliquoted and frozen at –20°C for 3 months. Avoid repeated freeze/thaw cycles.

Do not store the diluted Standard Solutions.

#### **Wash Solution Concentrate (10x)**

Dilute Wash Solution Concentrate (10x) ten-fold in distilled water to prepare a 1x working solution. Example: 100 ml of Wash Solution Concentrate (10x) + 900 ml of distilled water for use of all 96-wells.

#### Stability and storage:

The diluted Wash Solution is stable 1 month when stored at 2-8°C. Opened Wash Solution Conc. (10x) is stable 3 months when stored at 2-8°C.



## 10. PREPARATION OF SAMPLES

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The kit measures MxA protein in whole blood (cell lysate).

Whole blood samples should be collected in EDTA collection tubes and assayed immediately after collection or should be stored at -20°C.

Cell lysis: Dilute whole blood samples prior to the assay 10x with Dilution Buffer, e.g. 25 µl of sample + 225 µl of Dilution Buffer, mix well (not to foam), and incubate for 30 minutes at room temperature.

### Stability and storage:

Whole blood samples are stable at 4°C for 24 hours. Whole blood samples should be stored at -20°, or preferably at -70°C, for long-term storage.

*Note: It is recommended to use a precision pipette and a careful technique to perform the dilution in order to get precise results.*

## 11. ASSAY PROCEDURE

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1. Pipet **100 µl** of diluted Standards, Dilution Buffer (=Blank) and samples, preferably in duplicates, into the appropriate wells.
2. Incubate the plate at room temperature (ca. 25°C) for **1 hour**, shaking at ca. 300 rpm on an orbital microplate shaker.
3. Wash the wells **5-times** with Wash Solution (0.35 ml per well). After final wash, invert and tap the plate strongly against paper towel.
4. Add **100 µl** of POD Conjugate into each well.
5. Incubate the plate at room temperature (ca. 25°C) for **30 minutes**, shaking at ca. 300 rpm on an orbital microplate shaker.
6. Wash the wells **5-times** with Wash Solution (0.35 ml per well). After final wash, invert and tap the plate strongly against paper towel.
7. Add **100 µl** of TMB Substrate Solution into each well. Avoid exposing the microtiter plate to direct sunlight. Covering the plate with e.g. aluminium foil is recommended.
8. Incubate the plate for **10 minutes** at room temperature. The incubation time may be extended [up to 20 minutes] if the reaction temperature is below than 20°C. Do not shake the plate during the incubation.
9. Stop the colour development by adding **100 µl** of Stop Solution.
10. Determine the absorbance by reading the plate at 450 nm. The absorbance should be read within 5 minutes following step 9.

*Note: If some samples and standard/s have absorbances above the upper limit of your microplate reader, perform a second reading at 405 nm. A new standard curve, constructed using the values measured at 405 nm, is used to determine SP-D concentration of off-scale standards and samples. The readings at 405 nm should not replace the readings for samples that were “in range” at 450 nm.*

*Note 2: Manual washing: Aspirate wells and pipet 0.35 ml Wash Solution into each well. Aspirate wells and repeat twice. After final wash, invert and tap the plate strongly against paper towel. Make certain that Wash Solution has been removed entirely.*

	strip 1+2	strip 3+4	strip 5+6	strip 7+8	strip 9+10	strip 11+12
<b>A</b>	<b>Standard 24</b>	Sample 1	Sample 9	Sample 17	Sample 25	Sample 33
<b>B</b>	<b>Standard 12</b>	Sample 2	Sample 10	Sample 18	Sample 26	Sample 34
<b>C</b>	<b>Standard 6</b>	Sample 3	Sample 11	Sample 19	Sample 27	Sample 35
<b>D</b>	<b>Standard 3</b>	Sample 4	Sample 12	Sample 20	Sample 28	Sample 36
<b>E</b>	<b>Standard 1.5</b>	Sample 5	Sample 13	Sample 21	Sample 29	Sample 37
<b>F</b>	<b>Standard 0.75</b>	Sample 6	Sample 14	Sample 22	Sample 30	Sample 38
<b>G</b>	<b>Standard 0.38</b>	Sample 7	Sample 15	Sample 23	Sample 31	Sample 39
<b>H</b>	<b>Blank</b>	Sample 8	Sample 16	Sample 24	Sample 32	Sample 40

Figure 1: Example of a work sheet.

## 12. CALCULATIONS

Most microplate readers perform automatic calculations of analyte concentration. The Standard curve is constructed by plotting the mean absorbance at 450 nm (Y) of Standards against log of the known concentration (X) of Standards, using the four-parameter algorithm. Results are reported as concentration of MxA protein ng/ml.

Alternatively, the logit log function can be used to linearize the standard curve (i.e. logit of the mean absorbance (Y) is plotted against log of the known concentration (X) of Standards.

