

YK210 Mouse/Rat Urocortin 1 EIA

FOR LABORATORY USE ONLY

<Distributed by>

SCETI

Kasumigaseki Place, 3-6-7, Kasumigaseki, Chiyoda-ku
Tokyo 100-0013 Japan

URL: <http://www.sceti.co.jp/export/> e-mail: medical@sceti.co.jp

Contents

I . Introduction	2
II . Characteristics	3
III . Composition	4
IV . Method	5-6
V . Notes	7
VI . Performance Characteristics	8-11
VII . Stability and Storage	12
VIII . References	12

- Please read all the package insert carefully before beginning the assay -

YK210 Mouse/Rat Urocortin 1 EIA Kit

I. Introduction

Urocortin (Urocortin1: Ucn1) is first identified in rat ¹⁾, and later in human²⁾ and mouse³⁾. It is the second mammalian member of the CRF family. Rat and mouse Ucn1 have the same amino acid sequence and display 95% structure homology to human Ucn1, 45% to CRF and 63% to urotensin. In the rat, Ucn1 immunoreactivity (IR) was shown to distribute widely in central nervous system, endocrine organs, and digestive system and its concentration was highest in pituitary (11 pmol/g, w.w.) ⁴⁾. Koziez, Yanaihara et al. used a polyclonal antibody against rat Ucn1 to define the distribution of Ucn1-IR in rat central nervous system and found a large number of neurons with Ucn1-IR in rat brain ⁵⁾.

Synthetic human Ucn1 binds with high affinity to CRF receptor type 1(CRFR1), 2 alpha(CRFR2 α) and 2 beta (CRFR2 β). CRFR1 and CRFR2 have been shown to link to the development of stress-related disorders, such as mood and eating disorders. CRFR1 is expressed predominantly in the brain and pituitary, whereas CRFR2 expression is limited to particular brain areas and to some peripheral organs ⁶⁾. Data were also presented supporting the hypothesis that this peptide is the endogenous ligand for the CRFR2. ⁵⁾

Synthetic human Ucn1 stimulates cAMP accumulation in cells stably transfected with those receptors and acts in vitro to release ACTH from dispersed rat anterior pituitary cells. In addition, the CRF-binding protein binds human Ucn1 with high affinity and can prevent Ucn1-stimulated ACTH secretion in vitro²⁾. Ucn1 was suggested to play important roles in various tissues in normal rats, but shown not to behave as a hypothalamic hypophysiotropic factor in mediating adrenocorticotropin secretin in adrenalectomized rats ⁴⁾. Ucn1 has been implicated in various endocrine responses, such as blood pressure regulation, as well as in higher cognitive functions ⁵⁾.

Synthetic human Ucn1 also stimulates plasma ACTH, cortisol and atrial natriuretic peptide (ANP) secretin and suppresses plasma ghrelin in healthy male volunteers⁷⁾. In the human, plasma Ucn1 is elevated in heart failure, especially in its early stages. This fact may useful in the diagnosis of early heart failure ⁸⁾.

We have already developed mouse urocortin 2 (Ucn2) EIA kit (YK190), rat urocortin 2 (Ucn2) EIA kit (YK191) and mouse/rat urocortin 3 (Ucn3) EIA kit (YK200). This time, as a part of tools for urocortin research, our laboratory developed mouse/rat Ucn1 EIA kit (YK210), which is highly specific for mouse/rat Ucn1 with almost no crossreaction with Ucn2 (mouse), Ucn2 (rat), Ucn3 (mouse, rat), ACTH (mouse, rat), ACTH (human) and CRF (mouse, rat, human). The kit can be used for measurement of Ucn1 in mouse/rat plasma or serum with high sensitivity. It will be a specifically useful tool for Ucn1 research.