**The measured concentration of samples calculated from the standard curve must be multiplied by their respective dilution factor, because samples have been diluted prior to the assay, e.g. 1.55 ng/ml (from standard curve) x 10 (dilution factor) = 15.5 ng/ml.**

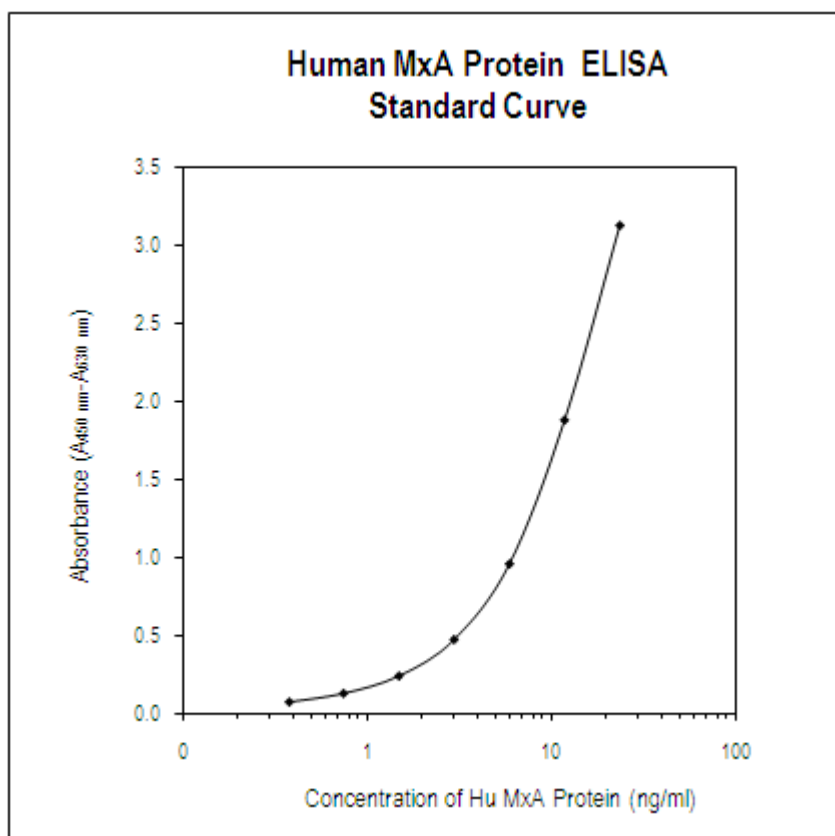


Figure 2: Typical Standard Curve for Human MxA Protein ELISA.

## 13. PERFORMANCE CHARACTERISTICS

» Typical analytical data of BioVendor Human MxA Protein ELISA are presented in this chapter.

### • Sensitivity

Limit of Detection (LOD) (defined as concentration of analyte giving absorbance higher than mean absorbance of blank\* plus three standard deviations of the absorbance of blank:  $A_{\text{blank}} + 3 \times \text{SD}_{\text{blank}}$ ) is calculated from the real human MxA in wells and is 0.03 ng/ml.

\*Dilution Buffer is pipetted into blank wells.

### • Limit of assay

Results exceeding resistin MxA protein of 24 ng/ml should be repeated with more diluted samples. Dilution factor needs to be taken into consideration in calculating the MxA protein concentration.

- **Precision**

Intra-assay (Within-Run, n=8)

<i>Sample</i>	<i>Mean (ng/ml)</i>	<i>SD (ng/ml)</i>	<i>CV (%)</i>
1	11.77	0.32	2.7
2	5.94	0.18	3.0

- **Linearity**

Serum samples were serially diluted with Dilution Buffer and assayed.

<i>Sample</i>	<i>Dilution</i>	<b><i>O</i></b> <i>bserved (ng/ml)</i>	<b><i>E</i></b> <i>xpected (ng/ml)</i>	<i>Recovery O/E (%)</i>
1	-	11.77	-	-
	2x	6.18	5.89	105.0
	4x	2.76	2.94	93.8
	8x	1.30	1.47	88.1
2	-	5.94	-	-
	2x	3.34	2.97	112.5
	4x	1.56	1.49	105.1
	8x	0.71	0.74	95.6

## 14. TROUBLESHOOTING AND FAQs

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### »» Weak signal in all wells

Possible explanations:

- Omission of a reagent or a step
- Improper preparation or storage of a reagent
- Assay performed before reagents were allowed to come to room temperature
- Improper wavelength when reading absorbance

### »» High signal and background in all wells

Possible explanations:

- Improper or inadequate washing
- Overdeveloping; incubation time with Substrate Solution should be decreased before addition of Stop Solution
- Incubation temperature over 30°C

### »» High coefficient of variation (CV)

Possible explanation:

- Improper or inadequate washing
- Improper mixing Standards, Quality Controls or samples







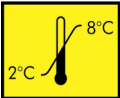

## 15. REFERENCES

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### »» References to MxA protein:

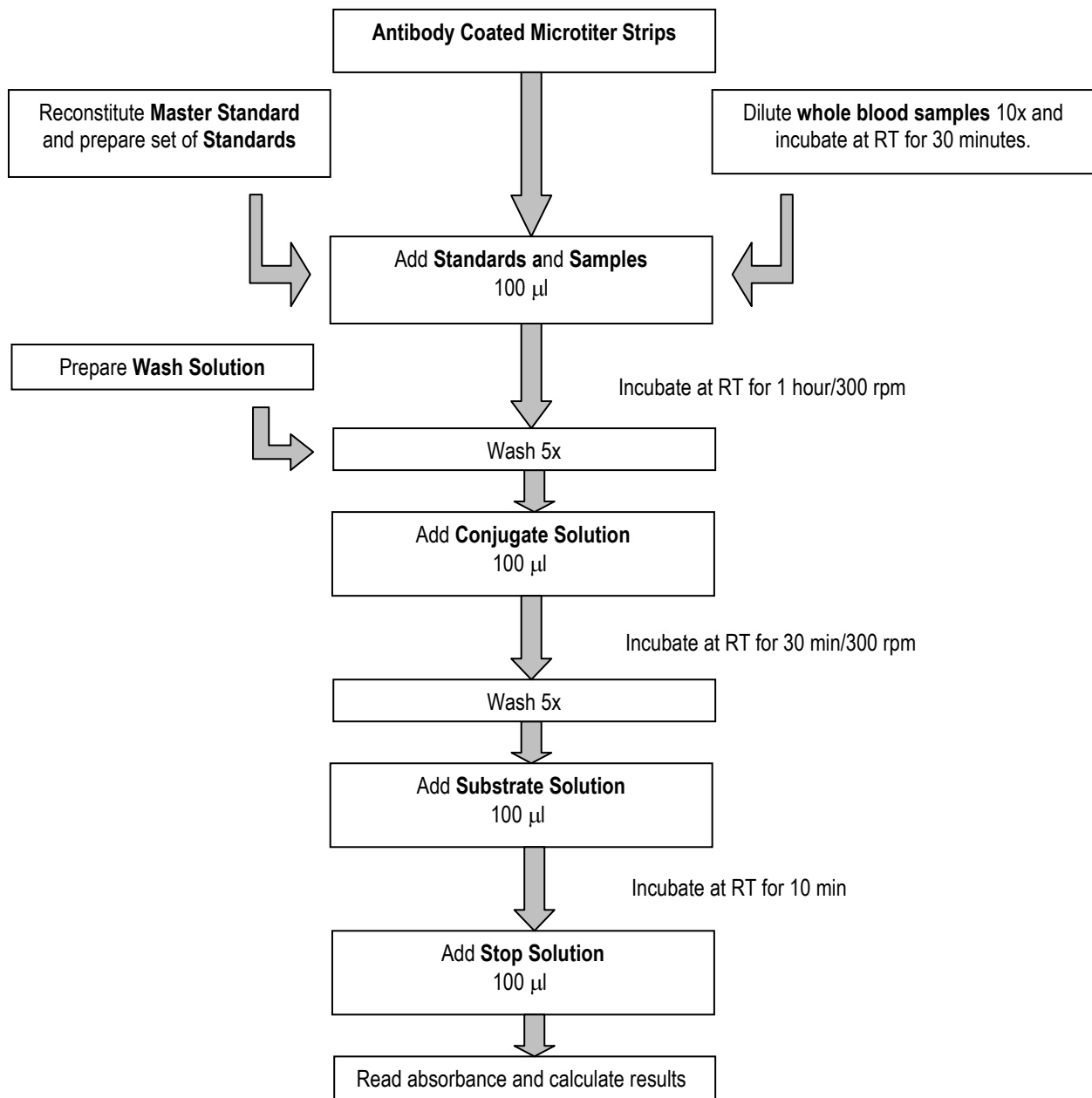
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## 16. EXPLANATION OF SYMBOLS

	Catalogue number
	Content
	Lot number
	See instructions for use
	Biological hazard
	Expiry date
	Storage conditions
	Identification of packaging materials

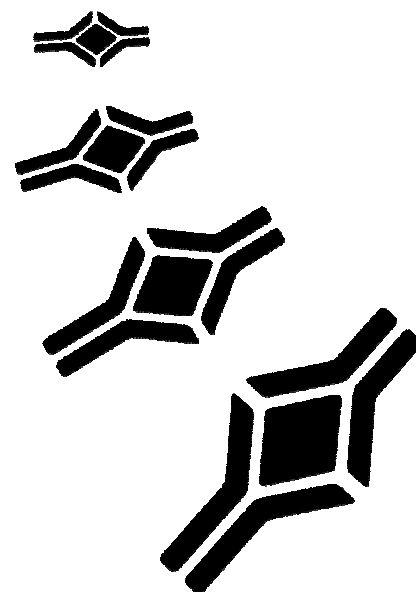


## Assay Procedure Summary




NOTES





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