YK210 Mouse/Rat Urocortin 1 EIA Kit	Contents
<ul style="list-style-type: none"> ▼ The assay kit can measure mouse/rat urocortin 1 in mouse/rat plasma and serum within the range of 1.563-100 ng/mL. ▼ The assay is completed within 16-18 hr + 3 hr. ▼ With one assay kit, 40 samples can be measured in duplicate. ▼ Test sample: Mouse/rat plasma and serum Sample volume: 10 µL ▼ The 96-wells plate in kit is consisted by 8-wells strips, and the strips can be used separately. ▼ Precision and reproducibility Intra-assay CV (%): Rat serum 2.87-9.48 Rat plasma 1.70-13.01 Mouse serum 3.51-5.73 Mouse plasma 3.14-5.32 Inter-assay CV (%): Rat serum 4.44-7.76 Rat plasma 5.71-15.72 Mouse serum 5.45-9.83 Mouse plasma 8.70-10.12 ▼ Stability and storage Store all of the components at 2-8°C. The kit is stable under the condition for 24 months from the date of manufacturing. The expiry date is stated on the label of kit. 	<ol style="list-style-type: none"> 1) Antibody coated plate 2) Standard 3) Labeled antigen 4) SA-HRP solution 5) Enzyme substrate solution (TMB) 6) Stopping solution 7) Buffer solution 8) Washing solution (concentrated) 9) Adhesive foil

II. Characteristics

This EIA kit is used for quantitative determination of urocortin 1 in mouse/rat plasma and serum samples. The kit is characterized by its sensitive quantification and high specificity. In addition, it has no influence by other components in samples. Mouse/rat urocortin 1 standard is highly purified synthetic product.

< Specificity >

This EIA kit has high specificity to mouse/rat urocortin 1 and shows no crossreactivity to urocortin 2 (mouse), urocortin 2 (rat), urocortin 3 (mouse, rat), ACTH (mouse, rat), ACTH (human), and CRF (mouse, rat, human).

< Assay principle >

This EIA kit for determination of mouse/rat urocortin 1 in samples is based on a competitive enzyme immunoassay using combination of highly specific antibody to mouse/rat urocortin 1 and biotin-avidin affinity system. To the 96-wells of plate coated with rabbit anti-mouse/rat urocortin 1 antibody, standards or samples and labeled antigen (biotinylated antigen) are added for competitive immunoreaction. After incubation and plate washing, horseradish peroxidase (HRP) labeled streptoavidin (SA) is added to form HRP labeled SA-labeled antigen-antibody complex on the surface of the wells. Finally, HRP enzyme activity is determined by 3,3',5,5'-Tetramethyl benzidine (TMB) and the concentration of mouse/rat Ucn1 is calculated.

III. Composition

Component	Form	Quantity	Main ingredient
1. Antibody coated plate	microtiter plate	1 plate (96 wells)	Rabbit anti mouse /rat urocortin1 antibody coated
2. Standard	lyophilized	1 vial (100 ng)	Synthetic mouse/rat urocortin1
3. Labeled antigen	lyophilized	1 vial	Biotinylated mouse/rat urocortin1
4. SA-HRP solution	liquid	1 bottle (12 mL)	Horseradish peroxidase labeled streptoavidin
5. Enzyme substrate solution	liquid	1 bottle (12 mL)	3,3',5,5'-Tetramethylbenzidine (TMB)
6. Stopping solution	liquid	1 bottle (12 mL)	1M H ₂ SO ₄
7. Buffer solution	liquid	1 bottle (15mL)	Tris-HCl buffer
8. Washing solution (concentrated)	liquid	1 bottle (50 mL)	Concentrated saline
9. Adhesive foil		3 pieces	

IV. Method

< Equipments required >

1. Photometer for microtiter plate (plate reader) which can read extinction 2.5 at 450 nm
2. Microtiter plate shaker
3. Washing device for microtiter plate and dispenser with aspiration system
4. Micropipettes, multi-channel pipettes for 8 wells or 12 wells and their tips
5. Glass test tubes for preparation of standard solution
6. Graduated cylinder (1,000 mL)
7. Distilled or deionized water

< Preparatory work >

1. Preparation of standard solution:
Reconstitute the mouse/rat urocortin 1 standard with 1 mL of buffer solution, which affords 100 ng/mL standard solution. The reconstituted standard solution (0.1 mL) is diluted with 0.1 mL of buffer solution that yields 50 ng/mL standard solution. Repeat the dilution procedure to make each standard solution of 25, 12.5, 6.25, 3.125 and 1.563 ng/mL. Buffer solution itself is used as 0 ng/mL standard solution.
2. Preparation of labeled antigen solution:
Reconstitute labeled antigen with 6 mL of distilled or deionized water.
3. Preparation of washing solution:
Dilute 50 mL of washing solution (concentrated) to 1,000 mL with distilled or deionized water.
4. Other reagents are ready for use.

< Procedure >

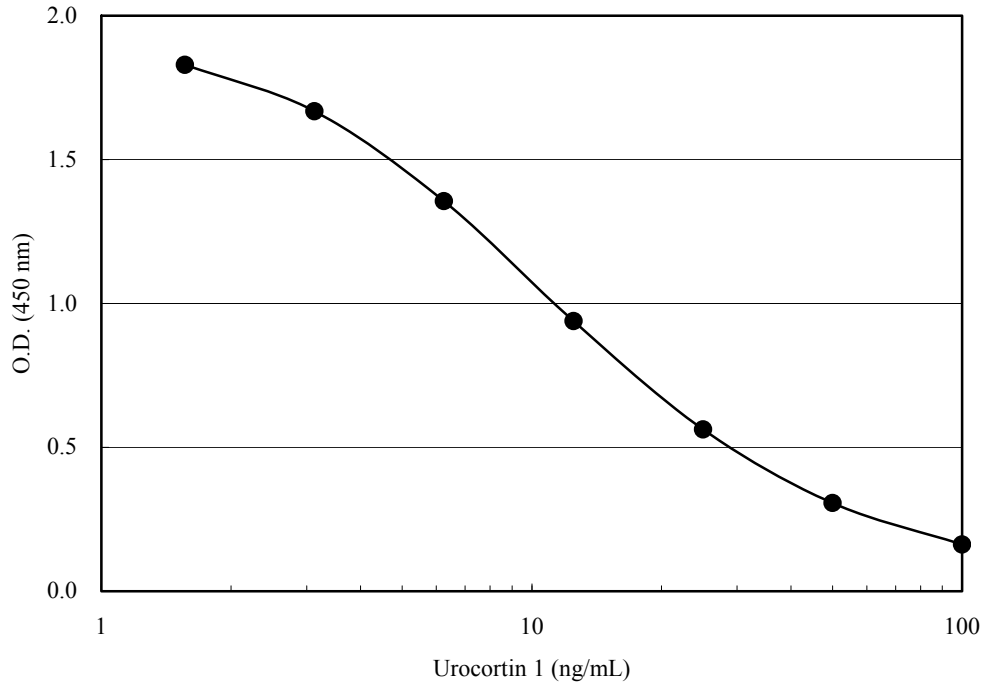
1. Before starting the assay, bring all the reagents and samples to room temperature (20~30°C).
2. Fill 0.3 mL/well of washing solution into the wells and aspirate the washing solution in the wells. Repeat this washing procedure further twice (total 3 times). Finally, invert the plate and tap it onto an absorbent surface, such as paper toweling, to ensure blotting free of most residual washing solution.
3. Add 40 μ L of buffer solution to the wells first, then introduce 10 μ L of each of standard solutions (0, 1.563, 3.125, 6.25, 12.5, 25, 50 and 100 ng/mL) or samples and finally add 50 μ L of labeled antigen to the wells. The total pipetting time of standard solutions and samples for a whole plate should not exceed 30 minutes.
4. Cover the plate with adhesive foil and incubate it at 4°C for 16~18 hours (keep still, plate shaker not need).
5. After incubation, move the plate back to room temperature keeping for approximately 40 minutes (keep still, plate shaker not need) and take off the adhesive foil, aspirate and wash the wells 4 times with 0.3 mL/well of washing solution. Finally, invert the plate and tap it onto an absorbent surface, such as paper toweling, to ensure blotting free of most residual washing solution.
6. Add 100 μ L of SA-HRP solution to each of the wells.
7. Cover the plate with adhesive foil and incubate it at room temperature for 2 hours. During the incubation, the plate should be shaken with a plate shaker (approximately 100 rpm).
8. Take off the adhesive foil, aspirate and wash the wells 4 times with 0.3 mL/well of washing solution. Finally, invert the plate and tap it onto an absorbent surface, such as paper toweling, to ensure blotting free of most residual washing solution.
9. Add 100 μ L of Enzyme substrate solution (TMB) to each of the wells, cover the plate with adhesive foil and keep it for 30 minutes at room temperature in a dark place for color reaction (keep still, plate shaker not need).
10. Add 100 μ L of stopping solution to each of the wells to stop color reaction.
11. Read the optical absorbance of the solution in the wells at 450 nm. The dose-response curve of this assay fits best to a 4 (or 5)-parameter logistic equation. The results of unknown samples can be calculated with any computer program having a 4 (or 5)-parameter logistic function. Otherwise calculate mean absorbance values of wells containing standards and plot a standard curve on semi logarithmic graph paper (abscissa: concentration of standard; ordinate: absorbance values). Use the average absorbance of each sample to determine the corresponding value by simple interpolation from this standard curve.

V. Notes

1. EDTA-2Na (1 mg/mL) additive blood collection tube is recommended for the plasma collection. Serum and plasma samples must be used as soon as possible after collection. If the samples are tested later, they should be divided into test tubes in small amount and frozen at or below -30°C. Avoid repeated freezing and thawing of samples.
2. Standard and labeled antigen solutions should be prepared immediately before use. This kit can be used dividedly in strips of the plate. In such case, the rest of reconstituted reagents (standard and labeled antigen) should be stored at or below -30°C (stable for 1 month).
3. The total pipetting time of standard solutions and samples for a whole plate should not exceed 30 minutes.
4. During storage of washing solution (concentrated) at 2~8°C, precipitates may be observed, however they will be dissolved when diluted. Diluted washing solution is stable for 6 months at 2~8°C.
5. Pipetting operations may affect the precision of the assay, so that pipette standard solutions or samples precisely into each well of plate. In addition, use clean test tubes or vessels in assay and use new tip for each standard or sample to avoid cross contamination.
6. When sample concentration exceeds 100 ng/mL, it needs to be diluted with buffer solution to proper concentration.
7. During the incubation with SA-HRP solution at room temperature, the assay plate should be shaken gently by a plate shaker to promote immunoreaction (approximately 100 rpm).
8. Perform all the determination in duplicate.
9. Read plate optical absorbance of reaction solution in wells as soon as possible after stop color reaction.
10. To quantitate accurately, always run a standard curve when testing samples.
11. Protect reagents from strong light (e.g. direct sunlight) during storage and assay.
12. Satisfactory performance of the test is guaranteed only when reagents are used from combination pack with identical lot number.
13. Some reagents contain human serum (tested and found negative for HBsAG, HIV 1/2, HCV, HIV-1 AG or HIV-1 NAT, ALT and a test for Syphilis by FDA approved methods), care should be taken when handling.

VI. Performance Characteristics

Typical standard curve



<Analytical recovery>

<Rat serum A>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	4.34		
2.0	6.62	6.34	104.42
7.0	12.64	11.34	111.46
20.0	28.45	24.34	116.89

<Rat serum B>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	2.61		
2.0	4.24	4.61	91.97
7.0	7.88	9.61	82.00
20.0	22.51	22.61	99.56

<Rat serum C>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	2.92		
2.0	4.61	4.92	93.70
7.0	7.99	9.92	80.54
20.0	22.29	22.92	97.25

<Rat plasma A>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	3.03		
2.0	5.52	5.03	109.74
7.0	9.55	10.03	95.21
20.0	20.46	23.03	88.84

<Rat plasma B>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	3.03		
2.0	5.11	5.03	101.59
7.0	9.29	10.03	92.62
20.0	19.43	23.03	84.37

<Rat plasma C>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	2.55		
2.0	4.55	4.55	100.00
7.0	7.87	9.55	82.41
20.0	20.01	22.55	88.74

<Mouse serum A>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	4.79		
2.0	6.54	6.79	96.32
7.0	11.00	11.79	93.30
20.0	24.79	24.79	100.44

<Mouse serum B>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	4.25		
2.0	6.10	6.25	97.60
7.0	10.68	11.25	94.93
20.0	25.01	24.25	103.13

<Mouse serum C>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	4.07		
2.0	5.99	6.07	98.68
7.0	10.29	11.07	92.95
20.0	27.01	24.07	112.21

<Mouse plasma A>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	5.09		
2.0	7.22	7.09	101.83
7.0	13.06	12.09	108.02
20.0	29.60	25.09	117.98

<Mouse plasma B>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	4.41		
2.0	6.39	6.41	99.69
7.0	11.27	11.41	98.77
20.0	28.22	24.41	115.61

<Mouse plasma C>

Added Urocortin 1 (ng/ml)	Observed (ng/ml)	Expected (ng/ml)	Recovery (%)
0.0	4.24		
2.0	6.88	6.24	110.26
7.0	11.90	11.24	105.87
20.0	28.20	24.24	116.34

<Dilution test>**<Rat serum>**

Dilution ratio	Rat A (ng/mL)	Rat B (ng/mL)
x 1	4.77	3.15
x 1.5	3.50	2.87
x 2	2.34	1.84
x 3	1.83	0.83

<Rat plasma>

Dilution ratio	Rat A (ng/mL)	Rat B (ng/mL)
x 1	2.63	3.41
x 1.5	2.08	2.47
x 2	1.37	1.60
x 3	0.99	1.07

<Mouse serum>

Dilution ratio	Mouse A (ng/mL)	Mouse B (ng/mL)
x 1	5.15	4.31
x 1.5	3.86	3.86
x 2	3.65	2.83
x 3	2.25	1.50

<Mouse plasma>

Dilution ratio	Mouse A (ng/mL)	Mouse B (ng/mL)
x 1	5.65	5.10
x 1.5	4.20	3.47
x 2	3.22	2.98
x 3	2.66	2.17

<Crossreactivity>

Related peptides	Crossreactivity (%)
Urocortin 1 (mouse, rat)	100.0
Urocortin 1 (human)	51.3
Urocortin 2 (mouse)	0
Urocortin 2 (rat)	0
Urocortin 3 (mouse, rat)	0
ACTH (mouse, rat)	0
ACTH (human)	0
CRF (mouse, rat, human)	0

<Precision and reproducibility>

Test sample	Intra-assay CV(%)	Inter-assay CV (%)
Rat serum	2.87-9.48	4.44-7.76
Rat plasma	1.70-13.01	5.71-15.72
Mouse serum	3.51-5.73	5.45-9.83
Mouse plasma	3.14-5.32	8.70-10.12

<Assay range>

1.563 ~ 100 ng/mL

VII. Stability and Storage

- < Storage > Store all of the components at 2~8°C.
- < Shelf life > The kit is stable under the condition for 24 months from the date of manufacturing. The expiry date is stated on the label of kit..
- < Package > For 96 tests per one kit including standards

VIII. References

1. Vaughan J, Donaldson C et al (1995): Urocortin, a mammalian neuropeptide related to fish urotensin I and to corticotropin-releasing factor. *Nature* **378**, 287-292.
2. Donaldson CJ, Sutton SW et al (1996): Cloning and characterization of human urocortin. *Endocrinology* **137**, 2167-2170.
3. Zhao L, Donaldson CJ et al (1998): The structures of the mouse and human urocortin genes (Ucn and UCN). *Genomics* **50**, 23-33
4. Oki Y, Iwabuchi M et al (1998): Distribution and concentration of urocortin, and effect of adrenalectomy on its contents in rat hypothalamus. *Life Sci* **62**, 807-812
5. Koziez F, Yanaihara H et al (1998): Distribution of urocortin-like immunoreactivity in the central nervous system of the rat. *Comp Neurol* **391**, 1-10
6. Dautzenberg FM, Kilpatrick GJ et al (2001): Molecular biology of the CRH receptors in the mood. *Peptides* **22**, 753-760
7. Davis ME, Pemberton CJ et al (2004): Urocortin-1 infusion in normal humans. *J Clin Endocrinol Metab* **89**, 1402-0409
8. Ng Li, Loke IW et al (2004): Plasma urocortin in human systolic heart failure. *Clin Sci (Lond)* **106**, 383-388

<Manufacturer>

Yanaihara Institute Inc.
2480-1 Awakura, Fujinomiya-shi
Shizuoka, Japan 418-0011
TEL: +81-544-22-2771 FAX: +81-544-22-2770
Website: <http://www.yanaihara.co.jp> E-mail: ask@yanaihara.co.jp

Update at Apr. 13, 2012